

Micro Biotec'13

PORTUGUESE CONGRESS OF
MICROBIOLOGY AND BIOTECHNOLOGY

6th - 8th December | Aveiro Portugal

Abstracts Book



Industrial Microbiology & Biotechnology

P079

DISTINCT BEHAVIOR BETWEEN MULTI-ANTIBIOTIC RESISTANT *ESCHERICHIA COLI* STRAINS TOWARDS REACTIVE OXYGEN SPECIES

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Antibiotic resistant bacteria have been implicated in a large number of nosocomial infections. This worldwide problem drew the attention to the development of new disinfection techniques, such as photoinactivation. During photocatalysis, the generation of reactive oxygen species (ROS) such as hydrogen peroxide (H₂O₂), hydroxyl radicals (•OH) and superoxide radical (O₂^{•-}) are supposed to occur. Hence, photocatalysis has been used to inactivate organisms, through ROS attack. However, several studies reported that in *Escherichia coli* an increased tolerance to oxidative stress induced by ROS may occur, and that it may be related to antibiotic resistance. In this study, the susceptibility to photoinactivation of two antibiotic resistant *E. coli* environmental strains (A5EL5 and E5EL20) belonging to the same phylogenetic group and with similar antibiotic resistance phenotype was assessed.

After 40 min of UV/TiO₂ exposure, viability losses of 44.2 % and above 99.0 % were recorded for strains A5EL5 and E5EL20, respectively. Based on the hypothesis that these two strains have distinct tolerance to different ROS generators, the ROS formed after contact with H₂O₂, KO₂ or H₂O₂ + Fe²⁺ (Fenton reaction) were quantified using a fluorescence method. The probe 2', 7'-dichlorohydrofluorescein diacetate (DCFH-DA) is widely used to measure the ROS content in cells. After hydrolysis by cellular esterases, DCFH is subsequently oxidized by ROS to highly fluorescent DCF. After 60 min of contact with DCFH-DA, cells were incubated in the presence of H₂O₂ or KO₂ for O₂^{•-} and H₂O₂ + Fe²⁺ (Fenton reaction) for •OH. Fluorescence units were measured in the crude cell extracts and converted into ROS concentration and the data was normalized by total protein content. Significant differences (p<0.05) between ROS content in A5EL5 and E5EL20 crude cell extracts were found for H₂O₂ (1.4x10⁻⁵ ± 2.9x10⁻⁶ and 1.1x10⁻⁴ ± 1.1x10⁻⁶ μmol ROS.μg⁻¹ total protein, respectively) and Fenton reaction (9.6x10⁻⁵ ± 5.6x10⁻⁶ and 1.5x10⁻⁴ ± 5.2x10⁻⁶ μmol ROS.μg⁻¹ total protein, respectively). No significant difference was found for KO₂. These preliminary tests, suggest that, in fact, the different tolerance to photocatalysis of the environmental *E. coli* may be related to oxidative stress response, but not to antibiotic resistance.

Acknowledgements: This work was partially financially supported by FCT (project PTDC/EQU EQU/115614/2009).