



Candida albicans: the current status regarding vaginal infections

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Abstract

Vaginal infections caused by *Candida albicans* are a significant global health concern due to their recurrence and negative impact on quality of life. This review examines the pathogenesis of *C. albicans* infections, emphasizing critical virulence factors such as biofilm formation, adherence, and phenotypic switching. Risk factors include immune system suppression, antibiotic use, and hormonal changes, all of which can lead to fungal overgrowth and infection. Current prevention and/or treatment strategies primarily rely on antifungal therapies, personal hygiene practices, and probiotics. However, challenges like antifungal resistance, recurrence, and limited treatment efficacy highlight the need for innovative approaches. Therefore, emerging methods such as novel antifungal agents, vaccines, and nanotechnology-based delivery systems offer promising advancements to improve infection control. Additionally, the immune system plays a key role in preventing *C. albicans* infections, with both innate and adaptive immunity acting to restrict fungal colonization and growth. Commercially available products, such as antifungal creams, vaginal probiotics, and hygiene solutions, are practical options but often lack long-term efficacy. Persistent challenges, including resistance, patient noncompliance, and restricted access to emerging therapies, hinder comprehensive prevention and treatment efforts. Thus, future research should focus on promoting interdisciplinary approaches, integrating personalized medicine, and enhancing healthcare accessibility. This review intends to present the current state of the art within the abovementioned issues and to enhance the understanding of the multifactorial nature of *C. albicans* infections and advanced prevention strategies, which are essential to reduce the burden of vaginal candidiasis worldwide and improve patient quality of life outcomes.

Key points

- *Candida albicans* pathogenesis involves biofilms, adherence, and phenotypic switching.
- Vaccines, nanotechnology, and new drugs offer improved prevention and treatment.
- Addressing antifungal resistance and patient compliance is key for prevention success.

Keywords *Candida albicans* · Vaginal infections · Pathogenesis · Prevention strategies · Antifungal resistance · Review

Introduction

Candida albicans is a diploid polymorphic fungus commonly present in several human surfaces such as skin, throat, or vagina mucosa (Parambath et al. 2024). Under certain conditions, such as a weakened immune system, diabetes, pregnancy, or antibiotics therapy, a dysbiosis occurs, and the situation evolves into an infection (Rosati et al. 2020). In the vaginal environment, *Candida* spp. infection is also known as vaginal candidiasis or vulvovaginal candidiasis (VVC). VVC is the second most common vaginal infection (after bacterial vaginosis) and affects 75% of women at least once in their lifetime, although up to 9% of them face recurrent infection episodes, which may surpass 4 per year (recurrent vulvovaginal candidiasis—RVVC) (Rosati et al. 2020).

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Besides *C. albicans* (the most common species causing invasive diseases), other common species of *Candida* spp. that can overgrow are *Candida glabrata*, *Candida parapsilosis*, *Candida tropicalis*, and *Candida krusei* (Rosati et al. 2020; CDC 2024a).

Although prevention can be achieved through simple strategies such as using cotton underwear and maintaining good hygiene habits, morbidity rates of VVC or RVVC are increasing, as well as the medical costs associated. In fact, *C. albicans* is responsible for more than 150 million mucosal infections per year, leading to healthcare costs of ca. \$2 billion in the USA (Richardson 2022). Furthermore, symptoms experienced by women when an infection occurs (vaginal itching, burning, redness, and a white discharge) are often underestimated but limiting to their quality of life (Mayo Clinic 2024). Additionally, despite the pressing need for reliable diagnostic tests and new, safe, and effective treatments and vaccines, research into the mechanisms of fungal vaginal infections is not as advanced as that for other types of diseases. Thus, a systematic overview of the knowledge gathered so far is crucial to understand the current point of situation in terms of available treatments, as well as the new avenues of research that are being drawn for the future. The current review covers the abovementioned issues, focusing on vaginal infections by *Candida albicans*.

Risk factors for vaginal infections

C. albicans infections are influenced by a variety of factors that alter the vaginal microbiota's normal equilibrium, promoting fungal overgrowth. Recent study continues to provide light on the following critical factors.

- a) Vaginal microbiome: this dynamic microecosystem plays a crucial role in maintaining vaginal health and preventing infections such as VVC. In healthy women, the vaginal microbiota is typically dominated by *Lactobacillus* sp. which help maintain a low pH environment and produce antimicrobial compounds. However, vaginal dysbiosis, characterized by a decrease in *Lactobacillus* sp. and an increase in diverse microorganisms, can contribute to VVC susceptibility (Ceccarani et al. 2019; Zeise et al. 2021). Studies have shown that women with VVC exhibit significant alterations in their vaginal microbiome compared to healthy individuals. These changes include a reduction in Firmicutes (primarily *Lactobacillus* sp.) and an increase in Actinobacteria, Bacteroidetes, and Proteobacteria (Ceccarani et al. 2019). The dysbiotic state creates a favorable environment for *Candida* sp. overgrowth, particularly *C. albicans*, which is the primary causative agent of VVC (Ceccarani et al. 2019; Valeriano et al. 2024). Furthermore,
- b) Antibiotic use: antibiotics with a broad spectrum remain a major risk factor for vaginal *C. albicans* infections. Antibiotics can reduce protective vaginal flora, particularly *Lactobacillus* species, which help by producing antimicrobials such as lactic acid. This microbial imbalance enables *C. albicans* to overgrow and invade the vaginal mucosa. *Lactobacillus* sp. depletion lowers competitive exclusion and fungal colonization resistance (Pirotta and Garland 2005; Achkar and Fries 2010; Chee et al. 2020). Antibiotic-induced dysbiosis can increase the recurrence of infections and treatment resistance.
- c) Hormonal changes: hormonal oscillations, particularly estrogen, affect the vaginal environment. Elevated estrogen levels, which occur during pregnancy, hormonal contraception use, or hormone replacement treatment, stimulate glycogen deposition in the vaginal epithelium, providing a perfect substrate for *C. albicans* development. Estrogen also influences the immunological response, reducing local immune defenses in the vaginal mucosa and generating an environment conducive to fungus persistence (Sobel 2007). This implies that hormonal regulatory mechanisms play an important role in determining susceptibility to infections.
- d) Immunocompromised states: immunosuppression caused by illnesses such as human immunodeficiency virus (HIV) infection, diabetes mellitus, or the use of immunosuppressive medications dramatically increases the risk of vaginal infections. A compromised immune system reduces the body's ability to mount effective defenses against fungal infection (Patil et al. 2018). Patients undergoing chemotherapy or those suffering from chronic illnesses frequently encounter recurring vaginal infections because of weakened cell-mediated immunity. Recent research has focused on the involvement of neutrophils and T-cells in vaginal homeostasis and how their failure leads to increased fungal colonization (Köhler et al. 2017). Diabetes and unregulated blood sugar levels also increase the likelihood of recurrent vulvovaginal candidiasis in individuals because increased glucose levels offer a plentiful energy supply to *C. albicans*, boosting its growth and biofilm formation capabilities. Additionally, high blood sugar inhibits the

body's natural defense system, decreasing neutrophils' effectiveness in fighting off fungal infections (Goderidze et al. 2022).

- e) Lifestyle factors and hygiene practices: Certain lifestyle behaviors and hygiene habits, such as wearing tight and non-breathable clothing, not practicing good hygiene, and excessive douching, can disrupt the natural balance of vaginal flora, making individuals more prone to *C. albicans* infections. Recent research emphasizes the importance of educational programs on correct hygiene practices to lower infection risks, particularly in women prone to recurring infections (Ventolini and Baggish 2006).

Pathogenesis of *Candida albicans*

The virulence of a microbial species arises from the interaction between the pathogen and the host rather than being an inherent trait of the microorganism itself. Unlike primary pathogens, which can cause disease without needing a compromised host, opportunistic or facultative pathogens, like *Candida* spp., typically cause infections only in hosts with predisposing vulnerabilities.

Environmental factors play a crucial role in shaping how pathogens evade or overcome the host's natural defenses. Virulence is not a static property; it can vary, increasing, decreasing, or potentially returning based on the conditions present (Méthot and Alizon 2014). *C. albicans* has the ability to transform from commensal to pathogenic due to its ability to adhesion, biofilm growth, hydrolytic enzyme release, morphological change, and metabolic adaptability

(Fig. 1, Mayer et al. (2013)). In addition, it rapidly adapts to the host environment and infects people with predisposing factors such as antibiotic treatment, malignancy, or weakened immune systems (Tsui et al. 2016; Ciurea et al. 2020).

C. albicans is a polymorphic fungus capable of existing as a yeast, pseudo hyphae, or true hyphae. This dimorphism is central to its pathogenicity. In its yeast form, *C. albicans* is adapted for commensal growth on mucosal surfaces. Upon encountering favorable conditions, such as tissue damage or changes in the local environment, *C. albicans* can transition to a hyphal form, which is more invasive. Hyphae penetrate host tissues, facilitating tissue invasion and destruction (Mayer et al. 2013; Chow et al. 2021).

Mechanisms of infection

There has been a fascinating and extensive range of research into the modes of infection of *C. albicans*. These can be considered as a series of stages, as the organism's fixation onto the host's tissues is followed by an evasion from its immune response. In the yeast phase, *C. albicans* resides in the gastrointestinal mucosa, while in the filamentous stage, it penetrates deep into the host tissues to bypass the immune reaction. This transition from yeast to filamentous and vice versa is influenced by favorable environmental conditions, such as tissue damage or changes in temperature, pH, and oxygen levels (Nielsen et al. 2021). The ability of *C. albicans* to stick on epithelial cells is further enhanced by the agglutinin-like sequence (ALS) proteins, which play a fundamental role in the process of infection establishment (Cangui-Panchi et al. 2023).

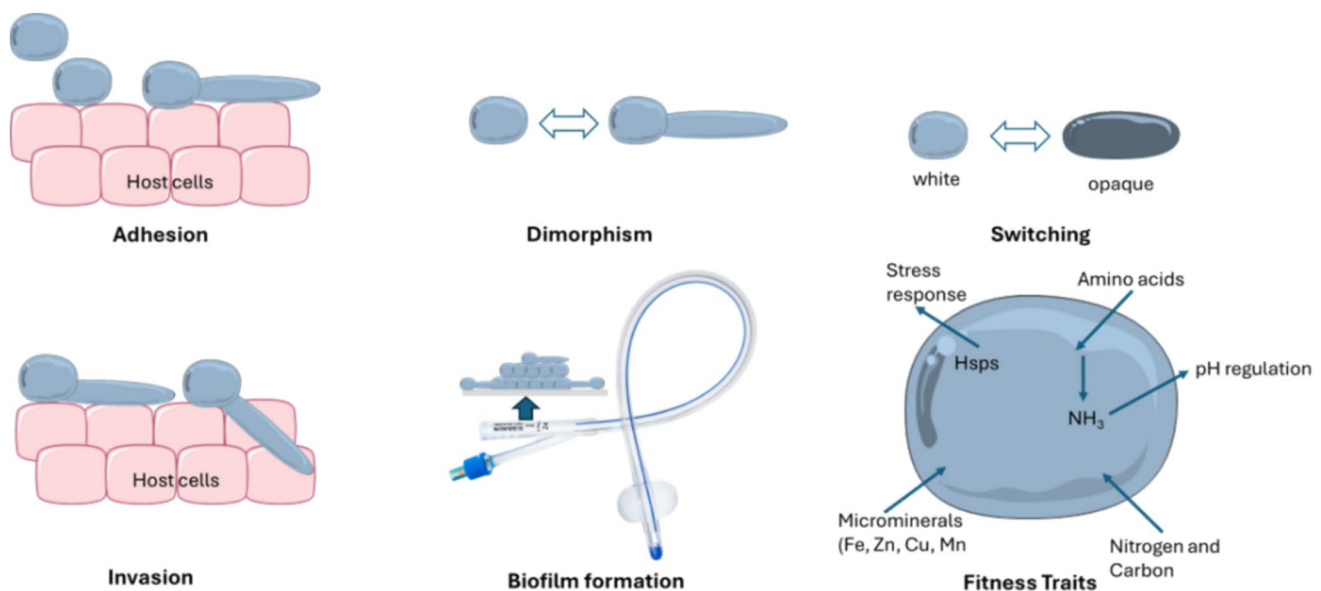


Fig. 1 A summary of key pathogenic mechanisms utilized by *Candida albicans*. Adapted from Mayer et al. (2013). The figure was partly generated using Servier Medical Art, provided by Servier, licensed under a Creative Commons Attribution 3.0 unported license

C. albicans initiates infection through a well-orchestrated sequence of events. The first step is adhesion to host tissues, primarily through the activity of ALS proteins. ALS proteins were identified as the first group of factors that mediate adhesion of the fungus to its host cells, by binding to the host cell surface and facilitating colonization of the mucosa. The *C. albicans* fungus contains many ALS proteins, with structural diversities which allow for successful colonization and adaptation to different environmental niches of the host or an indwelling device (Mayer et al. 2013; Lopes and Lionakis 2022).

Once adhered, *C. albicans* invades the host tissues by means of two principal mechanisms: induced endocytosis and active penetration of the host tissues. In induced endocytosis, the fungus is taken up by epithelial cells using a mechanism akin to bacterial intrusion whereby invasins bind to cellular targets and compel their internalization (Wächtler et al. 2012; Sheppard and Filler 2015; Maza et al. 2017; Lachat et al. 2022). In active penetration, *C. albicans* hyphal cells injure epithelial through the secretion of hydrolytic enzymes (mainly aspartyl proteases), which assist in the dissolution of extracellular matrix allowing the invasion into deeper tissues (Wächtler et al. 2012; Mayer et al. 2013).

A critical factor in the persistence of *C. albicans* infections is its capability to develop biofilms. Biofilms suppress fungal exposure to antifungal interventions and immune mechanisms by providing a coating around the fungus. Cells inside biofilms have different patterns of gene expression and metabolite production than their planktonic counterparts, heightening their tolerance to environmental challenges. This ability to develop biofilms is also related with chronic and recurrent infections, as it enables the organism to persist even despite treatment (Nobile and Johnson 2015; Malinová et al. 2023).

The virulence of *C. albicans* is also potentiated by its ability to produce many hydrolytic enzymes, such as secreted aspartyl proteinases (SAPs) and phospholipases, that are very important in tissue penetration and immune escape. SAPs breakdown host proteins and biofilm structures, assisting the fungus in deeper invasion of the vaginal epithelium (Galocha et al. 2019; Bras et al. 2024). Phospholipases break down the membranes of host cells, hence encouraging them to burst and allowing the fungus to invaginate and spread into the surrounding tissues (Mayer et al. 2013; D'Enfert et al. 2021; Lopes and Lionakis 2022).

Finally, candidalysin is an important virulence factor. Candidalysin is a peptide toxin produced by *C. albicans*, specially by its pathogenic hyphal form and functions by causing direct epithelial damage and provoking inflammatory signaling (Ho et al. 2020). Ho et al. (2019) found that candidalysin activates the epidermal growth factor receptor (EGFR), triggering mitogen-activated protein kinase

(MAPK) signaling. In addition, candidalysin stimulates the release of alarmins and antimicrobial peptides, further underpinning host immune responses (Ho et al. 2020).

These adhesion, invasion, biofilm formation, and enzyme and toxin production processes show how complex *C. albicans* mechanisms are in acquiring lesions, maintaining them and re-infecting the host.

Host–pathogen interactions

The interaction between *C. albicans* and the immune system of the host is a complex and dynamic system that encompasses innate and adaptive immune components. The innate immune system is predominantly effective in the initial stages of recognition and control of *C. albicans* colonization and infection, through the actions of neutrophils, macrophages, and dendritic cells (DC). These immune cells express a few pattern recognition receptors (PRRs) such as Toll-like receptors (TLRs), C-type lectin receptors (CLRs), and nucleotide-binding oligomerization domain (NOD)-like receptors (NLRs) that recognize and respond to the fungal cell wall and promote intracellular processes such as ingestion of the fungi and secretion of inflammatory cytokines to curb the fungal propagation (Dühring et al. 2015; Zheng et al. 2015; Lionakis et al. 2023). However, *C. albicans* has evolved various immune evasion strategies that allow it to persist within the host, often leading to chronic or recurrent infections. One such mechanism involves masking key elements of its cell wall, particularly β -glucan, during hyphal growth. β -Glucan is a potent activator of immune responses via PRRs like Dectin-1, but by camouflaging it, *C. albicans* reduces its visibility to immune cells, thereby preventing robust immune activation (Mata-Martínez et al. 2022). In addition, the biofilm formed by *C. albicans* not only enhances its survival in hostile environments but also provides a physical shield that impedes immune cell activity, further complicating the elimination of the pathogen (Hernández-Chávez et al. 2017; Garcia-Rubio et al. 2019).

In addition to innate immunity, the adaptive immune system, particularly Th17 cells (subsets of T cells), plays a crucial role in controlling *C. albicans* infections. Th17 cells produce IL-17, a cytokine that recruits neutrophils and enhances fungal clearance. However, *C. albicans* can reduce the Th17 response in chronic infections, promoting persistent infections by reducing immune control (Garcia-Rubio et al. 2019).

Role of the immune system

Innate immune responses

The innate immune response acts as the first line of defense against *C. albicans* in the vaginal mucosa. In the context

of vaginal infections, four key components play dominant roles: epithelial cells, PRRs, cytokines, and the NLRP3 inflammasome.

- a) Epithelial cells: vaginal epithelial cells (VECs) can distinguish between the commensal and pathogenic forms of *C. albicans*, responding differently based on fungal morphotype and load (Gaziano et al. 2023). Although VECs exhibit some antifungal activity, their effectiveness is lower compared to oral epithelial cells, for example (Barousse et al. 2005). Another factor influencing the protective efficacy of VECs is the vaginal environment, particularly pH levels. More acidic environments have been shown to reduce the antifungal activity of VECs (Barousse et al. 2005). On the other hand, certain interventions can enhance the efficacy of VECs against *C. albicans*. For instance, specific probiotic strains have been shown to strengthen the barrier function of VECs and improve their ability to distinguish between commensal and pathogenic forms of *C. albicans* (Dong et al. 2023). Additionally, topical estrogen application can promote VEC proliferation and maturation, potentially reinforcing the vaginal epithelial barrier against *C. albicans* (Sibeko et al. 2024).
- b) Pattern recognition receptors (PRRs): innate immune cells recognize *C. albicans* through various PRRs, including CLRs like Dectin- 1 and Dectin- 2 (Richardson and Moyes 2015). While these PRRs are critical for detecting *C. albicans*, they also recognize other fungi and microorganisms, leading to broader immune responses. This broad recognition capability allows these receptors to play important roles in defending against multiple fungal pathogens, though it means their activation is not specific to *C. albicans* alone (Robinson et al. 2009; Saijo and Iwakura 2011). Furthermore, *C. albicans* has evolved sophisticated strategies to evade or modulate PRR recognition, potentially limiting the effectiveness of the innate immune response. One such example involves shielding of β - 1,3-glucan, a key pathogen-associated molecular pattern (PAMP), with an outer mannan layer. This “hiding” strategy reduces the reactivity of Dectin- 1, against *C. albicans* yeast cells (Hernández-Chávez et al. 2017).
- c) NLRP3 inflammasome: the NLR family pyrin domain-containing 3 (NLRP3) is a multiprotein complex comprising an amino-terminal pyrin domain (PYD), a carboxy-terminal leucine-rich repeat domain (LRR domain), and a central NACHT domain thus named to its presence in the neuronal apoptosis inhibitor protein (NAIP), the major histocompatibility complex class II transcription activator (CIITA), the incompatibility protein locus from the fungus *Podospira anserine* (HET-

E), and the mammalian telomerase-associated proteins (TP1) (Koonin and Aravind 2000; Swanson et al. 2019). After host cells membrane damage (namely potassium efflux) by candidalysin or *Candida albicans* recognition via binding of β -glucan to Dectin- 1, NLRP3 inflammasome is activated in macrophages, leading to the activation of caspase- 1 that promotes the formation of the proinflammatory cytokines interleukin- 1 β (IL- 1 β) and interleukin- 18 (IL- 18) and pyroptosis (Tavares et al. 2015; Rogiers et al. 2019; Swanson et al. 2019; Chen et al. 2023; Xu et al. 2024). In women with VVC and RVVC, the strong inflammation response driven by these cytokines, as a defense mechanism against infection, causes several symptoms such as irregular and thick white vaginal discharge, pain, burning, itching, redness, and swelling in the vulva and/or vagina (Cheng et al. 2024). Roselletti et al. (2017) have reported that the expression of NLRP3 inflammasome is significantly higher in women with VVC when compared to asymptomatic carriers. Furthermore, genetic polymorphisms in the gene encoding NLRP3 have shown to prompt exacerbation of the inflammatory response to *C. albicans* (Bruno et al. 2015), thus emphasizing the role of genetic polymorphisms of the NLRP3 inflammasome in the pathogenesis of VVC and RVVC. NLRP3 inflammasome inhibitors have gained interest as a therapeutic strategy, with several research studies reporting reduced inflammation in several disease models (Zahid et al. 2019; Mezzaroma et al. 2021; Ambrus-Aikelin et al. 2023; Xu et al. 2024; Tantra et al. 2024). A reduction in the inflammatory response in VVC, namely, the decrease of IL- 1 β expression levels, has also been shown for mice when intravaginally administered with glyburide, an effective inhibitor of the NLRP3 inflammasome (Bruno et al. 2015). This observation supports the belief that by modulating the inflammasome activity, NLRP3 inflammasome inhibitors are promising therapeutic agents in the management of VVC and RVVC, not only by alleviating the symptoms and discomforts experienced by women with VVC/RVVC but also by reducing the frequency of recurrent infections.

- d) Cytokine production: Activated epithelial cells release pro-inflammatory cytokines and chemokines that help combat *C. albicans* infections, and the application of specific pro-inflammatory cytokines or their analogs could enhance the local immune response against *C. albicans* (De Bernardis et al. 2015). However, while pro-inflammatory cytokines are essential for recruiting immune cells, excessive inflammation can lead to tissue damage and symptoms associated with vaginal candidiasis (Santoni et al. 2002) Furthermore, an overproduction of cytokines may disrupt the balance between tolerating commensal *C. albicans* and

responding to its pathogenic forms. Additionally, *C. albicans* has evolved mechanisms to modulate host cytokine responses, potentially limiting the effectiveness of this immune strategy. These mechanisms include manipulating T cell responses (Swidergall and LeibundGut-Landmann 2022), altering macrophage phenotypes, and producing cytolytic peptide toxins (Zhao et al. 2022).

Adaptive immune responses

The adaptive immune response, particularly T cell-mediated immunity, is essential for long-term protection against *C. albicans* infections. Here, several particularly important factors subsist; CD4⁺ T helper (Th) cells, DCs, CD8⁺ T cells, and antibody-mediated immunity.

- a) CD4⁺ Th cells: these, especially the Th1 and Th17 subsets, play a pivotal role in this defense. CD4⁺ T cells are regarded as the primary cell-mediated adaptive immune response against *C. albicans* (Richardson and Moyes 2015). This is evident in patients with acquired immunodeficiency syndrome (AIDS), who, due to a lack of CD4⁺ T cells, exhibit higher susceptibility to *C. albicans* infections (van de Veerdonk and Netea 2010). Th1 cells contribute to antifungal defense by producing IFN- γ , which induces nitric oxide production in macrophages and promotes the generation of *Candida*-specific antibodies (van de Veerdonk and Netea 2010). Meanwhile, the Th17 subset is a double-edged sword regarding VVC control. Th17 cells produce cytokines IL-17 (among others), which are both beneficial and detrimental in host defense against disseminated candidiasis. In one hand, overproduction of IL-17A is linked to inflammatory and neutrophil recruiting, which in turn lead to tissue damage and symptomatic VVC (Bagri et al. 2022). On the other hand, studies show that mice deficient in Th17 responses are highly susceptible to oropharyngeal candidiasis, and IL-17 receptor knockout mice display increased vulnerability to systemic *Candida* infections compared to wild-type mice (Pathakumari et al. 2024). Together, these findings underscore the vital roles of Th1 and Th17 responses in protecting against *C. albicans*.
- b) Antibody-mediated immunity: In addition to T cells, B cells and antibodies also contribute to protection, albeit to a lesser extent (Santoni et al. 2002). For example, authors have demonstrated that when B cells were transferred to naive rats, these animals showed fewer *Candida* sp. colony-forming units compared to controls. However, the rate of fungal clearance was slower than that observed in animals receiving immune T cells (Santoni et al. 2002).
- c) Dendritic cells: although not part of the adaptive system, they play a pivotal role in bridging innate and

adaptive immunity by driving specific T cell responses against *C. albicans*. Different subsets of DCs are specialized in promoting distinct immune pathways. For instance, Langerhans cells, a type of DC found in epithelial tissues, are key in eliciting Th17 responses, which are critical for antifungal defense. On the other hand, Langerin⁺ dermal DC stimulate Th1 responses and cytotoxic T lymphocyte activity, thereby contributing to a broader immune response that includes both antifungal and cytotoxic mechanisms (Richardson and Moyes 2015).

- d) CD8⁺ T cells: Although CD8⁺ T cells are less effective than CD4⁺ T cells in controlling *C. albicans* infections, they still play a contributory role in fungal clearance. By recognizing and targeting infected cells, CD8⁺ T cells add an additional layer of defense to the immune response. Their cytotoxic activity, although not as central as the helper functions of CD4⁺ T cells, supports overall fungal elimination, particularly in disseminated infections where robust immune engagement is required (Santoni et al. 2002).

The right balance

Maintaining a balance between immune activation and tolerance is crucial for preventing unnecessary inflammation while ensuring effective responses to *C. albicans* infections. Several factors are key to prevent infection in this context:

- a) Commensal vs. pathogenic forms: The immune system must accurately distinguish between commensal and pathogenic forms of *C. albicans* to prevent harmful immune overactivation. In its commensal state, *C. albicans* exists as a harmless yeast, coexisting with the host without causing tissue damage. However, under certain conditions, such as immunosuppression or environmental changes, it can switch into its pathogenic hyphal form, triggering immune activation (Gaziano et al. 2023). This distinction is primarily achieved through pattern recognition receptors (PRRs), such as C-type lectin receptors (CLRs) like Dectin-1 and Dectin-2, which sense fungal cell wall components and distinguish morphotypes. The epithelial cells and immune cells work together to detect fungal load, morphology, and other danger signals, ensuring that only pathogenic forms elicit a significant immune response.
- b) Immune tolerance: in healthy women, VECs exhibit a remarkable ability to tolerate *C. albicans* colonization without initiating an excessive inflammatory response (Gaziano et al. 2023). This immune tolerance is essential for maintaining a symbiotic relationship with *C. albicans*, preventing unnecessary tissue damage caused

by immune overactivation. VECs achieve this through tightly regulated signaling pathways that modulate the production of pro-inflammatory cytokines and antimicrobial peptides in response to commensal fungal forms (Gaziano et al. 2023).

Together, the delicate balance between immune activation and tolerance relies on a dynamic interplay of host recognition systems, fungal behavior, and environmental factors. Understanding how this balance is maintained, and what factors disrupt it, is key to identifying strategies for preventing and managing vaginal candidiasis.

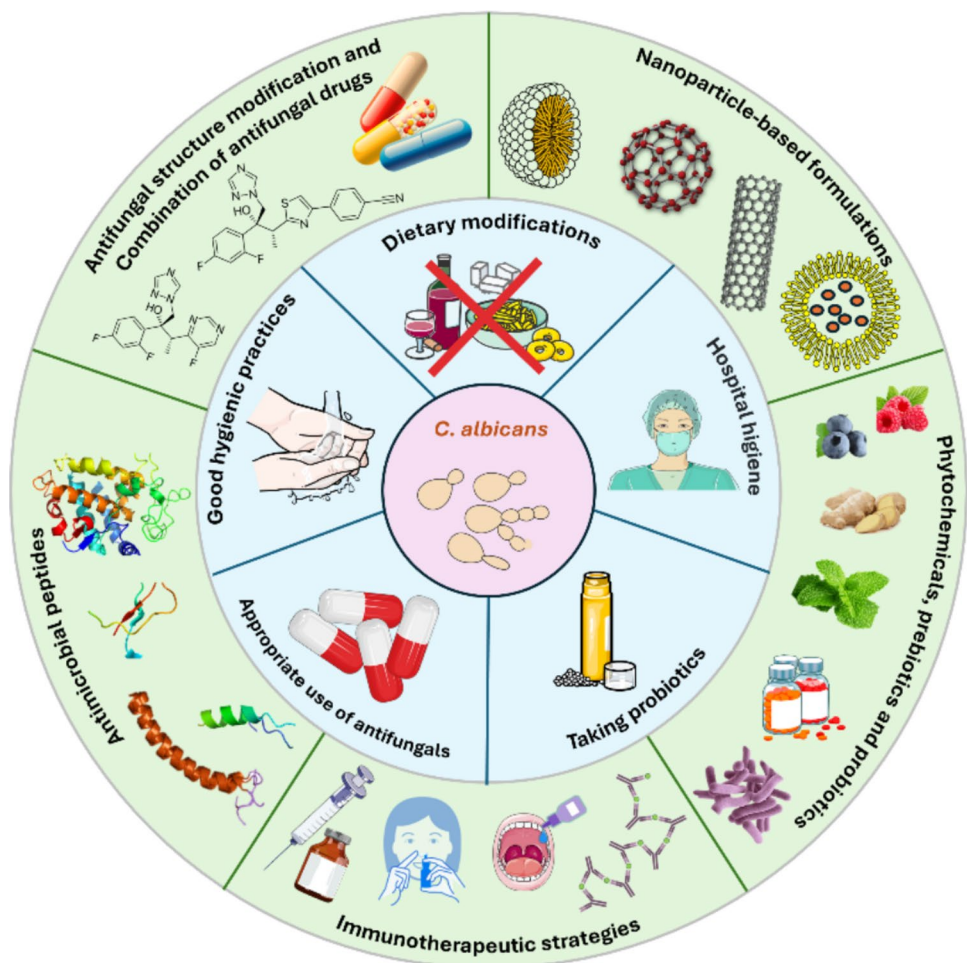
Current prevention strategies

C. albicans infections are practically inevitable, as these opportunistic fungal pathogens are ubiquitous commensals of humans (Shao et al. 2022), while asymptotically colonizing the skin and mucosal surfaces of the oral cavity and the gastrointestinal and reproductive tracts of healthy individuals (Achkar and Fries 2010; Alonso-Monge et al. 2021; Lemberg et al. 2022). *C. albicans* can switch from harmless

to pathogenic under certain conditions such as microbiota dysbiosis, impaired immune system, damage to the skin or mucosal barriers, changes in environmental pH or nutrient availability, and virulence gene expression by the fungus (Fidel 1999; Talapko et al. 2021; Jacobsen 2023).

Several approaches can be used to prevent *C. albicans* infections, namely (Fig. 2), maintaining good hygienic practices (hands and bathing with suitable soap regularly, keeping the external genitalia clean and dry), wearing cotton and breathable underwear, practicing safe sex, avoiding the consumption of dispensable antibiotics, and managing diabetes (CDC 2024b; Martins et al. 2014), as patients with this disease are more susceptible to yeast infections. Taking probiotics can also be beneficial for managing *Candida* spp. infections. Indeed, different *Lactobacillus* and *Bifidobacterium* genera species have been widely used for many decades as probiotics, as they provide a health benefit to the host by inhibiting *Candida* spp. growth (Chew et al. 2015; Matsubara et al. 2016; Hernández-Bautista et al. 2020). Dietary modifications could also be beneficial to prevent *C. albicans* overgrowth. Accordingly, alcoholic beverages, bread, and fermented products should be avoided, as these

Fig. 2 Conventional (inner blue circle) and emerging (outer green circle) strategies to prevent *C. albicans* infections



foods already contain a high quantity of fungi or yeast. A low-sugar diet (deprived of refined sugars, sucrose, honey, corn syrup, and fruit juice) should also be adopted since *C. albicans* uses sugar as carbon source to grow. In addition, allergenic foods such as milk, dairy products, eggs, wheat, or peanuts should be removed from the diet to prevent the weakening of the immune system and subsequent growth of the pathogen (Martins et al. 2014).

Nosocomial *Candida* spp. infections can also be prevented by adopting standard precaution methodologies. To avoid the dissemination of these hospital-acquired fungal infections, it is crucial for healthcare workers to wear personal protective equipment (gowns, gloves, masks, eye and face shields, shoe covers, head covers...), frequently wash their hands with soap (plain or with antiseptic), and rub them with an alcohol-based sanitizer (CDC 2024c). Noncritical environmental surfaces must be cleaned and disinfected, as well as medical equipment, instruments, and devices that require thorough decontamination before being used on a different patient. Indwelling devices (such as enteral feeding tubes, endotracheal tubes, arterial and central venous and catheters, urinary catheters and drains) should be used only, when necessary, inserted after disinfection of the skin and removed as early as possible. Additionally, high risk patients should be given appropriate antifungal prophylaxis, infected patients should be kept apart from other patients (either in single room isolation or cohousing), and the use of unnecessary antibiotics (particularly broad-spectrum antibiotics) on patients must be avoided (Ture and Alp 2018; CDC 2024c). However, *C. albicans* infections can still occur and become threatening if not treated properly.

Current available pharmacotherapeutic approaches for *C. albicans* treatment rely only on four classes of US Food and Drug Administration (FDA) approved antifungal drugs: polyenes, azoles, echinocandins, and 5-flucytosine (Salazar et al. 2020; Wall and Lopez-Ribot 2020). The following table (Table 1) summarizes the main classes of antifungal drugs, some examples and mechanisms of action.

The increased use of antifungal agents over the years resulted in the emergence of antifungal resistance, namely, among *Candida* species. Resistance phenomenon in yeast can be intrinsic (primary) or acquired (secondary). Intrinsic resistance occurs prior to antifungal exposure because of their fundamental structure that hinders the binding to its drug target (de Oliveira Santos et al. 2018; Czajka et al. 2023). This is the case, for example, of the fluconazole-resistant *C. krusei* or the less echinocandins-susceptible *C. parapsilosis* (Salazar et al. 2020; Murphy and Bicanic 2021; Czajka et al. 2023). Differently, acquired resistance is an evolutionary response to antifungal selective pressure, leading to genetic mutations such as modification or elimination of antifungal drug target and protein channels required for the uptake of the antifungals by the yeasts, modification of

Table 1 Overview of antifungal drug classes and their mechanisms of action

| Class of antifungal drugs | Examples | Mechanism of action | References |
|---------------------------|--|--|--|
| Polyenes | Amphotericin B, nystatin | Penetrate fungal cell membrane and bind to ergosterol, forming pores that cause leakage of cellular content, leading to cell death | (de Oliveira Santos et al. 2018; Salazar et al. 2020; Wall and Lopez-Ribot 2020) |
| Azoles | Fluconazole, ketoconazole, clotrimazole, butoconazole, miconazole, econazole, itraconazole | Inhibit ergosterol biosynthesis, reducing membrane integrity, resulting in cell lysis and death | (Maertens 2004; de Oliveira Santos et al. 2018; Scorzoni et al. 2021) |
| Echinocandins | Caspofungin, anidulafungin, micafungin | Inhibit β -(1,3)-D-glucan synthase, blocking β -(1,3)-D-glucan synthesis, a key fungal cell wall component, causing cell lysis and death | Death (Salazar et al. 2020; Mroczyńska and Brillowska-Dąbrowska 2020; Szymański et al. 2022) |
| 5-Flucytosine | 5-Fluorocytosine | Enters fungal cells via cytosine permeases and is converted to 5-fluorouracil, inhibiting RNA and DNA synthesis, leading to cell death | (Salazar et al. 2020; Wall and Lopez-Ribot 2020) |

Unfortunately, none of these conventional antifungal drugs has the optimum profile, as all of them are associated with some limitations

the composition of the fungal cell membrane, or the over-expression of efflux pumps critical for the expelling of the drugs from yeasts (de Oliveira Santos et al. 2018; Wall and Lopez-Ribot 2020; Murphy and Bicanic 2021; Czajka et al. 2023). Examples of this type of resistance mechanisms have been observed in *Candida* species such as *C. albicans*, *C. glabrata*, and *C. auris*, in response to azoles, echinocandins, polyenes, and 5-flucytosine (Salazar et al. 2020; Murphy and Bicanic 2021; Czajka et al. 2023). Additionally, phenotypic alterations often decrease fungal susceptibility to antifungals. Indeed, *Candida* species are known to form biofilms, thus living within a self-produced polymeric matrix that acts as an additional barrier that limits the distribution of the antifungal drugs (Murphy and Bicanic 2021; Kaur and Nobile 2023; Czajka et al. 2023).

Other limitations of the current conventional antifungal drugs regard their pharmacokinetics and cytotoxicity (Ostrosky-Zeichner et al. 2010; de Oliveira Santos et al. 2018; Wall and Lopez-Ribot 2020; Roy et al. 2023; Czajka et al. 2023). For example, within the azole family, not all antifungals can be used as systemic medication. Due to their high toxicity and low bioavailability, imidazole-based azoles (such as clotrimazole, miconazole, econazole, butoconazole, or ketoconazole) can only be formulated for superficial fungal infections (Ostrosky-Zeichner et al. 2010). While exhibiting low toxicity, echinocandins have the great disadvantage exhibiting poor oral bioavailability due to their larger molecular weight (being available only in intravenous formulations), and amphotericin-B, although extremely efficient, has been reported to be responsible for nephrotoxicity, electrolyte irregularities, and anaphylaxis events, besides being practically insoluble in water (Wall and Lopez-Ribot 2020; Murphy and Bicanic 2021). In addition, recurrent usage of many antifungal agents is limited due to their nephrotoxicity and hepatotoxicity (de Oliveira Santos et al. 2018; Murphy and Bicanic 2021).

To overcome all the abovementioned challenges related to the current antifungal pharmacotherapies, novel approaches must be exploited.

Innovative and emerging prevention methods

To minimize the different limitations associated with conventional antifungal drugs, different pharmacological strategies have been exploited (Fig. 2). Structure modifications and new formulations of the available antifungals have been explored for the development of new antifungal compounds. New modified triazoles (e.g., ravuconazole, albaconazole, and isavuconazole) and tetrazoles (oteseconazole) have been shown to have in vitro activity against *Candida* species such as *C. albicans*, *C. glabrata*, *C. parapsilosis*, *C. krusei*, and *C. auris* strains while exhibiting low cytotoxicity. Specifically, voriconazole and posaconazole, two second

generation triazoles with broad-spectrum activity, have been reported to have superior antifungal activity against most *Candida* species (de Oliveira Santos et al. 2018; Scorzoni et al. 2021). Similarly, alterations in the molecular structure of echinocandins give rise to new antifungal molecules (such as rezafungin) with reduced probability to elicit fungal resistance, increased antifungal potency against *Candida* species, and better pharmacokinetic profile (Scorzoni et al. 2021; Jospe-Kaufman et al. 2024). Liposomal formulations of amphotericin B were also developed aiming to surpass its limitations. The incorporation of amphotericin B into liposomes, lipid complexes, or colloidal dispersions allowed the development of new antifungal targeted therapies while reducing its toxicity and enhancing its pharmacokinetics (Torrado et al. 2008; Salazar et al. 2020). Three intravenous infusions of these liposomal formulations (Ambisome®, Abelcet®, and Amphotec®) are already commercially available (Torrado et al. 2008), and an oral formulation (CAmB/MAT2203) is under clinical trials for the treatment of invasive candidiasis (Jenssen et al. 2006).

Exploitation of antimicrobial peptides is a promising alternative to antifungal drugs. Ubiquitous in nature, these short, amphipathic, positively charge molecules are acknowledged to electrostatically interact with the membrane, leading to membrane permeabilization and consequent inhibition of nucleic and ribonucleic acid synthesis, protein and cell wall synthesis, and enzymatic activity (Jensen et al. 2006). Within these antimicrobial peptides, several have been reported to have antifungal activity, namely, anti-*Candida* activity in both planktonic and sessile physiological states (Fernández de Ullivarri et al. 2020; Perez-Rodriguez et al. 2022). These include human defensins- 1, - 2, and - 3 (HBD- 1, HBD- 2, and HBD- 3, respectively), found to be active against *C. albicans*, human lactoferrin (hLF) that has been reported to display activity against *C. albicans* and *C. krusei*, bovine cateslytin (bCAT), found to be active against *C. albicans*, *C. glabrata*, and *C. tropicalis*, human histatin- 5 (Hst 5) that has revealed activity against *C. albicans* and to inhibit its biofilm formation, LL- 37, that holds antifungal activity against planktonic and sessile *C. albicans* and *C. auris*, and also lysozyme that has been demonstrated to be active against several *Candida* species (de Oliveira Santos et al. 2018; Perez-Rodriguez et al. 2022).

Another strategy to enhance the therapeutic efficiency of the existing antifungal drugs consists in their combination with each other or with new molecules, as it can overcome their toxicity hurdle by reducing the individual doses of each agent, besides reducing the risk of antimicrobial resistance emergence due to the implication of multiple drug targets simultaneously (Scorzoni et al. 2021). The synergetic effect of the combination of 5-flucytosine and amphotericin B has been reported to have an improved effect to treat invasive candidiasis (Scorzoni et al. 2021). Similarly, the

combination of azoles and echinocandins (e.g., fluconazole + micafungin, voriconazole + micafungin, posaconazole + caspofungin), the combination of polyenes and echinocandins (e.g., amphotericin B + caspofungin, amphotericin B + anidulafungin, amphotericin + micafungin), and the combination of polyenes and azoles (e.g., amphotericin + posaconazole) have shown that their synergistic antifungal effect against *Candida* spp. infections and biofilms was enhanced when compared to their individual activities (de Oliveira Santos et al. 2018; Scorzoni et al. 2021). Moreover, the combination of lactofungin, a lactoferrin-derived antifungal peptide, with amphotericin B demonstrated the potential of this new compound to be efficient used in *anti-Candida* therapies while using a low dosage and, thus, prompting a reduction on amphotericin B cytotoxicity (Fernandes et al. 2020).

As stated before, nanoparticles such as liposomes have been exploited as antifungal delivery systems to fight candidiasis fungal infections. Different nano systems have been assessed for alternative antifungal therapies such as solid lipid nanoparticles, nanostructured lipid carriers, polymers, and metals with innate antimicrobial properties (e.g., zinc, gold, and silver). Their small size and high surface area-to-volume ratio allows the encapsulation, adsorption or chemical attachment of considerable amounts of antifungal agents, and transportation to specific targets, without being recognized by efflux pump proteins (Salazar et al. 2020; Scorzoni et al. 2021). Nanostructured lipid carriers loaded with amphotericin B revealed enhanced antifungal activity when compared with the commercial amphotericin B colloidal system Fungizone®, and solid lipid nanoparticles loaded with fluconazole demonstrated improved activity against *C. albicans*, *C. glabrata*, and *C. parapsilosis* (Scorzoni et al. 2021). Likewise, fluconazole-loaded chitosan nanoparticles exhibited considerable activity against the same *Candida* species, and silver nanoparticles (either alone or conjugated with fluconazole) have also been shown to be highly effective against *Candida* species (Salazar et al. 2020; Scorzoni et al. 2021).

In recent years, immunotherapeutic strategies have been gaining a special interest for the treatment of *Candida* infections. Particularly, great efforts are being made in the development of vaccines to prevent *Candida* infections through stimulation of the immune system to recognize the fungus and consequently attack it by eliciting an immune response. Although no *Candida* sp. vaccines are currently clinically available, two formulations are already in clinical trials (Costa-Barbosa et al. 2023; van de Veerdonk et al. 2010), and many others are being developed and assessed for their application in *anti-Candida* therapies. Several types of vaccines with different routes of administration (injectable, oral, intranasal) are being explored to prevent *Candida* infections (Sahu et al. 2022;

Alapan et al. 2024). This immunotherapeutic approach has significantly evolved by taking advantage of genetic editing techniques such as the clustered regularly interspaced short palindromic repeats (CRISPR)-Cas9 system that not only allows the identification of new targets for novel antifungal compounds but also allows the creation of attenuated *Candida* strains that can be used as live vaccines (Uthayakumar et al. 2020). Live attenuated whole-cells are designed to contain a weakened form of the whole fungus that will cause a minor infection which will not evolve to a severe disease but instead will produce a robust long-lasting immune response against subsequent exposure to the pathogen (Sahu et al. 2022; Alapan et al. 2024). This kind of vaccination has already proved to avoid reinfection in animal models after being inoculated with a low virulent *C. albicans* strains (van de Veerdonk et al. 2010; Sahu et al. 2022). However, some concerns regarding this type of vaccines concern their application in immunocompromised patients, who may become seriously ill after inoculation (Sahu et al. 2022). Differently, killed whole cell vaccines use inactivated *Candida* cells (killed by radiation, chemicals, or heat) to stimulate an immune response, thus being safer for patients with a reduced ability to fight infections and diseases (Sahu et al. 2022; Alapan et al. 2024). Indeed, inoculation of inactivated *C. neoformans* conferred immune protection against several fungi pathogens in immunocompetent and immunocompromised animal models (Alapan et al. 2024). Alternative *anti-Candida* vaccines have been designed to use only parts of *Candida* such as proteins, peptides and polysaccharides, or genetic material, such as DNA and RNA, to trigger an immune response (Sahu et al. 2022; Alapan et al. 2024). Recombinant vaccines based of *C. albicans* adhesins (agglutinin-like sequence 1 and 3; Als1p and Als3p) and *C. parapsilosis* secreted aspartic protease 2 (Sap2) protein revealed enhanced immunological protection against fungal infections in animal models (van de Veerdonk et al. 2010; Sahu et al. 2022), and a heat shock protein from *C. albicans* (hsp90-CA) encoding DNA vaccine showed a protective immunological response to vaginal candidiasis in animal models. In addition, conjugate fungal vaccines composed of fungal antigen such as (polysaccharides) covalently linked to carrier proteins have also demonstrated to be a promising alternative, namely, against *Candida* vaginal infections (van de Veerdonk et al. 2010; Sahu et al. 2022).

Non-pharmacological approaches, using phytochemicals, prebiotics, and probiotics, have also been exploited against candidiasis. Several oily formulations of plant extracts have already been topically used as therapeutics for vulvovaginal candidiasis, namely, oils from lemon balm, garlic, fennel, chamomile, ginger, and sage (Picheta et al. 2024). Additionally, molecules such as allicin, curcumin, cannabidiol

(CBD), and dill oil have shown to have a beneficial effect on vulvovaginal candidiasis, whereas green tea, cinnamon, garlic, propolis, and ginger revealed *anti-Candida* activity against oral candidiasis (Gharibpour et al. 2021; Picheta et al. 2024). These findings support the idea that plant-based bioactive compounds can be a good option for the treatment of different *Candida* infections. Finally, it is well known that the consumption of probiotics has several benefits for human gut health. Similarly, the use of these “generally recognized as safe” (GRAS) live microorganisms has also been used for the maintenance and improvement of women vaginal microenvironment. When the abundance of lactobacilli within the vaginal tract (mainly *L. gasseri*, *L. jensenii*, and *L. crispatus*) decreases, an increase on the pathogenesis of *Candida* species (namely, *C. albicans* and *C. glabrata*) is observed (Salazar et al. 2020). For that reason, the consumption of probiotic lactobacilli can enhance and restore vaginal homeostasis, thus inhibiting the growth of pathogenic fungi and the formation of fungal biofilms, as it has been widely demonstrated (Wu et al. 2022; Liu et al. 2023). Indeed, several combinations of lactobacilli strains, to be administered either via oral or intravaginal route, are currently under clinical trials (Liu et al. 2023). Furthermore, studies regarding the combination of lactobacilli with lactoferrin (Superti De Seta 2020) and the synergistic effect obtained from the combination of lactobacilli with mannan oligosaccharides (Faustino et al. 2024) demonstrated an improvement in the health of women with vaginal fungal infections and an inhibitory effect on the adhesion of the *C. albicans*, thus highlighting the potential of probiotics and prebiotics as efficient therapeutic candidates for the treatment of fungal infection.

Ultimately, all the abovementioned strategies reported interesting results that represent a significant advancement in the ongoing development of new therapeutic approaches against candidiasis.

Commercial products

The most common treatments for yeast vaginal infections involve antifungal medications. These can be administered in various forms, including creams, ointments, tablets, and suppositories. Preferable options consist in the use of a cream applied topically in the vaginal area (and used daily for up to 7 days), or a single dose of fluconazole taken orally. Clotrimazole (Lotrimin) and miconazole (Monistat) are widely used and typically effective for mild to moderate infections, as well as oral medication such as fluconazole (Diflucan) (Cleveland Clinic 2022). A list of the top sellers from Amazon is available in Table 2.

When infections are resistant to standard treatments, more doses of fluconazole or boric acid capsules

(topically administered) may be recommended (CDC 2024 d; Mayo Clinic 2024). Another two drugs that have been recently approved by the FDA for recurrent VVC are ibrexafungerp (Brexafemme) in 2021 and oteseconazole (Vivjoa) in 2022. Only the former was approved in the EU by the European Medicines Agency (EMA).

As previously mentioned, natural treatments (e.g., probiotics, tea tree oil, coconut oil, and garlic) can also be an option for those looking to avoid the side effects of conventional medications or for mild infections. These products are known for their antifungal properties, although the scientific validation for their action against *Candida* infections is often absent. Furthermore, they are generally not as well-studied or as effective as conventional antifungal medications. Nevertheless, and especially for probiotics, they are usually also in the consumers list of preferences, as observed in Table 2.

The global yeast infection treatment market size for VV candidiasis was valued at 4.37 billion in 2023 and is expected to reach USD 7.01 billion by 2031, with a compound annual growth rate (CAGR) of 6.1% during the forecast period of 2024 to 2031. These predictions are based on the rise in autoimmune disorders and yeast infections, as well as market expansion due to the global economic recovery, which enables consumers to access better and higher-quality healthcare (Sabyasachi Ghosh 2023; Data Bridge 2024).

Conclusion

Vaginal infections caused by *Candida albicans* remain a significant global health concern due to their high recurrence rates and negative impact on quality of life. Understanding the pathogenesis of *C. albicans* infections, particularly its key virulence factors such as biofilm formation, adherence, and phenotypic switching, is crucial for developing more effective prevention and treatment strategies. While antifungal therapies, probiotics, and hygiene practices are widely used, challenges such as antifungal resistance, recurrence, and limited efficacy highlight the need for innovative approaches. Emerging strategies, including novel antifungal agents (e.g., antimicrobial peptides), vaccines, and nanotechnology-based delivery systems (e.g., liposomes), offer promising solutions to improve outcomes and reduce dependency on current treatments.

The immune system plays a critical role in preventing *C. albicans* infections, emphasizing the importance of both innate and adaptive immune responses in restricting fungal colonization and overgrowth. Despite the availability of commercial products such as antifungal creams and/or oral pills and pre-probiotics, long-term efficacy remains limited. Addressing persistent challenges, including patient

Table 2 Available commercial products, presentation format, claims, ingredients concentration per serving, treatment duration, and goal

| Producer | Product | Presentation | Claims | Ingredients concentration per serving | Treatment duration | Treatment goal |
|---------------|-----------------------------|--------------------------------|---|--|---|-------------------------------|
| NutraBlast | Boric Life | Vaginal suppositories | Supports odor control; promotes vaginal balance | Boric acid powder (600 mg) | 1 for 7 days | Treatment |
| Lemme | Lemme Purr | Vaginal probiotic gummies | Balanced pH, healthy odor, yeast balance, and flora support + vitamin C for immune health | <i>Bacillus coagulans</i> SNZ 1969 (1 billion CFU), pineapple (<i>Ananas comosus</i>) powder (100 mg), vitamin C (20 mg) | 2 gummies daily | Preventive dietary supplement |
| Monistat | Monistat 1 combination pack | Vaginal insert + vaginal cream | Cures most vaginal yeast infections; relieves associated external itching and irritation | Miconazole nitrate 1200 mg in vaginal insert and 2% in external cream | External cream – 2 times daily for up to 7 days | Treatment |
| Love Wellness | Love Wellness – the killer | Vaginal suppositories | Controls vaginal odor and promotes a fresh scent | Boric acid (600 mg) | Use after being intimate, at the end of period, or when experiencing irritation from pH imbalance (for up to 14 days) | Treatment |
| Azo | Yeast Plus | Oral tablets | Relief from vaginal itching, burning, odor, and discharge | <i>Candida albicans</i> and other homeopathic ingredients | 1 tablet, 3 times a day, as long as symptoms persist | Symptom relief |
| Amazon | Miconazole 3 | Vaginal cream | Yeast infection relief | Miconazole nitrate 4% | 3 days | Treatment |
| Amazon | Tioconazole 1 | Ointment | Yeast infection relief | Tioconazole 300 mg (6.5%) | 1 dose treatment | Treatment |

compliance, access to care, and resistance, requires an interdisciplinary approach that integrates personalized medicine, advanced therapeutic options, and improved healthcare accessibility.

Future research efforts must focus on bridging these gaps to advance prevention and treatment strategies. By fostering a multi-faceted approach that combines scientific innovation with clinical application, significant strides can be made toward reducing the burden of *C. albicans* infections and improving patient quality of life.

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Declarations

Ethics approval This article does not contain any studies with human participants or animals performed by any of the authors.

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References

- Achkar JM, Fries BC (2010) *Candida* infections of the genitourinary tract. Clin Microbiol Rev 23:253–273. <https://doi.org/10.1128/CMR.00076-09>
- Alapan D, Bisweswar O, Prasenjit S, Prasanjit D, Arkapal B (2024) Recent advances in the clinical development of antifungal vaccines: a narrative review. Front Trop Dis 5:1446477. <https://doi.org/10.3389/FITD.2024.1446477/BIBTEX>
- Alonso-Monge R, Gresnigt MS, Román E, Hube B, Pla J (2021) *Candida albicans* colonization of the gastrointestinal tract: a double-edged sword. PLoS Pathog 17:e1009710. <https://doi.org/10.1371/journal.ppat.1009710>
- Ambrus-Aikelin G, Takeda K, Joetham A, Lazic M, Povero D, Santini AM, Pranadinata R, Johnson CD, McGeough MD, Beasley FC, Stansfield R, McBride C, Trzoss L, Hoffman HM, Feldstein AE, Stafford JA, Veal JM, Bain G, Gelfand EW (2023) JT002, a small molecule inhibitor of the NLRP3 inflammasome for the treatment of autoinflammatory disorders. Sci Rep 13:13524. <https://doi.org/10.1038/s41598-023-39805-z>
- Bagri P, Anipindi VC, Kaushic C (2022) The role of IL-17 during infections in the female reproductive tract. Front Immunol 13. <https://doi.org/10.3389/fimmu.2022.861444>
- Barousse MM, Espinosa T, Dunlap K, Fidel PL (2005) Vaginal epithelial cell anti-*Candida albicans* activity is associated with protection against symptomatic vaginal candidiasis. Infect Immun 73:7765–7767. <https://doi.org/10.1128/IAI.73.11.7765-7767.2005>
- Bras G, Satala D, Juszcak M, Kulig K, Wronowska E, Bednarek A, Zawrotniak M, Rapala-Kozik M, Karkowska-Kuleta J (2024) Secreted aspartic proteinases: key factors in *Candida* infections and host-pathogen interactions. Int J Molecular Sci 25:4775. <https://doi.org/10.3390/IJMS25094775>
- Bruno VM, Shetty AC, Yano J, Fidel PL, Noverr MC, Peters BM (2015) Transcriptomic analysis of vulvovaginal candidiasis identifies a role for the NLRP3 inflammasome. mBio 6. <https://doi.org/10.1128/mBio.00182-15>
- Cangui-Panchi SP, Nacato-Toapanta AL, Enríquez-Martínez LJ, Salinas-Delgado GA, Reyes J, Garzon-Chavez D, Machado A (2023) Battle royale: immune response on biofilms – host-pathogen interactions. Curr Res Immunol 4:100057. <https://doi.org/10.1016/J.CRIMMU.2023.100057>
- CDC (2024) Candidiasis basics. <https://www.cdc.gov/candidiasis/about/index.html>. Accessed 18 Dec 2024
- CDC (2024) Preventing candidiasis. <https://www.cdc.gov/candidiasis/prevention/index.html>. Accessed 24 Nov 2024
- CDC (2024) Standard precautions for all patient care. <https://www.cdc.gov/infection-control/hcp/basics/standard-precautions.html>. Accessed 24 Nov 2024
- CDC (2024) Treatment of candidiasis. <https://www.cdc.gov/candidiasis/treatment/index.html>. Accessed 18 Dec 2024
- Ceccarani C, Foschi C, Parolin C, D'Antuono A, Gaspari V, Consolandi C, Laghi L, Camboni T, Vitali B, Severgnini M (2019) Marangoni A (2019) Diversity of vaginal microbiome and metabolome during genital infections. Sci Rep 9(1):1–12. <https://doi.org/10.1038/s41598-019-50410-x>
- Chee WJY, Chew SY (2020) Than LTL (2020) Vaginal microbiota and the potential of *Lactobacillus* derivatives in maintaining vaginal health. Microb Cell Factories 19(1):1–24. <https://doi.org/10.1186/S12934-020-01464-4>
- Chen Y, Ye X, Escames G, Lei W, Zhang X, Li M, Jing T, Yao Y, Qiu Z, Wang Z, Acuña-Castroviejo D, Yang Y (2023) The NLRP3 inflammasome: contributions to inflammation-related diseases. Cell Mol Biol Lett 28:51. <https://doi.org/10.1186/s11658-023-00462-9>
- Cheng KO, Montañó DE, Zelante T, Dietschmann A, Gresnigt MS (2024) Inflammatory cytokine signalling in vulvovaginal candidiasis: a hot mess driving immunopathology. Oxf Open Immunol 5. <https://doi.org/10.1093/oxfimm/iqae010>
- Chew SY, Cheah YK, Seow HF, Sandai D, Than LTL (2015) Probiotic *Lactobacillus rhamnosus* GR-1 and *Lactobacillus reuteri* RC-14 exhibit strong antifungal effects against vulvovaginal candidiasis-causing *Candida glabrata* isolates. J Appl Microbiol 118:1180–1190. <https://doi.org/10.1111/JAM.12772>

- Chow EWL, Pang LM, Wang Y (2021) From Jekyll to Hyde: the yeast–hyphal transition of *Candida albicans*. *Pathogens* 10. <https://doi.org/10.3390/PATHOGENS10070859>
- Ciurea CN, Kosovski IB, Mare AD, Toma F, Pinteá-Simon IA, Man A (2020) *Candida* and candidiasis—opportunism versus pathogenicity: a review of the virulence traits. *Microorganisms* 8:1–17. <https://doi.org/10.3390/MICROORGANISMS8060857>
- Cleveland Clinic (2022) Vaginal yeast infection: causes, symptoms & treatment. <https://my.clevelandclinic.org/health/diseases/5019-vaginal-yeast-infection>. Accessed 18 Dec 2024
- Costa-Barbosa A, Pacheco MI, Carneiro C, Botelho C, Gomes AC, Real Oliveira MECD, Collins T, Vilanova M, Pais C, Correia A, Sampaio P (2023) Design of a lipid nano-delivery system containing recombinant *Candida albicans* chitinase 3 as a potential vaccine against fungal infections. *Biomed Pharmacother* 166:115362. <https://doi.org/10.1016/J.BIOPHA.2023.115362>
- Czajka KM, Venkataraman K, Brabant-Kirwan D, Santi SA, Verschoor C, Appanna VD, Singh R, Saunders DP, Tharmalingam S (2023) Molecular mechanisms associated with antifungal resistance in pathogenic *Candida* species. *Cells* 2:2655. <https://doi.org/10.3390/CELLS12222655>
- D'Enfert C, Kaune AK, Alaban LR, Chakraborty S, Cole N, Delavy M, Kosmala D, Marsaux B, Fróis-Martins R, Morelli M, Rosati D, Valentine M, Xie Z, Emritoll Y, Warn PA, Bequet F, Bounoux ME, Bornes S, Gresnigt MS, Hube B, Jacobsen ID, Legrand M, Leibundgut-Landmann S, Manichanh C, Munro CA, Netea MG, Queiroz K, Roget K, Thomas V, Thorat C, Van Den Abbeele P, Walker AW, Brown AJP (2021) The impact of the fungus–host–microbiota interplay upon *Candida albicans* infections: current knowledge and new perspectives. *FEMS Microbiol Rev* 45:14. <https://doi.org/10.1093/FEMSRE/FUAA060>
- Data Bridge (2024) Yeast infection market size, share & trends report by 2031. <https://www.databridgemarketresearch.com/reports/global-yeast-infection-market>. Accessed 18 Dec 2024
- de Oliveira Santos GC, Vasconcelos CC, Lopes AJO, de Sousa Cartágenes M do S, Filho AKDB, do Nascimento FRF, Ramos RM, Pires ERB, de Andrade MS, Rocha FMG, de Andrade Monteiro C (2018) *Candida* infections and therapeutic strategies: mechanisms of action for traditional and alternative agents. *Front Microbiol* 9. <https://doi.org/10.3389/fmicb.2018.01351>
- De Bernardis F, Arancia S, Sandini S, Graziani S, Norelli S (2015) Studies of immune responses in *Candida vaginitis*. *Pathogens* 4:697–707. <https://doi.org/10.3390/pathogens4040697>
- Dong M, Dong Y, Bai J, Li H, Ma X, Li B, Wang C, Li H, Qi W, Wang Y, Fan A, Han C, Xue F (2023) Interactions between microbiota and cervical epithelial, immune, and mucus barrier. *Front Cell Infect Microbiol* 13. <https://doi.org/10.3389/fcimb.2023.1124591>
- Dühring S, Germerodt S, Skerka C, Zipfel PF, Dandekar T, Schuster S (2015) Host–pathogen interactions between the human innate immune system and *Candida albicans*—understanding and modeling defense and evasion strategies. *Front Microbiol* 6:625. <https://doi.org/10.3389/FMICB.2015.00625>
- Faustino M, Pereira JO, Pereira AM, Oliveira AS, Ferreira CMH, Pereira CF, Durão J, Pintado ME, Carvalho AP (2024) Vaginal prevention of *Candida albicans*: synergistic effect of lactobacilli and mannan oligosaccharides (MOS). *Appl Microbiol Biotechnol* 108:1–17. <https://doi.org/10.1007/S00253-023-12909-2/FIGURES/7>
- Fernandes KE, Payne RJ, Carter DA (2020) Lactoferrin-derived peptide lactofungin is potently synergistic with amphotericin B. *Antimicrob Agents Chemother* 64. https://doi.org/10.1128/AAC.00842-20/SUPPL_FILE/AAC.00842-20-SD002.XLSX
- Fernández de Ullivarri M, Arbulu S, García-Gutierrez E, Cotter PD (2020) Antifungal peptides as therapeutic agents. *Front Cell Infect Microbiol* 10:518678. <https://doi.org/10.3389/FCIMB.2020.00105/BIBTEX>
- Fidel PL (1999) *Candida albicans*: from commensal to pathogen. Medical Importance of the Normal Microflora. Springer, US, Boston, MA, pp 441–476
- Galocha M, Pais P, Cavalheiro M, Pereira D, Viana R, Teixeira MC (2019) Divergent approaches to virulence in *C. albicans* and *C. glabrata*: two sides of the same coin. *Int J Mol Sci* 20(9):2345. <https://doi.org/10.3390/IJMS20092345>
- García-Rubio R, de Oliveira HC, Rivera J, Trevijano-Contador N (2019) The fungal cell wall: *Candida*, *Cryptococcus*, and *Aspergillus* species. *Front Microbiol* 10. <https://doi.org/10.3389/FMICB.2019.02993>
- Gaziano R, Sabbatini S, Monari C (2023) The interplay between *Candida albicans*, vaginal mucosa, host immunity and resident microbiota in health and disease: an overview and future perspectives. *Microorganisms* 11. <https://doi.org/10.3390/microorganisms11051211>
- Gharibpour F, Shirban F, Bagherniya M, Nosouhian M, Sathyapalan T, Sahebkar A (2021) The effects of nutraceuticals and herbal medicine on *Candida albicans* in oral candidiasis: a comprehensive review. *Adv Exp Med Biol* 1308:225–248. https://doi.org/10.1007/978-3-030-64872-5_16
- Goderidze T, Durglishvili N, Durglishvili G, Machitidze M, Odzelashvili I (2022) Frequency of vaginal *Candida* in diabetes patients – overview. *Georgian Sci* 4:315–322. <https://doi.org/10.52340/gS.2022.04.04.35>
- Hernández-Bautista LM, Márquez-Preciado R, Ortiz-Magdaleno M, Pozos-Guillén A, Aranda-Romo S, Sánchez-Vargas LO (2020) Effect of five commercial probiotic formulations on *Candida albicans* growth: in vitro study. *J Clin Pediatr Dent* 44:289–295. <https://doi.org/10.17796/1053-4625-44.5.5/HTML>
- Hernández-Chávez M, Pérez-García L, Niño-Vega G, Mora-Montes H (2017) Fungal strategies to evade the host immune recognition. *J Fungi* 3:51. <https://doi.org/10.3390/jof3040051>
- Ho J, Yang X, Nikou S-A, Kichik N, Donkin A, Ponde NO, Richardson JP, Gratacap RL, Archambault LS, Zwirner CP, Murciano C, Henley-Smith R, Thavaraj S, Tynan CJ, Gaffen SL, Hube B, Wheeler RT, Moyes DL, Naglik JR (2019) Candidalysin activates innate epithelial immune responses via epidermal growth factor receptor. *Nat Commun* 10:2297. <https://doi.org/10.1038/s41467-019-09915-2>
- Ho J, Wickramasinghe DN, Nikou S-A, Hube B, Richardson JP, Naglik JR (2020) Candidalysin is a potent trigger of alarmin and antimicrobial peptide release in epithelial cells. *Cells* 9:699. <https://doi.org/10.3390/cells9030699>
- Jacobsen ID (2023) The role of host and fungal factors in the commensal-to-pathogen transition of *Candida albicans*. *Curr Clin Microbiol Rep* 10:55–65. <https://doi.org/10.1007/s40588-023-00190-w>
- Jenssen H, Hamill P, Hancock REW (2006) Peptide antimicrobial agents. *Clin Microbiol Rev* 19:491–511. <https://doi.org/10.1128/CMR.00056-05/ASSET/8CCE8620-B1C9-4084-9712-3CC0AF74247D/ASSETS/GRAPHIC/ZCM0030621790002.JPEG>
- Jospe-Kaufman M, Ben-Zeev E, Mottola A, Dukhovny A, Berman J, Carmeli S, Fridman M (2024) Reshaping echinocandin antifungal drugs to circumvent glucan synthase point-mutation-mediated resistance. *Angew Chem Int Ed* 63:e202314728. <https://doi.org/10.1002/ANIE.202314728>
- Kaur J, Nobile CJ (2023) Antifungal drug-resistance mechanisms in *Candida* biofilms. *Curr Opin Microbiol* 71:102237. <https://doi.org/10.1016/J.MIB.2022.102237>
- Köhler JR, Hube B, Puccia R, Casadevall A, Perfect JR (2017) Fungi that infect humans. *Microbiol Spectr* 5. <https://doi.org/10.1128/MICROBIOLSPEC.FUNK-0014-2016/ASSET/D4236>

DD4-FD5C-4A73-9395-0CF33D7B696A/ASSETS/GRAPHIC/FUNK-0014-2016-FIG1.GIF

- Koonin EV, Aravind L (2000) The NACHT family – a new group of predicted NTPases implicated in apoptosis and MHC transcription activation. *Trends Biochem Sci* 25:223–224. [https://doi.org/10.1016/S0968-0004\(00\)01577-2](https://doi.org/10.1016/S0968-0004(00)01577-2)
- Lachat J, Pascault A, Thibaut D, Le Borgne R, Verbavatz JM (2022) Weiner A (2022) Trans-cellular tunnels induced by the fungal pathogen *Candida albicans* facilitate invasion through successive epithelial cells without host damage. *Nat Commun* 13:1–15. <https://doi.org/10.1038/s41467-022-31237-z>
- Lemberg C, de San M, Vicente K, Fróis-Martins R, Altmeyer S, Tran VDT, Mertens S, Amorim-Vaz S, Rai LS, d'Enfert C, Pagni M, Sanglard D, LeibundGut-Landmann S (2022) *Candida albicans* commensalism in the oral mucosa is favoured by limited virulence and metabolic adaptation. *PLoS Pathog* 18:e1010012. <https://doi.org/10.1371/journal.ppat.1010012>
- Li H, Miao MX, Jia CL, Cao YB, Yan TH, Jiang YY, Yang F (2022) Interactions between *Candida albicans* and the resident microbiota. *Front Microbiol* 13:930495. <https://doi.org/10.3389/FMICB.2022.930495/PDF>
- Lionakis MS, Drummond RA (2023) Immune responses to human fungal pathogens and therapeutic prospects. *Nat Rev Immunol* 23(7):433–452. <https://doi.org/10.1038/s41577-022-00826-w>
- Liu P, Lu Y, Li R, Chen X (2023) Use of probiotic lactobacilli in the treatment of vaginal infections: *in vitro* and *in vivo* investigations. *Front Cell Infect Microbiol* 13:1153894. <https://doi.org/10.3389/FCIMB.2023.1153894/BIBTEX>
- Lopes JP, Lionakis MS (2022) Pathogenesis and virulence of *Candida albicans*. *Virulence* 13:89. <https://doi.org/10.1080/21505594.2021.2019950>
- MacAlpine J, Daniel-Ivad M, Liu Z, Yano J, Revie NM, Todd RT, Stogios PJ, Sanchez H, O'Meara TR, Tompkins TA, Savchenko A, Selmecki A, Veri AO, Andes DR, Fidel PL, Robbins N, Nodwell J, Whitesell L, Cowen LE (2021) A small molecule produced by *Lactobacillus* species blocks *Candida albicans* filamentation by inhibiting a DYRK1-family kinase. *Nat Commun* 12(1):1–16. <https://doi.org/10.1038/s41467-021-26390-w>
- Maertens JA (2004) History of the development of azole derivatives. *Clin Microbiol Infect* 10:1–10. <https://doi.org/10.1111/j.1470-9465.2004.00841.x>
- Malinová Z, Čonková E, Váczi P (2023) Biofilm formation in medically important *Candida* species. *J Fungi* 9:955. <https://doi.org/10.3390/JOF9100955>
- Martins N, Ferreira ICFR, Barros L, Silva S, Henriques M (2014) Candidiasis: predisposing factors, prevention, diagnosis and alternative treatment. *Mycopathologia* 177:223–240. <https://doi.org/10.1007/s11046-014-9749-1>
- Mata-Martínez P, Bergón-Gutiérrez M, del Fresno C (2022) Decitin-1 signaling update: new perspectives for trained immunity. *Front Immunol* 13. <https://doi.org/10.3389/FIMMU.2022.812148>
- Matsubara VH, Bandara HMHN, Mayer MPA, Samaranyake LP (2016) Probiotics as antifungals in mucosal candidiasis. *Clin Infect Dis* 62:1143–1153. <https://doi.org/10.1093/CID/CIW038>
- Mayer FL, Wilson D, Hube B (2013) *Candida albicans* pathogenicity mechanisms. *Virulence* 4:119. <https://doi.org/10.4161/VIRU.22913>
- Mayo Clinic (2024) Yeast infection (vaginal) - diagnosis and treatment. <https://www.mayoclinic.org/diseases-conditions/yeast-infection/diagnosis-treatment/drc-20379004>. Accessed 18 Dec 2024
- Maza PK, Bonfim-Melo A, Padovan ACB, Mortara RA, Orikaza CM, Ramos LMD, Moura TR, Soriani FM, Almeida RS, Suzuki E, Bahia D (2017) *Candida albicans*: the ability to invade epithelial cells and survive under oxidative stress is unlinked to hyphal length. *Front Microbiol* 8:229839. <https://doi.org/10.3389/FMICB.2017.01235/BIBTEX>
- Méhot PO, Alizon S (2014) What is a pathogen? toward a process view of host-parasite interactions. *Virulence* 5:775–785. <https://doi.org/10.4161/21505594.2014.960726>
- Mezzaroma E, Abbate A, Toldo S (2021) NLRP3 inflammasome inhibitors in cardiovascular diseases. *Molecules* 26:976. <https://doi.org/10.3390/molecules26040976>
- Mroczynska M, Brillowska-Dąbrowska A (2020) Review on current status of echinocandins use. *Antibiotics* 9:227. <https://doi.org/10.3390/ANTIBIOTICS9050227>
- Murphy SE, Bicanic T (2021) Drug resistance and novel therapeutic approaches in invasive candidiasis. *Front Cell Infect Microbiol* 11:759408. <https://doi.org/10.3389/FCIMB.2021.759408/BIBTEX>
- Nielsen S, White K, Preiss K, Peart D, Gianoulis K, Juel R, Sutton J, McKinney J, Bender J, Pinc G, Bergren K, Gans W, Kelley J, McQuaid M (2021) Growth and antifungal resistance of the pathogenic yeast, *Candida albicans*, in the microgravity environment of the international space station: an aggregate of multiple flight experiences. *Life* 11:283. <https://doi.org/10.3390/LIFE11040283>
- Nobile CJ, Johnson AD (2015) *Candida albicans* biofilms and human disease. *Annu Rev Microbiol* 69:71–92. <https://doi.org/10.1146/annurev-micro-091014-104330>
- Ostrosky-Zeichner L, Casadevall A, Galgiani JN, Odds FC, Rex JH (2010) An insight into the antifungal pipeline: selected new molecules and beyond. *Nat Rev Drug Discov* 9(9):719–727. <https://doi.org/10.1038/nrd3074>
- Parambath S, Dao A, Kim HY, Zawahir S, Alastruey Izquierdo A, Tacconelli E, Govender N, Oladele R, Colombo A, Sorrell T, Ramon-Pardo P, Fusire T, Gigante V, Sati H, Morrissey CO, Alfenaar J-W, Beardsley J (2024) *Candida albicans* —a systematic review to inform the World Health Organization Fungal Priority Pathogens List. *Med Mycol* 62. <https://doi.org/10.1093/mmy/myae045>
- Pathakumari B, Liu W, Wang Q, Kong X, Liang G, Chokkakula S, Pathakamuri V, Nunna V (2024) Comparative evaluation of *Candida* species-specific T-cell immune response in human peripheral blood mononuclear cells. *Biomedicines* 12:1487. <https://doi.org/10.3390/biomedicines12071487>
- Patil S, Majumdar B, Sarode SC, Sarode GS, Awan KH (2018) Oropharyngeal candidosis in HIV-infected patients—an update. *Front Microbiol* 9:356560. <https://doi.org/10.3389/FMICB.2018.00980/BIBTEX>
- Perez-Rodríguez A, Eraso E, Quindós G, Mateo E (2022) Antimicrobial peptides with anti-*Candida* activity. *Int J Mol Sci* 23:9264. <https://doi.org/10.3390/IJMS23169264>
- Picheta N, Piekarz J, Burdan O, Satora M, Tarkowski R, Kułak K (2024) Phytotherapy of vulvovaginal candidiasis: a narrative review. *Int J Mol Sci* 25:3796. <https://doi.org/10.3390/IJMS25073796>
- Pirotta MV, Garland SM (2005) Her choice: dealing with lactobacilli, vaginitis, and antibiotics. *Curr Infect Dis Rep* 7:445–452. <https://doi.org/10.1007/S11908-005-0046-5/METRCS>
- Richardson JP (2022) *Candida albicans*: a major fungal pathogen of humans. *Pathogens* 11:459. <https://doi.org/10.3390/pathogens11040459>
- Richardson JP, Moyes DL (2015) Adaptive immune responses to *Candida albicans* infection. *Virulence* 6:327–337. <https://doi.org/10.1080/21505594.2015.1004977>

- Robinson MJ, Osorio F, Rosas M, Freitas RP, Schweighoffer E, Groß O, Verbeek JS, Ruland J, Tybulewicz V, Brown GD, Moita LF, Taylor PR, Reis e Sousa C (2009) Dectin-2 is a Syk-coupled pattern recognition receptor crucial for Th17 responses to fungal infection. *J Exp Med* 206:2037–2051. <https://doi.org/10.1084/jem.20082818>
- Rogiers O, Frising UC, Kucharíková S, Jabra-Rizk MA, van Loo G, Van Dijk P, Wullaert A (2019) Candidalysin crucially contributes to Nlrp3 inflammasome activation by *Candida albicans* hyphae. *mBio* 10. <https://doi.org/10.1128/mBio.02221-18>
- Rosati D, Bruno M, Jaeger M, ten Oever J, Netea MG (2020) Recurrent vulvovaginal candidiasis: an immunological perspective. *Microorganisms* 8:144. <https://doi.org/10.3390/microorganisms8020144>
- Roselletti E, Perito S, Gabrielli E, Mencacci A, Pericolini E, Sabbatini S, Cassone A, Vecchiarelli A (2017) NLRP3 inflammasome is a key player in human vulvovaginal disease caused by *Candida albicans*. *Sci Rep* 7:17877. <https://doi.org/10.1038/s41598-017-17649-8>
- Roy M, Karhana S, Shamsuzzaman M, Khan MA (2023) Recent drug development and treatments for fungal infections. *Braz J Microbiol* 54:1695–1716. <https://doi.org/10.1007/S42770-023-00999-Z/TABLES/2>
- Sabyasachi Ghosh (2023) Yeast infection treatment market revenue forecast 2024–2034. <https://www.futuremarketinsights.com/reports/yeast-infection-treatment-market>. Accessed 18 Dec 2024
- Sahu SR, Bose S, Singh M, Kumari P, Dutta A, Utkalaja BG, Patel SK, Acharya N (2022) Vaccines against candidiasis: status, challenges and emerging opportunity. *Front Cell Infect Microbiol* 12:1002406. <https://doi.org/10.3389/FCIMB.2022.1002406/BIBTEX>
- Saijo S, Iwakura Y (2011) Dectin-1 and Dectin-2 in innate immunity against fungi. *Int Immunol* 23:467–472. <https://doi.org/10.1093/intimm/dxr046>
- Salazar SB, Simões RS, Pedro NA, Pinheiro MJ, Carvalho MFNN, Mira NP (2020) An overview on conventional and non-conventional therapeutic approaches for the treatment of candidiasis and underlying resistance mechanisms in clinical strains. *J Fungi* 6:23. <https://doi.org/10.3390/jof6010023>
- Santoni G, Boccanera M, Adriani D, Lucciarini R, Amantini C, Morrone S, Cassone A, De Bernardis F (2002) Immune cell-mediated protection against vaginal candidiasis: evidence for a major role of vaginal CD4(+) T cells and possible participation of other local lymphocyte effectors. *Infect Immun* 70:4791–4797. <https://doi.org/10.1128/IAI.70.9.4791-4797.2002>
- Scorzoni L, Fuchs BB, Junqueira JC, Mylonakis E (2021) Current and promising pharmacotherapeutic options for candidiasis. *Expert Opin Pharmacother* 22:887–888. <https://doi.org/10.1080/14656566.2021.1873951>
- Shao T-Y, Haslam DB, Bennett RJ, Way SS (2022) Friendly fungi: symbiosis with commensal *Candida albicans*. *Trends Immunol* 43:706–717. <https://doi.org/10.1016/j.it.2022.07.003>
- Sheppard DC, Filler SG (2015) Host cell invasion by medically important fungi. *Cold Spring Harb Perspect Med* 5. <https://doi.org/10.1101/CSHPERSPECT.A019687>
- Sibeko S, Sanderson M, Moyo S, Botha MH (2024) Role of the epithelium in human papillomavirus and human immunodeficiency virus infections in the female genital tract. *Frontiers in Reproductive Health* 6. <https://doi.org/10.3389/frph.2024.1408198>
- Sobel JD (2007) Vulvovaginal candidosis. *Lancet* 369:1961–1971. [https://doi.org/10.1016/S0140-6736\(07\)60917-9](https://doi.org/10.1016/S0140-6736(07)60917-9)
- Superti F, de Seta F (2020) Warding off recurrent yeast and bacterial vaginal infections: lactoferrin and lactobacilli. *Microorganisms* 8:130. <https://doi.org/10.3390/MICROORGANISMS8010130>
- Swanson KV, Deng M, Ting JP-Y (2019) The NLRP3 inflammasome: molecular activation and regulation to therapeutics. *Nat Rev Immunol* 19:477–489. <https://doi.org/10.1038/s41577-019-0165-0>
- Swidrigall M, LeibundGut-Landmann S (2022) Immunosurveillance of *Candida albicans* commensalism by the adaptive immune system. *Mucosal Immunol* 15:829–836. <https://doi.org/10.1038/s41385-022-00536-5>
- Szymański M, Chmielewska S, Czyżewska U, Malinowska M, Tylicki A (2022) Echinocandins – structure, mechanism of action and use in antifungal therapy. *J Enzyme Inhib Med Chem* 37:876–894. <https://doi.org/10.1080/14756366.2022.2050224>
- Talapko J, Juzbašić M, Matijević T, Pustijanac E, Bekić S, Kotris I, Škrlec I (2021) *Candida albicans*—the virulence factors and clinical manifestations of infection. *J Fungi* 7:79. <https://doi.org/10.3390/jof7020079>
- Tantra T, Rahaman TAA, Nandini CS (2024) Therapeutic role of NLRP3 inflammasome inhibitors against Alzheimer's disease. *Bioorg Chem* 153:107912. <https://doi.org/10.1016/j.bioorg.2024.107912>
- Tavares AH, Bürgel PH, Bocca AL (2015) Turning up the heat: inflammasome activation by fungal pathogens. *PLoS Pathog* 11:e1004948. <https://doi.org/10.1371/journal.ppat.1004948>
- Torrado JJ, Espada R, Ballesteros MP, Torrado-Santiago S (2008) Amphoterin B formulations and drug targeting. *J Pharm Sci* 97:2405–2425. <https://doi.org/10.1002/JPS.21179>
- Tsui C, Kong EF, Jabra-Rizk MA (2016) Pathogenesis of *Candida albicans* biofilm. *Pathog Dis* 74:ftw018. <https://doi.org/10.1093/FEMSPD/FTW018>
- Ture Z, Alp E (2018) Infection control measures to prevent hospital transmission of *Candida*. *Hosp Pract* 46:253–257. <https://doi.org/10.1080/21548331.2018.1510282>
- Uthayakumar D, Sharma J, Wensing L, Shapiro RS (2020) CRISPR-based genetic manipulation of *Candida* species: historical perspectives and current approaches. *Front Genome Ed* 2:606281. <https://doi.org/10.3389/FGED.2020.606281/BIBTEX>
- Valeriano VD, Lahtinen E, Hwang I-C, Zhang Y, Du J, Schuppe-Koistinen I (2024) Vaginal dysbiosis and the potential of vaginal microbiome-directed therapeutics. *Front Microbiomes* 3:1363089. <https://doi.org/10.3389/FRMBI.2024.1363089>
- van de Veerdonk FL, Netea MG (2010) T-cell subsets and antifungal host defenses. *Curr Fungal Infect Rep* 4:238–243. <https://doi.org/10.1007/s12281-010-0034-6>
- van de Veerdonk FL, Netea MG, Joosten LA, van der Meer JWM, Kullberg BJ (2010) Novel strategies for the prevention and treatment of *Candida* infections: the potential of immunotherapy. *FEMS Microbiol Rev* 34:1063–1075. <https://doi.org/10.1111/J.1574-6976.2010.00232.X>
- Ventolini G, Baggish MS (2006) Recurrent Vulvovaginal Candidiasis. *Clin Microbiol News* 28:93–95. <https://doi.org/10.1016/J.CLINMICNEWS.2006.05.004>
- Wächtler B, Citiulo F, Jablonowski N, Förster S, Dalle F, Schaller M, Wilson D, Hube B (2012) *Candida albicans*-epithelial interactions: dissecting the roles of active penetration, induced endocytosis and host factors on the infection process. *PLoS ONE* 7:36952. <https://doi.org/10.1371/JOURNAL.PONE.0036952>
- Wall G, Lopez-Ribot JL (2020) Current antimycotics, new prospects, and future approaches to antifungal therapy. *Antibiotics* 9:445. <https://doi.org/10.3390/antibiotics9080445>
- Wu Y, Hu S, Wu C, Gu F, Yang Y (2022) Probiotics: potential novel therapeutics against fungal infections. *Front Cell Infect Microbiol* 11:793419. <https://doi.org/10.3389/FCIMB.2021.793419/BIBTEX>
- Xu J, Pickard JM, Núñez G (2024) FDA-approved disulfiram inhibits the NLRP3 inflammasome by regulating NLRP3 palmitoylation. *Cell Rep* 43:114609. <https://doi.org/10.1016/j.celrep.2024.114609>
- Zahid A, Li B, Kombe AJK, Jin T, Tao J (2019) Pharmacological inhibitors of the NLRP3 inflammasome. *Front Immunol* 10. <https://doi.org/10.3389/fimmu.2019.02538>

Zeise KD, Woods RJ, Huffnagle GB (2021) Interplay between *Candida albicans* and lactic acid bacteria in the gastrointestinal tract: impact on colonization resistance, microbial carriage, opportunistic infection, and host immunity. *Clin Microbiol Rev* 34. <https://doi.org/10.1128/CMR.00323-20/ASSET/01B0A05B-B683-4632-9DD8-F59735030961/ASSETS/IMAGES/LARGE/CMR.00323-20-F005.JPG>

Zhao S, Shang A, Guo M, Shen L, Han Y, Huang X (2022) The advances in the regulation of immune microenvironment by

Candida albicans and macrophage cross-talk. *Front Microbiol* 13. <https://doi.org/10.3389/fmicb.2022.1029966>

Zheng N-X, Wang Y, Hu D-D, Yan L, Jiang Y-Y (2015) The role of pattern recognition receptors in the innate recognition of *Candida albicans*. *Virulence* 6:347–361. <https://doi.org/10.1080/21505594.2015.1014270>

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