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Authors	Tânia Ribeiro, Margarida Maia, António Fonseca, Bianca Marques, Cristina Caleja, Ana Rosa, Rui Martins, André Almeida, Maria João Mota, Tiago Aires, Cristina Rocha, José A. Teixeira, Ana Rita Cabrita, Lillian Barros, Manuela Pintado

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8 Tânia Bragança Ribeiro^{a*}, Margarida R. G. Maia^{b,c}, António J. M.
9 Fonseca^b, Bianca Marques^d, Cristina Caleja^{e,f}, Ana Rosa^g, Rui Martins^h,
10 André Almeida^h, Maria J. Motaⁱ, Tiago Airesⁱ, Cristina M. R. Rocha^{d,j}, José
11 A. Teixeira^{d,j}, Ana R. J. Cabrita^b, Lillian Barros^{e,f}, Manuela Pintado^a
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14
15
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18

19 *^aUniversidade Católica Portuguesa, CBQF- Centro de Biotecnologia e Química Fina -*
20 *Laboratório Associado, Escola Superior de Biotecnologia, Rua Diogo Botelho 1327,*
21 *4169-005 Porto, Portugal; ^bREQUIMTE, LAQV, ICBAS, Instituto de Ciências*
22 *Biomédicas Abel Salazar, Universidade do Porto, Rua de Jorge Viterbo Ferreira 228,*
23 *4050-313 Porto, Portugal; ^cLEAF, TERRA, Instituto Superior de Agronomia,*
24 *Universidade de Lisboa, Tapada da Ajuda, 1349-017 Lisboa, Portugal, ^dCEB - Centre*
25 *of Biological Engineering, University of Minho, 4710-057 Braga, Portugal; ^eCentro de*
26 *Investigação de Montanha (CIMO), Instituto Politécnico de Bragança, Campus de*
27 *Santa Apolónia, 5300-253 Bragança, Portugal; ^fLaboratório Associado para a*
28 *Sustentabilidade e Tecnologia em Regiões de Montanha (SusTEC), Instituto Politécnico*
29 *de Bragança, Campus de Santa Apolónia, 5300-253 Bragança, Portugal; ^gSEBOL,*
30 *Comércio e Indústria de Sebo, S.A, Rua Padre Adriano n.º 61, Olivais do Machio 2660-*
31 *119, St.º Antão do Tojal, Portugal; ^hIndústria Transformadora de Subprodutos, S.A.,*
32 *Herdade da Palmeira - Olheiros do Meio - São José da Lamarosa Agolada 2100-011*
33 *Coruche, Portugal; ⁱSORGAL, Sociedade de Óleos e Rações, S.A., Estrada Nacional*
34 *109, Lugar da Pardala, 3880-728 S. João de Ovar, Portugal; ^jLABBELS - Associate*
35 *Laboratory, Braga, Portugal*
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50 *corresponding author (e-mail: tribeiro@ucp.pt)
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A comprehensive review of fish protein hydrolysates targeting pet food formulations

The fish industry generates significant amounts of fish by- and co-products (FBCPs) annually, projected to reach 160.8 million tonnes by 2030. This growth highlights the urgent need for sustainable FBCP management and an opportunity to improve the sector's environmental sustainability. Fish protein hydrolysates (FBCPHs) and bioactive peptides (BPs) derived from these FBCPs are gaining recognition in the pet food sector for their nutritional and bioactive benefits. FBCPHs, primarily sourced from category 3 by-products unsuitable for human consumption, could significantly enhance the economic viability of both industries. This review analyzes production processes, highlights the benefits and challenges of enzymatic hydrolysis, and reviews emerging technologies such as subcritical water hydrolysis (SWH), promising sustainable alternatives by enhancing extraction efficiency and reducing energy consumption. The review explores FBCPHs' applications in pet food, focusing on beneficial biological activities (e.g., antioxidant, prebiotic, neuroprotective). Findings show FBCPHs have significant potential in pet food formulations, providing palatability, hypoallergenic benefits, and addressing health concerns like gastrointestinal disorders and stress-related behaviors. However, further research is required to optimize production processes, scale industrial applications, and ensure regulatory compliance. In conclusion, FBCPHs present a valuable solution for promoting sustainability, improving pet nutrition, and supporting the circular economy.

1. Introduction

The food industry generates high amounts of animal-based waste annually. In 2022, the fish and aquaculture industry produced an estimated 185.4 million tons, up to 35 % lost or wasted annually. This waste includes heads, skins, trimmings, fins, bones, viscera, scales, and shells from crustaceans, which account for approximately 60% of the total fish processing volume. Additionally, by-catches and undersized fish are often rejected, further contributing to waste^[1-3]. Most of these were traditionally discarded as

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4 wastes with high costs and significant environmental impact, mostly due to their high
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6 organic load and rapid deterioration, especially when containing viscera [1-3].
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9 Despite being underutilized and generally low-valued, the use of fish by- and co-
10 products (FBCPs) has been increasing for the production of food and non-food
11 products, such as fishmeal, fish oil, and fertilizers. These food products, rich in proteins
12 and oil, have begun to account for a growing share of fish-derived resources, with
13 FBCPs now representing 34% of fishmeal and 53% of fish oil production [1]. While the
14 recovery of other by-products, like chitin and chitosan, has been explored, their uptake
15 by the industry remains limited. New sustainable extraction and conversion routes are
16 being researched to develop industrially feasible and high-quality polymers, as
17 conventional methods are energy-intensive and require strong acids, bases, and solvents.
18 Potential applications for these polymers include agrochemicals, water treatment agents,
19 packaging, coatings, biomedical devices, and dietary supplements[4]. Marine-sourced
20 collagen and gelatin are other valuable by-products; although they are still underutilized
21 compared to terrestrial sources, they are gaining attention in food, cosmetic, and
22 biomedical applications. Their future use as food ingredients, pharmaceuticals, and
23 biomedical devices is also promising due to their texturizing, film-forming, and
24 biocompatibility properties [5].
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42 The European Commission's goal of achieving zero waste by 2030, combined
43 with the growing global population and concerns over food security, is forcing the fish
44 industry toward more circular, sustainable solutions. The valorization of FBCPs,
45 particularly proteins, supports these objectives by meeting nutritional demands while
46 reducing waste and environmental impact. Adopting a circular bioeconomy approach
47 improves resource efficiency by transforming waste into high-value products [2, 3, 6].
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4 In the pet food industry, the European Pet Food Industry Federation (FEDIAF)
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6 identified protein as a key ingredient essential for growth, muscle maintenance,
7
8 digestion, and energy production [6]. While the pet food industry already utilizes a wide
9
10 range of by- and co-products (BCPs) as protein sources, including fish, meat, bone
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12 meal, poultry, and poultry by-product meal [6, 7], there is a trend towards replacing
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14 and/or enriching these rendered meals with novel functional ingredients [8]. Pet owners'
15
16 increasing concern for their animals' health and well-being has driven research toward
17
18 new ingredients, such as plant and algae extracts, protein isolates, and hydrolysates
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20 (e.g., from soy or fish). These functional ingredients offer additional health benefits
21
22 such as improved immune function, satiety, and digestive health while also addressing
23
24 specific health concerns such as halitosis, neuroprotection, or anti-carcinogenic
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26 activities. They also provide hypoallergenic properties, making them suitable for pets
27
28 with sensitivities [9].
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31 Protein hydrolysates, like soy-based hydrolysates, are already used in the pet
32
33 food industry to produce hypoallergenic formulations and improve palatability,
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35 although scientific data on their broader use remains limited [2, 3]. Companies like BRF
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37 Ingredients offer commercially available hydrolyzed proteins, such as BioActio, for
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39 functional feed and pet food formulations. Among these, fish-based hydrolysates, fish
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41 by- and co-products hydrolysates (FBCPHs), and their bioactive peptides (BPs) have
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43 attracted considerable attention for their nutritional benefits and biological properties [2,
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45 3, 6, 10].
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48 Numerous studies have reported that fish-based hydrolysates and purified BPs
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50 exhibit a variety of biological activities, namely antimicrobial, antidiabetic,
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52 antihypertensive, antioxidant, neuroprotective, anti-inflammatory, and
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54 immunomodulatory [6, 11, 12]. These properties make FBCPHs promising ingredients for
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4 functional pet food. Research by Folador et al. [11] demonstrated the potential of
5
6 FBCPHs in canned foods and dry extruded kibbles. Other studies have reported that
7
8 incorporating fish-based hydrolysates may improve digestion and enhance palatability
9
10 in pet food [6, 11–13]. For example, two recent studies found that incorporating FBCPHs
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12 into commercial dog food formulations was well accepted by beagle dogs and did not
13
14 negatively impact food intake, digestibility, or fecal characteristics [14, 15]. However,
15
16 despite these advantages, data on their application in commercial pet food and their
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18 effects on companion animals remain limited.
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21 Bioactive peptides (BPs) are inactive within their parent protein structure and
22
23 require proteolysis to be released. This process produces fish-based hydrolysates [2, 3].
24
25 Most fish BPs consist of 2 to 20 amino acid sequences with a molecular weight lower
26
27 than 6000 Da [2, 10]. Several methods have been used to hydrolyze protein and produce
28
29 fish-based protein hydrolysates and BPs, including chemical hydrolysis, microbial
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31 fermentation, and enzymatic hydrolysis. Enzymatic hydrolysis is preferred due to its
32
33 specificity and control [2, 3]. While chemical hydrolysis is cheaper, it is unspecific and
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35 difficult to control, and the extreme alkali or acidic conditions can lead to the
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37 destruction of certain amino acids as well as to the formation of D - amino acids and
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39 toxic substances, leading to reduced nutritional quality [3]. In contrast, enzymatic
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41 hydrolysis occurs under moderate and controllable reaction conditions, high substrate
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43 specificity, and minimal toxic residues, though the high cost of enzymes limits its
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45 industrial-scale use [2, 3]. Fermentation offers a more cost-effective alternative but
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47 introduces additional complexity in production [2, 3].
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51 This review aims to consolidate the available knowledge on the sustainable upcycling of
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53 fish waste and FBCPs into valuable FBCPHs, highlighting their potential as functional
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55 ingredients in pet food formulations. By focusing on their biological properties,
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4 production methods, and application in the pet food industry, this review seeks to
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6 present new insights that can help researchers and the industry overcome the challenges
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8 of producing high-value ingredients from undervalorized biomass, such as FBCPs.
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10 Furthermore, this approach supports the sustainable management of protein resources
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12 and aligns with the broader goals of reducing waste and environmental impact within
13
14 the food production system [2]. Therefore, the following sections include (1) Describing
15
16 the FBCPs generated by the fish processing industry, (2) Exploring various methods
17
18 used for producing and purifying FBCPHs and bioactive peptides, including innovative
19
20 technologies, (3) Analyzing the reported biological properties of these hydrolysates and
21
22 peptides; (4) Highlighting the potential of FBCPHs and purified peptides for pet food
23
24 formulations and (5) Discussing pet food demands, trends, and relevant EU legislation
25
26 regarding functional ingredients and fish hydrolysate application in pet food.
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29 -----Figure 1-----
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32 The relevant scientific data was collected by searching the Web of Science,
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34 Scopus, and Google Scholar using keywords like "fish hydrolysates," "fish by-
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36 products," "fish wastes valorization," "valorization of seafood," "fish hydrolysates in pet
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38 food," "fish protein hydrolysate," "hydrolyzed diets," and "upcycled foods." Information
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40 on pet food legislation was retrieved from FEDIAF and Global Alliance of Pet Food
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42 Associations (GAPFA) resources.
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49 2. Fish by-products and co-products

50 Over the past decades, global fish production and consumption have
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52 significantly increased. Fish production increased from 138 million tonnes in 2000 to
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54 179 million tonnes by 2019, with a corresponding rise in per capita fish consumption
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4 from 15.1 kg to 20.5 kg^[16]. This trend is expected to continue, with FAO predicting a
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6 13% increase in fish production by 2030, reaching 201 million tons^[16]. Consequently,
7
8 the fishing sector generates an estimated 20 million tons of FBCPs annually^[17].
9

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11 Around 70% of the fish consumed undergoes processing before reaching the
12 consumer. Depending on factors like type of processing, fish species, and geographical
13 area, fish processing generates FBCPs ranging from 20% to 80% of the total fish weight
14
15^[17]. These FBCPs are rich in valuable nutrients such as proteins (49 to 57% dry weight,
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17 e.g., collagen and gelatin and derived peptides), minerals (22 to 30% dry weight of ash,
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19 e.g., calcium and phosphorus), lipids (7 to 19% dry weight, e.g., eicosapentaenoic acid
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21 (EPA) and docosahexaenoic acid (DHA)), and enzymes (pepsin, trypsin, collagenase,
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23 and chymotrypsin)^[2, 3, 18].
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27 FBCPs are derived from various fish parts, each offering distinct bioactive
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29 compounds. For example, muscle contains protein, while viscera are rich in oil,
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31 enzymes, and protein^[19, 20]. Bones contain calcium and chondroitin sulfate^[19], while
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33 heads provide protein, oil, and chondroitin sulfate^[19]. Scales are a source of collagen,
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35 calcium, and gelatin^[19], and skin can provide collagen, gelatin, and protein^[19]. Fins can
36
37 be used for chondroitin sulfate production^[19] (Figure 1). These compositional
38
39 differences and specificities allow for the recovery of various valuable products
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41 depending on the FBCPs processed. For instance, fish skin can be enzymatically
42
43 hydrolyzed to produce hydrolysates rich in collagen peptides^[21], and fish viscera can be
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45 used to recover omega-3 fatty acids rich in EPA and DHA^[22] or even digestive
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47 enzymes^[20]. Overall, all FBCPs are suitable to be valorized by enzymatic hydrolysis, a
48
49 versatile technique that allows the production of FBCPHs by recovering the protein
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51 present in all types of FBPCs or even whole fish^[19, 23] while also recovering fatty acids
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53 or minerals present in FBCPs^[17, 19, 23].
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-----Figure 2-----

BCPs hold significant potential for higher-value applications in food and feed, including pet food. However, international regulations strictly govern the treatment and handling of animal BCPs. In this context, co-products refer to residual raw materials remaining from the slaughtering process that retain a quality suitable for human consumption and can be employed in food production. Conversely, materials considered unsuitable for human consumption due to commercial, safety, and/or regulatory considerations are designated by-products. The European Union categorizes non-edible animal by-products into three categories based on their potential risk (Regulation (EC) 142/2011) [24]. Category 1 (highest risk) is unsuitable for food or feed, while Category 2 can be repurposed for non-food applications like fuel. Category 3 (lowest risk) includes materials safe for human consumption but excluded for commercial or cultural reasons, such as packaged fish with minor damage or expired shelf-life.

Regulations regarding processing BCPs as animal feed have forced producers to adapt and introduce novel by-product processing strategies. Where BCP producers once freely disposed of these products, European Regulations (Regulation (EC) 142/2011 [24]) now mandate specific, detailed processing methods [24]. Despite the permitted applications of processed BCPs under European Regulations, the economic costs of implementing these techniques have increased. Therefore, there is a growing demand for novel methods that recover more valuable fractions from BCPs, increasing their economic value and offsetting processing costs.

Numerous investigations have shown that valorizing animal BCPs (category 3) for animal feed, including pet food, is an environmentally beneficial approach [25]. The

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4 European Union generates between 88 and 129 million tons of food waste annually,
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6 emitting 170 million tonnes of CO₂, with half of the waste generated in the production
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8 chain [26]. Reducing this waste is critical to lowering CO₂ emissions and mitigating
9
10 global warming. Recycling BCPs can significantly reduce CO₂ emissions in animal and
11
12 pet food production [25, 26]. For example, a study has shown that incorporating recycled
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14 meat (e.g., beef, pork, and poultry) from packaged food waste into pet food has
15
16 significantly reduced the industry's environmental footprint [25].

17
18 Similarly, life cycle assessment (LCA) studies on fish meal production plants
19
20 have shown that valorizing FBCPs into feed and pet food ingredients can decrease the
21
22 environmental impact of fish processing [27–29]. Laso and colleagues studied the
23
24 environmental management of FBCPs in the canned anchovy industry using LCA
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26 methods. Their analysis demonstrated that valorizing anchovy heads and bones for
27
28 fishmeal and oil production was significantly more environmentally advantageous than
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30 incineration or landfilling, mainly due to the avoided environmental burdens from
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32 traditional processes such as fishing fresh anchovy for fishmeal production. According
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34 to the authors, this valorization approach resulted in the emission of 37,8 kg of CO₂
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36 equivalents per ton of anchovy heads and bones, while landfilling led to emissions of
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38 810 kg of CO₂ equivalents (with gas recovery) and up to 830 kg without gas recovery.
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40 Although the study did not profoundly approach the financial aspects, the authors noted
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42 that valorizing FBCPs exhibits clear economic advantages over landfilling or
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44 incineration, as it produces a final product with commercial value and demand [28]. In
45
46 another study, Maiolo and colleagues assessed the supply chain of Italian rainbow trout
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48 (*Oncorhynchus mykiss*) and concluded that valorizing FBCPs for pet food production
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50 resulted in slightly higher emissions than incineration. However, the authors stated that
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52 this result was primarily due to the reliance on fossil fuels for energy, and using
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4 renewable energy could lower emissions significantly, offering a solid alternative to
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6 reduce the industry's environmental footprint [29]. Furthermore, Vázquez and colleagues
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8 explored the application of enzymatic treatments to FBCPs, finding that these
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10 treatments produce high-quality protein products with excellent digestibility and
11
12 bioactive properties, such as antioxidant and antihypertensive activities. Although
13
14 preliminary assessments indicate that enzymatic processing is energy-intensive, this
15
16 process presents a more sustainable, scalable, and flexible option compared to
17
18 traditional methods [30].

21 FBCPHs are particularly interesting for pet food due to several factors. They
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23 provide a sustainable protein source, rich in easily absorbable amino acids in the small
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25 intestine [13]. Additionally, FBCPHs possess hypoallergenic properties [12], making them
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27 suitable for pets with sensitivities. Furthermore, they may act as palatability enhancers
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29 [11], improving the taste and acceptance of pet food. Besides these benefits, FBCPHs
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31 also exhibit antioxidant and antihypertensive activities [6], enhancing their nutritional
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33 and functional value for pets. These combined properties make FBCPHs an interesting
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35 option for pet food formulation. However, careful consideration must be given to the
36
37 processing methods, source materials, and energy consumption involved in FBCPH
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39 production to ensure sustainability and cost-effectiveness [29, 31].

42 Overall, alternative methods for FBCPs valorization may be promising solutions
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44 to reduce the environmental impact of the fish processing industry by lowering the CO₂
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46 emission and producing valuable products such as fishmeal and FBCPHs. Besides
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48 environmental benefits, these products increase economic value, while landfilling or
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50 incineration represents an expense for producers. However, these methods still need
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52 improvement, particularly regarding energy efficiency. However, addressing energy
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54 efficiency challenges and performing further optimization studies are essential to fully
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4 exploit their potential benefits and accelerate their use in industries such as the pet food
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11 3. Peptides production from fish by-products

14 3.1. Enzymatic hydrolysis

16 FBCPHs can be produced by chemical hydrolysis, physical treatment, microbial
17 fermentation, or enzymatic hydrolysis. The choice of hydrolysis method significantly
18 influences the purity, yield, and overall cost of the process. Chemical hydrolysis is more
19 cost-effective, using acids or bases to break down proteins. However, this process is less
20 specific, resulting in the degradation of sensitive amino acids (tryptophan degradation)
21 and the formation of side products, such as salts, which reduce the purity and bioactivity
22 of the FBCPHs [6, 32]. Fermentation presents a more sustainable and economical
23 alternative, utilizing microbial enzymes to generate bioactive FBCPHs. However,
24 lower-purity FBCPHs were typically obtained due to the broader range of molecular
25 weights in the peptides generated [6, 32]. In contrast, enzymatic hydrolysis allows more
26 control over protein breakdown, producing peptides with higher bioactivity and
27 specificity. This method employs mild reaction conditions, protecting sensitive amino
28 acids' integrity. However, it is also more expensive due to the cost of enzymes and the
29 complexities involved in scaling up the process [6, 32]. Preliminary assessments suggest
30 valorizing FBCPs via enzymatic hydrolysis or fermentation may offer environmental
31 advantages compared to conventional fish meal production [30].
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50 Among the available methods, enzymatic hydrolysis is the most widely used due
51 to its many advantages. The selectivity of enzymes allows the controlled and specific
52 release of peptides, and using mild temperature and pH conditions results in
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4 hydrolysates with higher nutritional quality (e.g., less loss of amino acids) [33, 34]. In
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6 enzymatic hydrolysis, a proteolytic enzyme, or an enzyme mixture, breaks specific
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8 peptide bonds (H^+) of proteins, producing smaller (3–20 AA residues) and more water-
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10 soluble peptides (with increased ionizable groups) than the intact proteins [34, 35].
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12 The enzymatic hydrolysis process of FBCPs typically involves four steps: (1)
13
14 Homogenization of FBCPs through grinding and water addition, which ensures
15
16 thorough mixing and enzyme access while adjusting the protein concentration to 8–
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18 12%; (2) Establishment of hydrolysis parameters; such as temperature, pH, enzyme-to-
19
20 substrate ratio, water-to-substrate ratio, and processing time, followed by enzyme
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22 addition; (3) Enzyme inactivation post-hydrolysis, usually through heating (85–95°C for
23
24 5–20 min) or pH adjustment; (4) Separation of the hydrolysate (liquid fraction) from
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26 solids (unhydrolyzed proteins, bones, skin, scales, and others) and oil fraction via three-
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28 phase decanter, filtration, or centrifugation. The hydrolysate can then be concentrated
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30 through evaporation or dried using a spray or freeze-dryer [3, 34–37]. A schematic
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32 representation of the enzymatic hydrolysis process of FBCPs is illustrated in Figure 3.
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36 -----Figure 3-----
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40 Although enzymatic hydrolysis may appear straightforward, many factors must
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42 be considered to produce high-yield, high-quality FBCPHs. These factors include
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44 enzyme selection and extraction conditions, such as water-to-substrate ratio, pH,
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46 temperature, and processing time. Selecting an appropriate enzyme is crucial, as it
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48 determines the cleavage pattern of peptide bonds [33, 34]. Enzymes can be sourced from
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50 vegetables (papain and bromelain), animals (pepsin and trypsin), bacteria (alcalase and
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52 neutrase), or fungi (fungal protease), and they target different peptide bonds based on
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54 their endo- or exopeptidase activities [32–34]. The choice of enzyme should be tailored to
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4 the protein source and the desired peptide size distribution [34]. Other important
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6 considerations include enzyme concentration, pH, and temperature. Increasing enzyme
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8 concentration speeds up hydrolysis but raises production costs [32]. Enzyme efficiency
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10 also depends on maintaining optimal pH and temperature ranges, as deviations can
11
12 reduce hydrolysis efficiency and peptide yield [32, 34]. However, adjusting the optimum
13
14 pH may cause high salt levels in the final hydrolysate, negatively impacting the
15
16 FBCPH's nutritional value [34]. Similarly, while higher temperatures increase enzyme
17
18 activity, temperatures outside the optimal range can denature enzymes and slow down
19
20 reactions, reducing efficiency [6]. Other important factors include the hydrolysis time
21
22 and the water-to-substrate ratio. Longer hydrolysis times can increase peptide yield, but
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24 excessive time may cause over-hydrolysis, reducing the bioactivity [32]. A proper water-
25
26 to-substrate ratio is essential to prevent product inhibition and maximize yield, though
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28 adding more water increases drying costs [34, 38].

31 Processing FBCPs presents additional challenges due to the complexity and
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33 heterogeneity of the material. Therefore, controlling their proximate composition (e.g.,
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35 fat, protein, moisture, and ash) and amino acid composition is crucial for producing
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37 quality, stable FBCPHs [34]. For example, high lipid and solid content (e.g., skin, bones,
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39 scales) can lead to significant lipid oxidation and negatively affect enzymatic hydrolysis
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41 [36, 37]. Suitable antioxidants can be added, or lipids can be removed before hydrolysis to
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43 prevent lipid oxidation [36]. Mechanical deboning can address the removal of solid
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45 components. Autoclaving and hydrothermal (85–135 °C for 15–120 minutes) pre-
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47 treatments have been effective in softening hard FBCP components such as heads [39]
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49 and scales [40]. Other pre-treatments, like microwaves, offer benefits such as shorter
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51 hydrolysis times, higher antioxidant activity, and lower immunochemical reactivity [41].
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4 Lower pH values and higher temperatures also enhance oxidation and should be avoided
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6 by selecting the proper enzyme [34].
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10 Regarding control parameters, the extent of protein cleavage is a primary
11 consideration. A standard method for monitoring enzymatic protein hydrolysis is
12 measuring the degree of hydrolysis (%DH), i.e., the percentage of cleaved peptide
13 bonds. A higher %DH typically means higher protein recovery [34, 37]. The MW
14 distribution of peptides is another important parameter for ensuring protein hydrolysate
15 quality, directly measured in the final hydrolysate rather than relative to the starting
16 material, as with %DH. However, both methods are labor-intensive, limiting their
17 industrial use. Recently, Fourier-transform infrared (FTIR) spectra have shown promise
18 in monitoring enzymatic protein hydrolysates by predicting the degree of hydrolysis and
19 the average MW of protein hydrolysates [42].
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30 31 **3.2. Emergent technologies**

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33 As mentioned above, different methods can release BPs, including enzymatic
34 hydrolysis, chemical hydrolysis, and fermentation. However, these methods have some
35 disadvantages, including long processing times, high concentration of solvents, high
36 energy consumption, and/or high production costs (e.g., enzymes) [43]. Moreover,
37 extreme conditions, including high temperatures, long reaction times, and/or high
38 alkalinity/acidity, can affect or alter the functional properties of potential bioactive
39 compounds.
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47 To address some of these problems, innovative processing technologies have
48 emerged and been exploited for producing FBCPHs and extracting peptides [43, 44],
49 aiming at faster, more efficient, and highly selective processes with decreased
50 consumptions (solvents/chemicals and energy). These technologies include subcritical
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4 water hydrolysis (SWH), pulsed electric fields (PEF), ultrasound-assisted extraction
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6 (UAE), microwave-assisted extraction (MAE), and supercritical fluid extraction (SFE)
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8 [44].
9

11 3.2.1. Subcritical Water Hydrolysis

12 Subcritical Water Hydrolysis (SWH) is an extraction and hydrolysis method that
13
14 uses subcritical water (SCW) to extract and hydrolyze target compounds or fractions
15
16 from several matrices. SCW is defined as water under a range of pressures and
17
18 temperatures (i.e., ranging from 100 °C at 0.1 MPa to 374.5 °C at 22.06 MPa), above its
19
20 boiling temperature at the selected pressure and below its critical point, with pressure
21
22 high enough to maintain its liquid state. Under subcritical conditions, the
23
24 physicochemical features of water change, namely: (i) the ionic product increases,
25
26 which allows water to act as an acid or base catalyst (significant, for instance, in
27
28 hydrolytic reactions); and (ii) the dielectric constant decreases, which allows water to
29
30 behave as a less polar solvent [43]. Furthermore, due to the high temperature applied,
31
32 viscosity decreases, solubility of some target compounds increases, and reaction rates
33
34 increase, leading to faster and more efficient processes.
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41 -----Figure 4-----
42
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44

45 SWH properties are particularly relevant when aiming to hydrolyze proteins and
46
47 produce peptides. This is because, when high pressure and high temperature are applied
48
49 separately to proteins, they experience denaturation, i.e., the hydrogen bonds are
50
51 broken, and proteins loss their quaternary (if present), tertiary, and even secondary
52
53 structures [43, 45]. In fact, it is known that when applying high hydrostatic pressure at
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4 room temperature or high temperature under atmospheric pressure, the denaturation
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6 process of proteins occurs without altering their molecular size [45]. The protein's
7
8 hydrolysis only occurs when higher temperatures are only possible when combined with
9
10 high pressures, which happens when SWH is used [43, 45]. Once the SCW is present,
11
12 proteins break down into peptides, amino acids, and organic acids, depending on the
13
14 reaction's extensiveness [43, 45]. The main parameters influencing the extension of
15
16 hydrolysis are temperature, pressure, and reaction time. Up to now, the primary
17
18 limitation of using this technology for protein hydrolysis is the lack of complete
19
20 understanding regarding protein cleavage specificity and how it can be controlled or
21
22 predicted [43].

23
24
25 The main applications of SWH are related to the extraction of bioactive
26
27 compounds (polyphenols, flavonoids, and anthocyanins) and hydrolysis of biopolymers
28
29 (lignin, polysaccharides, and protein) from several food matrices. In particular, SWH
30
31 has been exploited for protein extraction and hydrolysis in a wide range of food
32
33 processing wastes from both vegetable and animal sources, including deoiled rice bran,
34
35 soy pulp, onion waste, bean dregs, poultry waste, and other meat by-products and, more
36
37 specifically, FBCPs/wastes [45].

38
39
40 Regarding the use of SWH to extract and hydrolyze protein from fish wastes,
41
42 conditions tested include temperatures ranging from 110 to 300 °C and times ranging
43
44 from five minutes to six hours. These hydrolysates displayed important biological
45
46 activities, including antioxidant, antimicrobial, antiproliferative, and anti-hypertensive
47
48 [43, 46–53]. Examples of the use of SWH to obtain protein-enriched fish hydrolysates and
49
50 bioactive peptides are described below.

51
52
53 Hao et al.^[46] used abalone viscera. Several temperatures (110 to 230 °C) during
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55 1 h were tested. The results showed that 170 °C was the temperature with the best
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4 extraction yields. Hydrolysates' protein content increased gradually with temperatures
5
6 up to 200 °C, ranging from 57 to 69%. The peptide profile of hydrolysates showed that
7
8 SWH at low and medium temperatures (110 – 170 °C) produced peptides with MW >
9
10 5000 Da, but for SWH at high temperatures (200 – 230 °C), the peptides were not
11
12 produced. Nevertheless, most peptides produced were between 1000 and 5000 Da and
13
14 180 and 1000 Da, respectively. All hydrolysates showed antioxidant activity by
15
16 scavenging free radicals, inhibiting lipid oxidation, and exerting reducing power.
17
18 Hydrophobic AAs, such as phenylalanine, leucine, methionine, and tyrosine, were
19
20 correlated with the bioactive features of hydrolysates produced.
21

22
23 **Melgosa et al.^[47] applied SWH at sardine and deoiled sardine wastes, aiming to**
24
25 **produce fish protein hydrolysates with bioactive properties. Results showed that prior**
26
27 **defatting by supercritical CO₂ of sardine BCPs increased protein yields in hydrolysates.**
28
29 **The bioactive properties of fish hydrolysates obtained were affected by temperature.**

30
31 Hydrolysates exhibited antioxidant activity by scavenging DPPH radical and presented
32
33 antiproliferative effects against HT-29 adenocarcinoma cells, the 250 °C temperature
34
35 producing the most bioactive hydrolysate.
36

37
38 Another example of SWH application in fish proteins was reported by Ahmed
39
40 and Chun ^[48], who used tuna skin and collagen previously extracted from tuna skin by
41
42 supercritical CO₂ as raw materials. SWH was performed using a ratio sample: water of
43
44 1:200 and 1:50 for collagen and skin, respectively. The time of SWH was 5 min, and
45
46 temperatures ranged from 120 to 300 °C, with a stirring rate of 150 rpm. The maximum
47
48 %DH was achieved at 250 °C for both raw materials. Hydrolysates exhibited
49
50 antioxidant activity by scavenging radicals, chelating metals, and exhibiting reducing
51
52 power, the best antioxidant features found in hydrolysates produced at 280 °C.
53

54
55 Hydrolysates also exhibited antimicrobial activities against *Staphylococcus aureus*,
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4 *Pseudomonas aeruginosa*, and *Bacillus cereus*. Functional properties of hydrolysates
5
6 vary with temperature and protein source, collagen-derivative hydrolysates being higher
7
8 than tuna skin-derivative hydrolysates. These functional properties arose from small
9
10 peptides (MW < 600 Da) and free AAs.

11
12
13 Other authors have also investigated the SWH of fish wastes from different
14
15 origins, such as shrimp [49], blue mussel [50], mackerel [51], oyster [52], and squid viscera
16
17 [53], and reported antioxidant and anti-hypertensive activity of hydrolysates obtained.

18
19 The advantages of these technologies include their sustainability,
20
21 reproducibility, and low toxicity when compared with other organic solvents and
22
23 chemical extraction methodologies, as well as the lower extraction times and costs when
24
25 compared with enzymatic hydrolysis [45]. The main drawbacks are the degradation of
26
27 some heat-sensitive compounds, the lack of information about cleavage specificity, and
28
29 specific equipment costs. So, additional studies are needed to implement this technology
30
31 in the industry, including an evaluation of energy and economic costs. Nevertheless,
32
33 SWH has been proven to be an effective and green technology for protein hydrolysis
34
35 and valorization of fish wastes.

3.2.2. Pulsed Electric Fields

36
37
38 Pulsed Electric field (PEF) is a non-thermal technology that submits a matrix to
39
40 high-voltage electrical pulses during short periods. Further, applying PEF disturbs cell
41
42 membranes' structure and facilitates the extraction of entrapped bioactive compounds,
43
44 with minimal changes in product sensorial and nutritional properties [54-56]. In fact,
45
46 when a cell membrane is placed in an electric field, the electric potential causes the
47
48 separation of membrane molecules based on their dipole nature and charge within the
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4 membrane. Once the transmembrane potential reaches around 1 V, there is a repulsion
5
6 between the charged molecules, creating pores in the membrane's weak spots
7
8 dramatically increasing permeability [57]. The effectiveness of extraction processes is
9
10 greatly influenced by the temperature of treatment, pulse shape, amplitude, frequency,
11
12 intensity, and duration of the electric pulses [57, 58].
13

14 PEF technology has been widely used in the food industry for dehydration,
15
16 sterilization, and preservation [59, 60]. PEF technology provides various benefits
17
18 compared to traditional pasteurization methods in the food sector, including prolonging
19
20 shelf-life, maintaining nutrients, preserving quality, and being cost-effective [60]. More
21
22 recently, this technology has been exploited to extract bioactive compounds (such as
23
24 polyphenols, anthocyanins, and carotenoids) from plants and fruits and to extract and
25
26 hydrolyze proteins from meat wastes and, more specifically, fish wastes [54, 57, 58, 61].
27
28 Though it was demonstrated that PEF treatment could also cause protein hydrolysis, it
29
30 has been mainly used as a pre-treatment in enzymatic hydrolysis processes to extract
31
32 and hydrolyze protein from fish wastes [59, 62]. Being a non-thermal and non-chemical
33
34 process, it may also have the advantage of lower degradation and better bioactivity
35
36 preservation.
37
38

39
40 Regarding the valorization of protein from fish wastes, the uses of PEF are still
41
42 scarce, but some examples are described below.
43

44 Zhou et al. [63] studied PEF for protein extraction using mussels as raw material.
45
46 PEF protein extraction yielded faster than those obtained for enzymatic and alkali
47
48 extraction methods. The maximum protein extraction yield was 77.08% and was
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50 achieved using an electric field intensity of 20 kV/cm, a pulse number of 8, and an
51
52 enzymolysis time of 2 h.
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4 Li et al. [59] evaluated PEF-assisted enzymatic extraction of the protein from
5 abalone viscera and resulted in a hydrolysate fully hydrolyzed, with high protein yield
6 and good emulsifying properties, compared to purely enzymatic extraction. Moreover,
7 water extraction assisted by PEF of fish residues improved the antioxidant capacity of
8 resulting protein extracts when compared to conventional water and methanol extraction
9 treatments [54]. PEF-assisted enzymatic extraction also showed promise for the
10 extraction of taurine from mussels [62]. Other studies used PEF to valorize fish wastes,
11 but they are focused more on valorizing fishbone [44, 55, 64–68].

21 Despite the increasing exploitation of PEF to hydrolyze proteins, the mechanism
22 behind PEF's effects on proteins is still poorly understood. It has been suggested that
23 proteins' polar groups may take in the energy of the PEF treatment, producing free
24 radicals, and that those created radicals could impact the intramolecular interactions in
25 protein molecules, including hydrophobic and electrostatic interactions, disulfide
26 bridges, hydrogen bonds, salt bridges, and Van der Waals forces. Furthermore, the
27 apparent charge of proteins may be altered by PEF due to changes in their ionic
28 interactions. Thus, further research is required to accurately assess the impact of PEF
29 treatments on the structural and techno-functional characteristics of proteins.
30 Nevertheless, PEF is gaining interest in this field due to its sustainability, reduced
31 operational costs, high scalability, low time consumption, and, particularly, because it
32 does not seem to affect the quality of the products [69].

3.2.3. Ultrasound-assisted extraction

49 Ultrasound-assisted extraction (UAE) consists of submitting a sample matrix at
50 frequencies above human hearing levels, ranging from 20 to 10 MHz. UAE efficiency is
51 due to the creation of acoustic cavitation and mechanical impact in the sample matrix,
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4 which increases the surface area of contact between the solvent and the biomolecules
5
6 present in the sample matrix, facilitating the penetration of the solvent into the sample
7
8 matrix and releasing the desired extractible bioactive compounds [44, 64].
9

10 Based on the frequency and amplitude of the applied ultrasound waves, there are
11
12 two types of UAE: high intensity and low intensity [44, 70, 71]. Low-intensity ultrasounds
13
14 (LUS) with high frequencies (100 kHz to 1 MHz) are mainly used to assess food
15
16 products' physical and chemical characteristics without causing damage. High intensity
17
18 ultrasound (HIUS) with low frequencies (20 kHz-100 kHz) is employed to accelerate
19
20 and enhance the effectiveness of sample preparation by modifying the physical or
21
22 chemical characteristics of food [44, 70-72].
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24

25 Recently, HIUS's capability to hydrolyze biopolymers has been increasingly
26
27 exploited, and UAE has shown to be also a useful technology to
28
29 depolymerize/hydrolyze polysaccharides like dextran, xanthan, carrageenan, chitosan,
30
31 and starch^[72].
32
33

34 Though it has been described to hydrolyze macromolecules and may have the
35
36 potential to hydrolyze proteins, it is usually more associated with protein extraction.
37

38 Currently, UAE alone or combined with other technologies has been explored for
39
40 extracting valuable compounds from fish wastes, including gelatin, collagen, and
41
42 proteins [44, 64, 65, 73-75].
43

44 Gelatin is a polypeptide derived from insoluble collagen denaturation with
45
46 valuable functional properties, and it is widely used in food and pharmaceuticals [44, 65].
47

48 FBCPs/wastes, especially skin and bones, have been exploited as novel sources of
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50 gelatin. For instance, Huang et al. [65] used fresh bighead carp (*Hypophthalmichthys*
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52 *nobilis*) scales as raw material and studied the gelatin extraction by conventional water
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54 bath (CWB) and UAE-assisted conventional extraction methods. The results suggested
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4 that UAE-assisted extraction can extract fish gelatin with better rheological properties,
5
6 higher gel strength, and emulsifying properties than the CWB extract. Another study
7
8 using the same raw material [73] also showed that UAE gelatin extraction resulted in
9
10 better extraction yields and quality of the gelatin than the one extracted using CWB.
11
12 However, UAE parameters must be adjusted to prevent acoustic cavitation from causing
13
14 protein degradation. In fact, the combination of high ultrasound intensity (over 200 W)
15
16 with longer extraction times (5 h or more) led to a decrease in gel strength and melting
17
18 points of gelatin extracted by UAE [73].
19
20

21 Collagen is a structural protein also found in the skin and bones of fish, which is
22
23 valuable and extensively used in the cosmetic, food, and pharmaceutical industries [52,
24
25 74]. So, Kim et al. [74] evaluated the collagen extraction by UAE using the skin of sea
26
27 bass (*Lateolabrax japonicus*) as raw material and showed that UAE at 80% amplitude,
28
29 0.1 mol L⁻¹ acetic acid, and 3 h could successfully extract collagen from this fish by-
30
31 product and could be an alternative to the existing methodology for collagen extraction,
32
33 which imply soaking the fish skin on a solution of 0.5 mol L⁻¹ of acetic acid for
34
35 approximately three days [74].
36
37

38 Moreover, Álvarez et al. [75] studied protein extraction from **mackerel wastes** and
39
40 showed that when combining UAE with alkaline extraction and isoelectric
41
42 solubilization precipitation, protein extraction yields could be higher, using lower
43
44 extraction times and less solvent. **Besides, when using abalone viscera as raw material,**
45
46 **UAE combined with alkaline extraction also improved the protein extraction rate by**
47
48 **around 17% when compared with alkaline extraction alone, and the produced peptides**
49
50 **demonstrated iron chelating properties [49].**
51
52

53 **UAE has been recognized as a fast, clean, reproducible, and alternative non-**
54
55 **thermal extraction method compared to traditional extraction methods [34].**
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4 Nevertheless, and similarly to the other innovative processing technologies, UAE is still
5
6 not a standard technology, and more studies are required to establish standardized
7
8 protocols for each desired application in order to make full use of this technology,
9
10 including but not limited to protein extraction from food wastes.
11

12 13 14 3.2.4. *Supercritical fluid extraction*

15
16 Supercritical fluid extraction (SFE) uses solvents above their critical point
17
18 (temperature and pressure) to separate compounds from the matrix. Under supercritical
19
20 conditions, the solvents have intermediary properties between gases and liquids,
21
22 facilitating the extraction of the desired compounds. Supercritical fluids possess
23
24 diffusion, viscosity, and surface tension similar to those of gases, thus accelerating mass
25
26 transfer phenomena to and from the solid matrix, and liquid-like density and solvation
27
28 power, thus facilitating the penetration of the solvent into the solid matrix. Carbone
29
30 dioxide (CO₂) is the most widely used SFE solvent in food applications (GRAS), and it
31
32 is usually used to extract non-polar compounds [44, 76]. SFE is reported as a fast,
33
34 efficient, selective, and environmentally friendly technology for the extraction of
35
36 valuable compounds from several matrices [44, 76]. However, this technique has some
37
38 drawbacks, namely the need for specific and expensive equipment to operate at elevated
39
40 pressures, the non-selectivity for extracting polar substances, and the high energy
41
42 consumption [44].
43

44
45 Regarding the valorization of fish wastes, SFE has been mainly exploited as a
46
47 pre-treatment for oil valorization and extraction of fatty acids [76]. Nevertheless, AA
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49 recovery from *Todarodes pacificus* squid is reported using SFE, followed by SWH [53].
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3.3. Purification of the peptide fraction

Several authors include a purification step in the process of extracting bioactive compounds. Purification is crucial for obtaining peptide fractions with specific composition or biological properties. Composed by several processes, purification aims to isolate a molecule from a mixture, allowing the characterization of the structure, function, and interactions of the peptide or protein of interest. Thus, expressing peptides often involves the fusion of tags to these molecules, which is essential in the purification process [77]. After this purification step, the peptides show a high degree of purity, a crucial factor in obtaining reliable results in the evaluation of their bioactive effects. In addition, this purity becomes mandatory for their intended industrial application, particularly in the pharmaceutical sector [78].

Purifying and characterizing peptides have been a major challenge. Classical purification methods, namely dialysis, membrane separation, gel chromatography, ion exchange chromatography, and reverse-phase high-performance liquid chromatography (RP-HPLC), have been applied and combined to purify the peptides [79]. However, these methods perform a separation based on the difference in molecular polar interaction or weight, which has been pointed out as a limitation of these methods.

Ultrafiltration and nanofiltration are widely used after enzymatic hydrolysis to fractionate peptides by molecular weight. These membrane-based techniques are scalable and cost-effective, a good option for applications that balance purity with economic efficiency [80, 81]. However, additional steps may be needed for higher purity [82]. In contrast, RP-HPLC offers high peptide purity by separating them based on hydrophobicity, but its high cost and limited scalability restrict its use to high-value products [80, 81]. Gel chromatography, frequently combined with ultrafiltration or RP-HPLC, separates peptides by molecular weight and is valuable for producing peptides

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4 with reliable bioactivity, though it is time-consuming and less efficient for large-scale
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6 production [80, 81]. Ion exchange chromatography separates peptides based on charge,
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8 yielding high-purity peptides, but it is more costly and less efficient for industrial-scale
9
10 operations [80, 81]. Affinity separation, based on reversible interactions between
11
12 molecules, provides high specificity but is costly and complex, limiting its industrial use
13
14 [80, 81, 83].

15
16
17 BPs obtained from FBCPs, such as skin, viscera, and scales with antioxidant
18
19 properties, may be a potential alternative to synthetic antioxidants, benefiting human
20
21 and animal nutrition [84]. Thus, several studies are dedicated to extracting,
22
23 characterizing, and purifying bioactive compounds from different fish parts. Liu et al.^[85]
24
25 proved the tyrosinase inhibitory activity *in vitro* of the peptide derived from zebrafish
26
27 phosvitin. Scale peptides from tilapia, a tropical fish originally from Africa, were
28
29 reported by Chai et al. [86], highlighting its advantages as natural ingredients, low MW,
30
31 and easy absorption by the human body.

32
33
34 Cholecystokinin-releasing peptides contained in FBCPHs (from blue whiting
35
36 (*Micromesistius poutassou*) and brown shrimp (*Penaeus aztecus*)) were partially
37
38 purified and characterized by size (of apparent molecular weight ranging from 1000 to
39
40 1500 Da) exclusion chromatography using a Toyopearl HW-40F column. This study
41
42 pioneered the potential use of peptide molecules from FBCPs and crustacean FBCPs for
43
44 application in appetite suppressant products [87].

45
46
47 In a study developed by Ko et al.^[88], two peptides derived from the muscle
48
49 protein of the olive flounder (*Paralichthys olivaceus*) were purified and showed a strong
50
51 antioxidant action. The authors prepared the hydrolysates by enzymatic reactions of
52
53 flounder fish muscle using eight commercial proteases such as papain, pepsin, trypsin,
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55 neutrase, alcalase, kojizyme, protamex, and α -chymotrypsin. Further separation of the
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4 α -chymotrypsin hydrolysate was performed by ultrafiltration, gel filtration, and RP-
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6 HPLC.
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8 A study developed by Saidi et al.^[89] intended to produce and fractionate an FPH
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10 from the FBCPs of tuna dark muscle, highlighting the performance of ultrafiltration and
11
12 nanofiltration processes in purifying these hydrolysates. In this study, Alcalase proved
13
14 to be the most suitable protease for producing peptide fractions, with optimal
15
16 conditions: temperature 55 °C, pH 8.5, time 60 min, enzyme-to-substrate ratio 1%. The
17
18 study also verified that fractionation is improved when combining membranes.
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21 Another study subjected a proteolysate generated by alcalase from *Stichopus*
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23 *horrens*, a popular sea cucumber species, to fractionation based on the peptide
24
25 hydrophobicity. In this study, peptides were fractionated using a gradient elution of
26
27 acetonitrile in a percentage between 0% and 31.3%^[90]. The results proved that the
28
29 hydrolyzed protein components have bioactive peptides with potential application in the
30
31 food industry as functional ingredients. Similar studies on peptide fractionation using
32
33 RP-HPLC showed the elution of peptides with acetonitrile in percentages below 50%.
34
35 Peptides isolated from proteolyzed cuttlefish were purified in the third step with an
36
37 acetonitrile concentration below 29%^[91]. The most active hydrolysate was obtained
38
39 with the crude protease extract from the hepatopancreas of cuttlefish ($64.47 \pm 1.0\%$ at 2
40
41 mg of dry weight/ml) with a degree of hydrolysis of 8%. Likewise, BPs isolated from
42
43 catfish muscle hydrolysate were purified with the aim of ultrafiltration, gel filtration,
44
45 and RP-HPLC with a C18 column with acetonitrile at a concentration below 50%^[92].
46
47

48 **The production of FBCPHs presents an environmentally sustainable and**
49
50 **valuable solution for the fish processing industry. Enzymatic hydrolysis is preferred for**
51
52 **producing highly bioactive and pure peptides despite its higher costs. Emerging**
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54 **technologies like SWH, PEF, and UAE offer more efficient alternatives by reducing**
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4 processing times and energy consumption. For peptide extraction/ purification,
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6 ultrafiltration and nanofiltration are considered ideal for industrial use due to their cost-
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8 effectiveness and molecular weight separation. At the same time, RP-HPLC and ion
9
10 exchange chromatography are better for high-purity needs, especially in research and
11
12 pharmaceutical applications. Overall, optimizing hydrolysis, integrating novel
13
14 technologies, and improving purification techniques are crucial for fully disclosing the
15
16 potential of FBCPHs in the nutraceutical and pet food industries. More research is
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18 required to enhance energy efficiency and scalability.
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24 4. Fish by-products protein hydrolysates as functional ingredients

25 4.1. Peptides biological activities

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27 Due to their biological activities and nutritional value, FBCPHs and their
28
29 peptides have a wide range of applications across several industries, including food,
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31 nutraceuticals, pharmaceuticals, and cosmetics. Recently, FBCPHs and their peptides
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33 have gained significant importance in animal nutrition [12, 93]. FBCPHs deliver a rich
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35 source of peptides with a complete amino acid profile, superior bioavailability, and
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37 higher levels of essential amino acids than those derived from mammalian or plant
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39 proteins [93, 94]. Besides that, FBCPH peptides have demonstrated the potential to
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41 enhance human and animal health, preventing and alleviating symptoms of various
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43 diseases [6, 12, 93]. FBCPHs and their peptides have been shown to possess several
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45 biological activities, such as antioxidant, anti-inflammatory, anti-hypertensive, immune-
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47 modulatory, antimicrobial, or hormone-regulating properties [35, 93, 95]. The development
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49 of FBCPHs through enzymatic hydrolysis has been essential in showing these biological
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51 activities, providing sustainable, bioactive, and nutrient-rich ingredients that support the
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4 health and well-being of animals [6, 12, 32]. Each of the above-enunciated biological
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6 activities will be discussed in detail in the following sections.
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8
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10 4.1.1. Oxidative Stress Control

11 Several FBCPHs and peptides exhibit potent antioxidant activity, countering
12 reactive oxygen species (ROS) to mitigate oxidative stress [96]. Uncontrolled ROS
13 generation contributes to health disorders such as diabetes, cardiovascular issues,
14 neurodegenerative diseases, and inflammation [6, 97]. On the other hand, Food
15 deterioration is linked to lipid oxidation and secondary lipid peroxidation products [98].
16 FBCPs peptides are suggested as alternative additives to synthetic antioxidants [88].
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24 The antioxidant efficacy of FBCPHs and peptides depends on production
25 parameters like enzymatic hydrolysis specificity and degree of hydrolysis (%DH).
26 These parameters influence peptide structure and, consequently, antioxidant activity [6,
27 95, 99]. Antioxidant peptides generally have shorter chain lengths (0.5–1.5 kDa),
28 hydrophobic amino acid-rich composition, and aromatic amino acids like tyrosine and
29 tryptophan, with valine and leucine at the N-terminus [100–104].
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37 Several authors commonly correlate low MW and antioxidant activity of
38 FBCPHs [48, 105, 106]. Low-molecular-weight antioxidant peptides interact more easily
39 with free radicals and have a higher capacity for elimination [102, 106]. However, Sierra et
40 al.^[100] reported higher antioxidant activity in hydrolysate fractions from red tilapia
41 scales with MW of 10–100 kDa and 3–10 kDa compared to the <3 kDa fraction.
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4 to an excess of electrons that could be donated by interacting with ROS/free radicals
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6 [103].
7

8 High metal ion-chelating activity of Stripped weakfish FBCPHs was detected by
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10 Lima et al.^[102]. Furthermore, metal ion-chelating activity is related to the presence of
11
12 basic (arginine and lysine) and aromatics AAs (e.g., phenylalanine) at the C-terminal
13
14 position, together with the high content of hydrophobic AAs (leucine). On the other
15
16 hand, tryptophan at the N-terminal position was associated with its capacity to act as a
17
18 hydrogen donor in the sequestration of hydroxyl radicals ^[102]. Positively charged AAs
19
20 as lysine at the N- and C-terminal is another valuable feature detected in antioxidant
21
22 FBCPHs ^[105, 106].
23
24

25 The mechanisms of antioxidant activity studied to FBCPHs include in vitro
26
27 radical scavenging [2,2-diphenyl-1-picrylhydrazyl (DPPH) and 2,2'-azino-bis-(3-
28
29 ethylbenzothiazoline-6-sulfonic acid) (ABTS)], superoxide and OH-radical scavenging
30
31 ^[105, 106], but also ferric reducing antioxidant power (FRAP), metal chelating activity ^{[48,}
32
33 ^{102, 104]}. Cell oxidative stress reduction has recently been assessed ^[100, 103]. Excessive
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35 ROS and oxidative stress are associated with several diseases, emphasizing the
36
37 significance of antioxidant peptides ^[12, 100].
38
39

40 The antioxidant activity of FBCPHs is commonly evaluated regarding in vitro
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42 radical scavenging activity by 2,2-diphenyl-1-picrylhydrazyl (DPPH) and 2,2'-azino-
43
44 bis-(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS); superoxide (O₂⁻) and OH-radical
45
46 scavenging activities ^[105, 106], but also by evaluation of ferric reducing antioxidant
47
48 power (FRAP) and metal chelating activity ^[48, 102, 104]. Cell oxidative stress reduction
49
50 has recently been assessed in red tilapia viscera and scales hydrolysates ^[100, 103]. Red
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52 tilapia viscera hydrolysate (0.1 mg mL⁻¹) and its < 1 kDa fraction (0.25 mg/mL)
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54 demonstrated no cytotoxic effect and cytoprotective effects preventing the decrease of
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4 the cell viability and ROS level accumulation in Caco-2 cells treated with 5 mM H₂O₂
5
6 to induce oxidative stress, being potential effective antioxidant agents against ROS-
7
8 mediated intestinal injuries, principally the < 1 kDa fraction. In another study, red
9
10 tilapia scales hydrolysate and fractions (2 mg/mL) reduced oxidative stress in A7r5 cells
11
12 stimulated by 1 μM of Ang II. Ang II activated ROS production in A7r5 vascular cells,
13
14 which can react with nitric oxide and trigger inflammation and proliferation, leading to
15
16 systemic vascular dysfunction and accelerating hypertension/ atherosclerosis
17
18 development [100].
19
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21

22 4.1.2. Prevention of obesity-associated diseases

23
24 Obesity is a global public health problem, affecting both humans and their pets,
25
26 with associated risks such as hypertension, type 2 diabetes, cardiovascular disease,
27
28 osteoarthritis, and cancer^[107–109]. Besides the capacity to reduce oxidative stress induced
29
30 by obesity, FBCPHs are reported to possess other vital activities against several obesity-
31
32 associated diseases, such as angiotensin I-converting enzyme (ACE) and dipeptidyl
33
34 peptidase IV (DPP-IV) inhibitory activities, offering potential benefits against
35
36 hypertension and type 2 diabetes^[104, 110–112], but also satiating, lipid-lowering, and
37
38 antiatherogenic potential effects^[109, 113–115].
39
40

41 ACE inhibitory peptides, which typically have low molecular weight (8-20 AA
42
43 residues) and moderate hydrophobicity, have been identified in FBCPHs from shortfin
44
45 scad (*Decapterus macrosoma*)^[116]; sturgeon skin^[104] and rainbow trout frames^[117].
46
47 Besides the low molecular weight (< 3 KDa), the antihypertensive hydrolysate (IC₅₀
48
49 2.20 mg/mL) from shortfin scad exhibited greater hydrophobicity at the C-terminal
50
51 (e.g., alanine, leucine) with an aliphatic AA (glycin) at the N-terminal. On the other
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53 hand, the ACE inhibition activity of sturgeon skin peptides was mainly attributed to
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4 strong hydrogen bonds with lysine at the C-terminal and aliphatic AAs (two proline) at
5
6 the N-terminal [104]. Moreover, a study of selective fractionation (electrodialysis with
7
8 ultrafiltration membrane) of rainbow trout frames hydrolysates highlighted the role of
9
10 hydrophobic and positively charged AAs in the ACE inhibitory activity [117]. Cationic
11
12 peptide fraction showed a potent inhibitory activity (IC₅₀ 0.0036 mg/ mL) compared to
13
14 initial FBPH (0.015 mg/ mL).
15

16
17 The mechanism of action of ACE inhibitory activity of FBCPHs is linked to the
18
19 modulation of the renin-angiotensin system (RAS), relaxing blood vessels, and
20
21 decreasing blood pressure [104]. Besides that, the inhibition of ACE may increase nitric
22
23 oxide availability and improve endothelial function, as observed in subjects with high
24
25 cardiovascular disease after ingesting an FBCPH from Nile tilapia FBCPs [118]. An
26
27 enhancement of muscle O₂ desaturation and resaturation parameters, with no changes in
28
29 blood pressure, was also detected [118]. In an *in vivo* study using a model of high-salt
30
31 and -fructose diet-induced hypertension in Wistar rats, a reduction of the systolic blood
32
33 pressure and consequent hypertension attenuation was verified after administration of a
34
35 viscera FBPH and its <1 kDa peptide fraction [115].
36

37
38 Regarding the DPP-IV inhibitor activity of peptides obtained from FBCPs, the
39
40 lower MW and presence of hydrophobic AAs at the N-terminal position have been
41
42 reported using salmon trimmings [116], and skin [119], tilapia FBCPs [112] and sturgeon
43
44 skin [104]. The hydrolysates obtained enzymatically from salmon skin gelatin and
45
46 trimmings with higher DPP-IV inhibitory activity exhibited a higher proportion of
47
48 peptides <1 kDa [119]. Hydrophobic AA residues such as phenylalanine, arginine, and
49
50 tyrosine were also detected. More recently, the molecular docking studies of DPP-IV
51
52 inhibitory peptides obtained from sturgeon skin revealed that their activity is mainly
53
54 attributed to hydrogen bonds and hydrophobic interactions involving tyrosine, arginine,
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4 and serine residues ^[104]. Besides that, the hydrolysates obtained from salmon skin
5
6 gelatin and trimmings also showed glucagon-like peptide-1 (GLP-1) secretory activity
7
8 and a positive effect on insulin release from BRIN- BD11 cells. Other FBCPHs also
9
10 increase insulin secretion by stimulating GLP-1 secretion, further increasing insulin
11
12 release and improving glucose homeostasis ^[95, 112]. Theysgeur et al.^[112] identified new
13
14 peptides from enzymatic hydrolysis of tilapia FBCPs that stimulate GLP-1 secretion
15
16 after canine gastrointestinal simulated digestion. GLP-1, as an intestinal anorexigenic
17
18 hormone, also exerts a satiating effect via different pathways. After a transport study
19
20 through the Caco-2 cell monolayer, peptides with DPP-IV inhibiting activity were also
21
22 found in the hydrolysate obtained from the tilapia FBCPs.
23
24

25
26 Regarding the effects of FBCPHs on body weight regulation, hydrolyzed fish
27
28 collagen peptide obtained from tuna skin by subcritical water was tested in a mouse
29
30 model of obesity induced by high-fat diet feeding ^[113]. The hydrolysate reduced body
31
32 weight gain and improved the serum lipid profiles of obese mice. Besides that,
33
34 differentiation inhibition of 3T3-L1 preadipocytes into adipocytes by decreasing the
35
36 expression of adipogenic master genes was also noticed.
37
38

39 40 *4.1.3. Reduction of anti-inflammatory response*

41
42 The hydrolysates from FCBPs have been reported to possess potent anti-
43
44 inflammatory properties ^[120]. Inflammation is a typical response of the immune system
45
46 to lesions and infection, modulated by inflammatory mediators and pro-inflammatory
47
48 compounds such as cytokines released by activated macrophages or other cells.
49
50 Excessive inflammation (uncontrolled production of pro-inflammatory compounds) can
51
52 contribute to various acute and chronic diseases, such as metabolic syndrome and
53
54 inflammatory bowel disease ^[121, 122]. Several FBCPHs have shown significant
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4 modulatory effects against inflammatory reactions through inhibition of the production
5
6 of pro-inflammatory markers, such as nitric oxide (NO), inducible nitric oxide synthase
7
8 (iNOS), and cytokines (e.g., Tumor necrosis factor- α (TNF- α)) [123]. Among them, the
9
10 FBCPHs from FBCPs of canned sardine [123], herring [124], sturgeon caviar muscle [120],
11
12 tuna fishery [125], mixed-species fish skin [126], and Atlantic salmon heads and backbones
13
14 [127] can be highlighted.

15
16
17 An anti-inflammatory hydrolysate was obtained from canned sardine FBCPs
18
19 hydrolysis using a brewing spent yeast protease extract. According to Vieira et al. [123],
20
21 the desalted sardine protein hydrolysate (<10 kDa) decreased all inflammation markers
22
23 (2.0 mg peptides/mL) in the endothelial cell line (EA.hy926), but also the co-culture
24
25 model with an intestinal cell line (Caco-2), compared to TNF- α -treated control. The
26
27 potential anti-inflammatory effects during endothelial inflammation detected in sardine
28
29 protein hydrolysate were related to the presence of the AAs glycine, histidine, and
30
31 cysteine despite its high content in glutamic acid, glutamine, aspartic acid, and alanine
32
33 (51.2%). Durand et al. [124] separated by electrodialysis with ultrafiltration membrane
34
35 (EDUF), a herring milt hydrolysate obtaining two anionic (higher acidic AAs content)
36
37 and two cationic fractions (higher presence of basic AAs). Positively charged peptides
38
39 characterized both cationic fractions, with arginine as one of the main AAs described as
40
41 characteristic of effective anti-inflammatories. Both cationic fractions revealed the
42
43 potential to prevent metabolic syndrome. These fractions exhibited anti-inflammatory
44
45 effects by reducing iNOS activation in the lipopolysaccharides. NO production was
46
47 induced in J774 mouse macrophage cells. Furthermore, FPH from the kingfish frame
48
49 [128] ameliorated the adverse effects of juvenile barramundi's poultry by-product meal-
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51 based diet in aquaculture. Chaklader et al. [128] showed the positive impact of FPH
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4 supplementation at 10% in terms of modulation of disease resistance and pro-
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6 inflammatory and inflammatory cytokines post-*Vibrio harveyi* infection.
7

8 FCBP hydrolysates' potential to prevent gastrointestinal inflammation-related
9
10 diseases such as ulcerative colitis has also been reported. Ulcerative colitis is a primary
11 form of inflammatory bowel disease [120]. Skin collagen of Pangas catfish and sturgeon
12 FBCPHs significantly decreased the severity of dextran sodium sulfate (DSS) -induced
13 damage mice model [120, 122]. Besides that, the *in vitro* experiments performed by
14
15 Sivaraman and Shanthi [122] revealed that skin collagen hydrolysate also attenuates TNF-
16
17 α induced tight junction barrier disruption. On the other hand, the suppression of DSS-
18
19 induced activation of the NF- κ B and MAPK pathways in the colon was detected after
20
21 the administration of sturgeon FBCPH [120]. Gao et al.[120] also noticed the restoration of
22
23 gut microbiota of colitic mice, namely an increase in the *Bacteroidetes/Firmicutes* ratio
24
25 and the relative abundance of other benefic bacteria while decreasing the abundance of
26
27 potentially harmful bacteria such as *Enterococcaceae*. Positive modulation of distal
28
29 intestinal microbiota with an enrichment of the *Firmicutes* and *Fusobacteria* was
30
31 reported in juvenile barramundi with a poultry BCPs meal-based diet after tuna BCPs
32
33 hydrolysate supplementation [125]. The upregulation of pro-inflammatory cytokines (IL-
34
35 1β and TNF- α) and downregulation of the anti-inflammatory cytokine IL-10 were also
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37 reported by Siddik et al.[125].
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46 4.1.4. Role in microorganisms' control and modulation

47 Several studies have highlighted the role of microbiota in health and diseases.
48
49 Gut microbiota dysbiosis is associated with obesity and other metabolic diseases such as
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51 diabetes, asthma, inflammatory bowel disease, and rheumatism [120, 129]. Some studies
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53 showed that peptides could beneficially modulate gut microbiota. However, the
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4 microbiota regulation by peptides obtained from FBCPs has barely been studied. Wang
5
6 et al.^[130] valorize Walleye pollock skin through the production of collagen peptides,
7
8 which significantly increased the levels of some beneficial bacteria and decreased the
9
10 levels of bacteria involved in inflammation in the high-fat diet (HFD) mouse model.
11
12 Besides, significant anti-obesity effects were also described.
13

14
15 The role of FBCPHs in microbiota regulation is also described relative to wound
16
17 healing. The oral administration of fish skin collagen peptides to the murine wound
18
19 model promoted wound healing by regulating microflora colonization in the wound
20
21 tissues, controlling the inflammatory reaction, and increasing angiogenesis and collagen
22
23 deposition ^[131]. Until now, peptides from FBCPs have not been evaluated regarding
24
25 their potential benefits in oral microbiota modulation, but they could be promising
26
27 agents in the equilibrium of oral microbiota. Oral microbiota equilibrium has not been
28
29 investigated as much as gut microbiota. However, its imbalance and dysbiosis are
30
31 linked to teeth and oral diseases, exerting relevant influence in systemic diseases such as
32
33 rheumatoid arthritis, inflammatory bowel disease, and chronic kidney disease ^[132].
34

35
36 Besides their microbiota modulation role, FBCPHs have been described as
37
38 potential antimicrobial compounds. Antimicrobial peptides (AMPs) are produced in fish
39
40 as part of their immune systems ^[133]. AMPs exhibit activity against Gram-positive and
41
42 Gram-negative bacteria, fungi, viruses, and unicellular protozoans ^[35]. AMPs such as
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44 piscidin, defensin, hepcidin, cathelicidin, and histone-based peptide families have been
45
46 isolated from several kinds of fish ^[134]. Besides that, AMPs have been produced
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48 enzymatically from FBCPs, such as yellowfin tuna viscera ^[135], mixed FBCPs of the
49
50 distribution market ^[105], tuna skin collagen ^[48], and stripped weakfish FBCPs ^[102].
51
52 However, its exploitation and application have been less studied than antioxidant and
53
54 anti-hypertensive activity.
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4 Most antimicrobial peptides identified in FBCPs are cationic with low MW and
5 hydrophobic properties. According to a study of fractionation of yellowfin tuna viscera
6 hydrolysate, the shortest peptides (< 3 kDa fraction) are the most active against Gram-
7 negative (*Escherichia coli*, *Pseudomonas aeruginosa*) and Gram-positive (*Listeria*
8 *monocytogenes*, *Staphylococcus aureus*) with a Minimum Inhibitory Concentration
9 (MIC) of 0.5 mg/mL [135]. Furthermore, the higher concentration of positively charged
10 and hydrophobic AAs was < 3 kDa fraction. The presence of AAs with a positive
11 charge (lysine, arginine, and histidine), in addition to the hydrophobic AA content
12 (60%), such as alanine, leucine were also associated with the antimicrobial activity of
13 10-30 kDa fraction of collagen hydrolysate derived from mixed FBCPs against *E. coli*
14 (10 mg/mL) [105]. Lima et al.[102] also reported a higher proportion of hydrophobic AAs
15 in the highest effective Stripped weakfish FBCPHs against *E. coli* (50 mg/mL) and *S.*
16 *aureus* (100 mg/mL) compared to the other hydrolysates. Moreover, Ahmed and
17 Chun[48] reported that a hydrolysate of tuna skin collagen obtained by subcritical water
18 hydrolysis at 280 °C exhibited either the highest content of hydrophobic AA and
19 antimicrobial activity (against *Bacillus cereus*, *Pseudomonas putida*, *P. aeruginosa*).
20 The hydrophobic character of peptides facilitates the interaction between the positively-
21 charged peptide with the negatively-charged surface of the bacteria, and the AAs
22 charged positively penetrate bacterial membranes interacting with the negative charges
23 of the cell wall of phospholipids, promoting cellular lysis [48, 105].
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46 Besides their broad antimicrobial spectrum, AMPs have been pointed out as
47 promising alternatives to antibiotics due to their low level of induced resistance and
48 potential capacity to modulate the inflammatory response and gut microbiota [136]. The
49 overuse of antibiotics is a severe problem in animal production, including aquaculture,
50 leading to increasing antimicrobial resistance, microbiota dysbiosis, and the
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4 development of other diseases. Recently, a novel AMP from rainbow trout
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6 (*Oncorhynchus mykiss*) with antibacterial activity against *Streptococcus iniae* (zoonotic
7
8 pathogen) was identified using an *in silico* approach [137]. Regarding its
9
10 immunomodulatory activity, treating rainbow trout cells with the AMP did not
11
12 upregulate the immune genes. However, it stimulated the cell Line RTS11 (rainbow
13
14 trout macrophage/monocyte-like cell line) and significantly upregulated the TNF- α
15
16 gene.
17
18

20 4.1.5. Neuroprotective ability and behavioral issues control

21
22 Peptides or protein hydrolysates have been described as promising
23
24 neuroprotective agents that act in neuronal survival via modulating signaling pathways
25
26 but also inhibit neuroinflammation and consequently suppress cognition decline [138, 139].
27
28 FBCPHs have been exhibited to be potential candidates for memory and learning-
29
30 improving functions [138, 140, 141]. Anchovy hydrolysates proved to be a potential
31
32 improver for memory in scopolamine-induced amnesia mice. The anchovy hydrolysate
33
34 obtained enzymatically after 8 h exhibited better results, probably due to its high
35
36 glutamate content [138]. Potential positive effects in memory and learning deficiency
37
38 were described for peptides from lantern fish (*Benthoosema pterotum*). Lantern fish
39
40 peptides revealed efficacy in neuron protection in human neuroblastoma SH-SY5Y cells
41
42 against H₂O₂-induced apoptosis and improved D-gal-induced deficit of memory and
43
44 learning ability in aging ICR mice. Lower levels of D-gal-induced thiobarbituric acid
45
46 reactive substances (TBARS) and endothelial NOS were detected in mice fed with
47
48 lantern fish hydrolysate and higher levels of glucose-6-phosphate dehydrogenase and
49
50 brain-derived neurotrophyl factor in comparison to the control group [140]. Sea cucumber
51
52 (*Cucumaria frondosa*) hydrolysate also ameliorates learning and memory deficits of D-
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4 galactose-induced aging mice^[141]. In Lin et al.^[141] study, peptides' neuroprotective
5
6 activity seems to be associated with the capacities to inhibit lipid peroxidation and
7
8 protein oxidation, increase antioxidant enzyme activity (superoxide dismutase and
9
10 glutathione peroxidase), and down-regulate acetylcholinesterase. The expression of
11
12 Klotho, which acts as an anti-aging humoral factor, is also upregulated. More recently,
13
14 peptides isolated from round scad hydrolysate exhibited neuroprotection action against
15
16 glutamate-induced neurotoxicity in PC12 cells ^[139]. Tyrosine and tryptophan also
17
18 proved to be related to the neuroprotection activity of peptides obtained from round
19
20 scad.
21

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23 Until now, few studies have assessed the neuroprotective ability of FBCPHss
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25 ^[142, 143]. In Chataigner et al.^[142] study, the supplementation with low MW peptides from
26
27 FBCP hydrolysate combined with *omega-3* long-chain polyunsaturated fatty acids
28
29 prevented the age-related spatial short-term memory deficits and modulated navigation
30
31 strategies adopted during spatial learning in aged mice. Besides that, FBCP peptides
32
33 also reduced anxiety-like behavior, restored plasmatic corticosterone levels like those of
34
35 adult animals following acute stress, and modulated the hypothalamic stress response. A
36
37 sardine byproduct hydrolysate (Peptidyss®) also decreased stress reactivity in Balb/c
38
39 mice by reducing corticosterone levels 30 min after stress induction compared to control
40
41 mice ^[143]. The sardine BCPs hydrolysate supplementation modulated stress-responsive
42
43 gene expression, especially in the hippocampus. Similar positive effects on stress and
44
45 anxiety responses were found in dogs supplemented with FBCPs ^[144, 145]. According to
46
47 Landsberg et al.^[145], the mackerel and cod hydrolysates (Gabolysat® PC60) exhibited
48
49 anxiolytic properties, decreasing hyperactivity and reducing cortisol response to stress.
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52 On the other hand, a new dietary supplement (derived from fish hydrolysate and melon
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4 juice concentrate rich in superoxide dismutase) promoted dog-human interactions and
5
6 tended to reduce subtle stress behaviors ^[144].
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8
9 Lastly, opioid-like peptides such as glyprolines (PGP and GP) are other peptides
10 that present the capacity to relieve the behavioral problems caused by stress in rats
11 subjected to a forced swimming test (10 min). Peptides Pro-Gly-Pro and Gly-Pro
12 intranasal ^[146] and intraperitoneal^[147] were administrated 15 minutes after the end and
13
14 before the stress test, respectively. Both studies demonstrate that Pro-Gly-Pro and Gly-
15 Pro peptides prevent and mitigate stress-induced anxiety and behavioral disorders,
16
17 potentially through interactions with CNS structures involved in stress responses.
18
19 However, Pro-Gly was less effective, highlighting the importance of specific peptide
20 sequences in conferring antistress properties. Despite this, the opioid-like activity of
21 peptides or hydrolysates from FBCPs has not yet been explored ^[148].
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29 Overall, FBCPHs exhibit a wide range of bioactivities. Their ability to mitigate
30 oxidative stress, regulate lipid metabolism, inhibit enzymes linked to obesity-related
31 diseases, and control inflammation highlights their potential as functional ingredients in
32 both human and animal health. In addition, FBCPHs present novel insights into the
33 modulation of gut microbiota and stress-related behaviors, particularly in
34 neuroprotection and stress reduction, showing their potential role in improving
35 cognitive function and emotional well-being. These findings highlight the FBCPHs'
36 potential as an environmentally sustainable source of proteins and bioactive peptides
37 that support overall health. However, some limitations must be addressed for their
38 effective application, namely the variability in peptide efficacy due to molecular weight,
39 amino acid composition, and hydrolysis conditions, emphasizing the need for
40 standardized production methods to improve bioactivity. Additionally, further in vivo
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4 studies are needed to elucidate the underlying mechanisms, validate their therapeutic
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6 potential, and establish safety, efficacy, and acceptable use levels.
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9 Future research on the potential of FBCPHs as alternatives to antibiotics in
10
11 animal nutrition, as well as their effects on oral microbiota and neuroprotection in
12
13 humans and animals, needs to be addressed. Understanding these aspects will contribute
14
15 to developing innovative bioactive ingredients from FBCPs.
16
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18 **5. Fish by-products hydrolysates application on pet food**

19 **5.1. Pet food demands and trends**

20
21
22 Pet ownership is steadily increasing globally, including in emerging economies,
23
24 and according to the European Pet Food Federation ^[149], 90 million European
25
26 households (46%) own a pet, with cats (113 million) and dogs (92 million) being the
27
28 most popular companion animals in Europe. Higher-income, increased urbanization,
29
30 demographic changes such as family size, millennial couples having children late in life,
31
32 people living alone, high level of education, and the recent COVID-19 pandemic
33
34 constitute drivers for this trend in the pet population ^[150, 151]. Consequently, sales of pet
35
36 food have increased dramatically, reaching 27.7 billion euros in Europe in 2021 with
37
38 3.1% annual growth for the pet food industry ^[149], and expected to progress at a
39
40 compound annual growth rate of 7.22% during the period 2022-2032 ^[152].
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44 Dry pet food is the most popular among the three commercially available pet
45
46 foods (dry, semi-dry, and wet) because of its long shelf life ^[153]. Pet food markets offer
47
48 various commercial diets formulated to meet nutritional requirements according to life
49
50 stage, health status, and activities. In Europe, regulatory guidelines and
51
52 recommendations are published by the FEDIAF, based primarily on research data
53
54 published by the National Research Council ^[154].
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4 To ensure sustainable pet ownership, pet food must be simultaneously
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6 sustainable and affordable while assuring the requirements for optimum animal health
7
8 and well-being ^[155]. Market research has been reporting a trend towards premiumization
9
10 ^[156], with pet owners demanding high-quality ingredients in line with the increasing
11
12 humanization of pets that are now considered family members ^[157, 158]. Concurrently,
13
14 pet owners increasingly favor food's functional value to promote pets' health and well-
15
16 being. Among the top health-related claims by pet food brands, high protein is the most
17
18 common and natural ingredient, the second being vitamins, antioxidants, and immune
19
20 system health ^[156]. Additionally, pet owners' sociocultural factors and eating habits have
21
22 been suggested to influence their companion animals' choice of pet foods and eating
23
24 habits ^[159].

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26
27 Protein requirements for dogs and cats are relatively high, set at 18-25% and 25-
28
29 33.3% (dry weight basis), respectively ^[149]. Compared to vegetable sources, animal-
30
31 based ingredients present several advantages, such as high protein content and
32
33 digestibility and provision of significant amounts of vitamins (e.g., vitamin B₁₂) and
34
35 minerals in organic forms, thus making them more bioavailable for the animal ^[8, 160]. A
36
37 growing trend in the pet food industry is replacing rendered animal meals, traditionally
38
39 representing the most common protein source in pet food, with raw animal proteins ^[161],
40
41 more in line with ancestral and unprocessed diets. However, this trend exacerbates the
42
43 pressure on natural resources ^[162] and increases the pet food industry's carbon footprint
44
45 ^[163]. A sustainable alternative lies in using BCPs from meat and fish processing, which
46
47 is negatively perceived by owners who demand human-grade meat as the main
48
49 ingredient for their pets ^[164]. The inclusion of BCPs under the new food category
50
51 'upcycled food' can contribute to these products rebranding and improve public
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53 acceptability while promoting industry sustainability and repurposing food that
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4 otherwise would be wasted ^[165, 166]. “Upcycled ingredients” have been gaining
5
6 popularity among pet food and treat brand manufacturers. However, consumer
7
8 consciousness remains limited. In a recent survey, 66% of pet owners reported
9
10 unfamiliarity with the term “upcycled ingredients”^[167]. However, only 7% indicated that
11
12 they have already fed their pets with “upcycled ingredients,” while 54% expressed
13
14 willingness to try them. For 24% of respondents, acquiring more information would be
15
16 the initial step toward “upcycled ingredients” adoption and only 7% of pet owners
17
18 indicated no interest in the concept. Besides that, the highest awareness was registered
19
20 among UK consumers and the lowest among US pet owners. Furthermore, younger
21
22 generations were more likely to recognize the term than older generations. In this
23
24 context, FBCPHs are particularly interesting to pet food. Indeed, FBCPHs provide AAs
25
26 more easily absorbed in the small intestine ^[13], with hypoallergenic ^[12] and bioactive
27
28 properties, acting as palatability enhancers ^[11] and contributing to pet food sector
29
30 sustainability and circular economy ^[164]. However, despite the high interest in these
31
32 ingredients, studies on companion animals are scarce ^[13].
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37 ***5.2. European Pet food legislation on functional ingredients***

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39 When developing novel pet food products enriched with functional ingredients,
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41 it is essential to consider that appropriate regulatory requirements must be complied
42
43 with to place the product on the market. Both pet food manufacturing and
44
45 commercialization are highly regulated worldwide, with legislation covering the
46
47 ingredients, the production process, and the marketing and sales. The aim is to ensure
48
49 that pet food products are safe, fulfill the nutritional needs of pets, and that the
50
51 information provided to consumers is accurate and truthful ^[168].
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4 The legal requirements for pet food are established explicitly by country and/or
5 region. This work will focus on the European Union (EU) legislation, where the EU
6 Commission regulates pet food, the EU Parliament, and the Council of the EU ^[168]. The
7 main legal requirements for pet food production and commercialization are outlined in
8 **Table 1.**
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----- Table 1 -----

For the use of functional ingredients in pet food, EU Regulation 767/2009 is particularly relevant since it provides basic rules for product claims (including functional claims), labeling, and packaging. It also provides a framework for establishing and marketing PARNUTs (products for particular nutritional purposes), i.e., legally “dietetic” pet food products.

Aiming to explain and clarify how EU Regulation 767/2009 requirements work in practice, FEDIAF developed the Code of Good Labelling Practice in 2011, revised in 2019 ^[169]. Both versions of the document were endorsed and recognized by the EU authorities. The Code intends to offer clear guidance to the EU Member States, avoiding different interpretations and applications of the defined rules. Therefore, the FEDIAF Code must be considered and analyzed when novel pet food products with functional claims are under development.

According to the Code, functional claims describe the effect of a complete or complementary pet food or a nutrient, substance, characteristic, or additive in the pet food on the body's growth, development, or normal functions. It provides a specific physiological benefit and may concern “optimization of the nutrition and support or protection of the physiological conditions” (R. 767/2009, Art. 13.2). These effects must

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4 go beyond meeting the basic nutritional needs of the animals. Functional claims can be
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6 considered as nutrient function claims, enhanced function claims, or health maintenance
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8 and decreased disease risk claims, according to the general requirements indicated in
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10 Table 2 ^[169].

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13 -----Table 2-----
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17 Functional claims (as well as other types of product claims) shall be objective,
18
19 verifiable to the competent authorities, and understandable by the pet food user, thus
20
21 meeting the following requirements ^[169]:

- 22
23
24 i. Must be substantiated at the time of putting on the market;
25
26 ii. Must not confuse or mislead purchasers;
27
28 iii. Must not denigrate other pet foods or suggest that other pet foods do not possess such
29
30 characteristics when it is not true.

31
32 All claims must be substantiated and verifiable, but the degree of substantiation
33
34 will depend on the type of claim made. In the case of functional claims, the level of
35
36 substantiation depends on whether the claim is considered generic or innovative.
37
38 Generic claims are considered when well-established and recognized knowledge exists,
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40 typically functions of approved additives and/or nutrient functions. In that case,
41
42 substantiation is based on general (scientific) knowledge, with published literature and
43
44 documentation demonstrating the beneficial effects of the pet food product. On the other
45
46 hand, innovative claims are not yet widely recognized. Their substantiation may be
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48 based on a comprehensive review and evaluation of all available scientific data related
49
50 to the claim's validity, whether published or in-house, irrespective of whether its impact
51
52 is favorable or otherwise ^[169].

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4 As a final remark, it is important to note that products with functional claims
5
6 should be clearly separated from PARNUTs, which correspond to “dietetic” pet food
7
8 products. These are conceived to meet the specific nutritional needs of animals whose
9
10 process of assimilation, absorption, or metabolism is or could be temporarily or
11
12 irreversibly impaired and which can benefit from ingestion of the feed appropriate to
13
14 their condition. In this case, the claims are strictly controlled, and thus, diets may only
15
16 be marketed as PARNUTs if their indication and nutritional characteristics are listed
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18 explicitly in Directive 2008/389 [168, 169].
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24 **5.3. Fish hydrolysates in Pet food**

25
26 In the last few years, the life expectancy of companion animals has increased
27
28 due to advances in pet nutrition and veterinary care. Simultaneously, a higher risk of
29
30 developing chronic diseases by pets was also verified. As a result, protein hydrolysates
31
32 have been rising as promising nutraceuticals of interest in the animal health market [12].
33
34 In the pet food sector, protein hydrolysates are among the most popular palatability
35
36 enhancers, being considered to exert satiety effects, and commonly used to manage
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38 adverse food reactions, gastrointestinal and dermatological diseases, and to reduce
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40 stress.
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43 The increasing demand for premium diets promoting nutritional and functional
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45 benefits, natural and sustainable ingredients, and organic and clean-label products has
46
47 amplified the fish protein hydrolysate industry. Indeed, a recent report from Global
48
49 Market Insights Inc. (2024) [170] outlined that the fish protein hydrolysate market size
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51 for animal feed and pet food was valued at USD 233.08 million in 2023, being projected
52
53 to grow at 5.5% CAGR from 2024 to 2032, with the enzymatic hydrolysis segment
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4 comprising the main market share due to its functional benefits. The valorization of fish
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6 by and co-products and their reintroduction into food and feed chains, namely on pet
7
8 food, is economically advantageous and contributes to counteracting their negative
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10 environmental impact while adding value for fisheries and aquaculture sectors under a
11
12 circular economy approach.
13

14
15 Despite the wide usage of protein hydrolysates, the development of FBCPHs
16
17 designed for formulating pet food has been scarcely studied compared to aquaculture
18
19 and animal feed [12]. Folador et al.[11] conducted one of the first studies assessing the
20
21 potential utilization of FBCPHs in pet formulations based on the chemical composition
22
23 characteristics and protein quality indices. In this study, different FBCPs were minced
24
25 (sole, pink salmon, red salmon, and pollock late season) after being cooked and
26
27 deboned, and the decanted liquid was hydrolyzed using commercial papain with the
28
29 addition of 0.01% ethoxyquin (antioxidant). FBCPHs' protein quality indices were
30
31 affected by the specific part of the fish used. More recently, the production of enzymatic
32
33 hydrolysates achieved the valorization of FBCPs (heads, viscera, trimmings, and
34
35 frames) generated from aquaculture turbot (*Scophthalmus maximus*) filleting. The
36
37 optimized FBCPHs were obtained using 0.2% (v/w) of alcalase for 3 h at 60 °C and pH
38
39 8.5. The application as an ingredient in the formulation of human protein concentrates,
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41 pet food diets, and aquaculture feeds was proposed based on the concentration of
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43 soluble protein, and the adequate balance of AA, protein digestibility, and bioactivities
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45 of FBCPHs attained [30].
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5.3.1. Cats

Scientific studies evaluating functional foods or supplements, including fish hydrolysates in cats, are almost inexistent (Table 3). Commercially available elimination diets with the inclusion of partially hydrolyzed salmon^[171] and hydrolyzed fish protein^[172] were shown to reduce the pruritus visual analog scale and to score feline allergic dermatitis, thus demonstrating their benefit to diagnose and manage cutaneous adverse food reaction, a common disease affecting cats. Moreover, the study conducted by Jeusette et al.^[173] with a prescription diet supplemented with 0.1% of fish peptides fed for 10 weeks showed a reduction in stress biomarkers, with a decrease in the average 24-hour urinary cortisol/creatinine ratio and an increase in serotonin levels. However, the supplement complexity, including L-tryptophan and lemon balm extract with a minimum of 5% rosmarinic acid and oligofructose, does not allow us to attribute the effects observed solely to fish hydrolysate. Similarly, effects on reducing joint degeneration and spondylitis deformans through a supplement with collagen peptides from fish scales and D-glucosamine from crabs are impossible only to be attributed to fish peptides^[174]. Although the authors remain optimistic about the use of FBCPHs on cats' diets, more research is needed to support their benefits on animal health and welfare.

-----Table 3-----

5.3.2. Dogs

Despite the considerable interest in hydrolyzed protein and its use in commercial diets, studies with functional foods and diets containing fish hydrolysates in dogs are scarce (Table 4).

A significant effort in diet formulation is focused on palatability as it strongly affects intake and, thus, nutrient provision. Protein hydrolysates are among commercial pet food's most popular palatability enhancers [11, 12], but research studies are scarce. Sensorial characteristics have been associated with a protein source and mixture of peptides [175], being short peptides and the amino acids taurine, glycine, arginine, glutamic acid, and alanine considered feeding stimulants for companion animals [12]. Additionally, bitterness was earlier associated with the molecular weight of peptides, with increased bitterness reported when the molecular weight of the peptides of soy protein hydrolysates ranged from 4 to 2kDa and decreased bitterness with peptides < 1kDa [176]. When comparing a diet with salmon hydrolysate with a chicken-based control diet, dogs showed a preference for the salmon hydrolysate diet with significant effects on the first approach and taste and intake ratio, suggesting that salmon hydrolysate provided a taste that the dogs preferred [11]. Additionally, in a recent study performed with adult Beagle dogs fed diets differing on protein hydrolysate and oil sources, with the experimental diet containing fish hydrolysate and oil in substitution of shrimp hydrolysate and salmon oil, no effect was observed on the first diet approached and tasted, and on intake ratio^[15]. Both diets presented the same kibble size, shape, and texture and similar amino acid profiles, but they contained different fatty acid profiles, and the peptide size was not evaluated, thus making it difficult to draw conclusions.

Additionally, the bitterness of protein hydrolysates has been suggested to reduce consumption in companion animals potentially. As referred above bitterness has been

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4 associated with the molecular weight of peptides [176]. Recent advancements regarding
5
6 the reduction of bitterness in FBCPs-derived peptides have been explored [177].
7
8 However, in studies with dogs, high palatability has been reported for fish protein
9
10 hydrolysates as pet food ingredients [11][15]. These findings may be explained by the fact
11
12 that pets have a distinct sensitivity to bitterness compared to humans and are likely less
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14 sensitive to bitterness overall, or at least to certain bitter compounds. Research assessing
15
16 the bitter taste receptors (Tas2rs) of dogs' receptive range has revealed a reduced
17
18 sensitivity to bitter compounds relative to humans [178]. Although fish hydrolysates and
19
20 peptides have not yet been specifically analyzed, palatability studies suggest that
21
22 bitterness is a less significant issue in pet food applications than in other sectors.
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25 Animal protein hydrolysates can also exert satiety effects [179], and although
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27 scarcely studied, Theysgeur et al. [112], using an *in vitro* simulated dog gastrointestinal
28
29 digestion model, found stimulation of cholecystokinin and glucagon-like peptide 1
30
31 secretion and inhibition of the dipeptidyl peptidase IV activity with a tilapia FBCPHs.
32

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34 Adverse food reactions involve non-immune (food intolerance) and immune
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36 mechanisms (food hypersensitivity), and their diagnosis is often performed through an
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38 elimination diet with novel protein sources or hydrolyzed proteins. As dogs are exposed
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40 to a wide variety of protein sources, it is not easy to identify a novel protein source;
41
42 thus, hydrolyzed proteins are increasingly considered. Indeed, by disrupting the protein
43
44 structure, protein hydrolysis removes existing allergens and allergenic epitopes,
45
46 preventing an immune response even by animals sensitized to the intact protein [180].
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48 Therefore, protein hydrolysates may play a role in preventing hypersensitivity in
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50 individuals who are at risk or already sensitized.
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53 Although studies with chicken hydrolysates [181, 182] have reported benefits when
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55 used as an elimination diet for diagnosing and treating adverse food reactions in dogs,
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4 results in conflict with allergic reactions reported in other studies ^[183]. Peptide size can
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6 contribute to these conflicting results as it conditions immunoglobulin (Ig) E-mediated
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8 reactions. Indeed, IgE recognizes protein allergens with low MW ^[184], suggesting that
9
10 hydrolyzed proteins with MW lower than 5 kDa should be used in elimination diets ^[185].
11
12 However, Bizikova and Olivry^[183] reported no induced hypersensitivity in dogs fed a
13
14 hydrolyzed poultry feather meal containing 95% hydrolyzed proteins with MW ≤ 1
15
16 kDa, whereas 78% hydrolyzed chicken liver proteins with MW ≤ 1 kDa induced food
17
18 reactions in 40% of the dogs. Additionally, a recent study ^[185] suggested that hydrolyzed
19
20 diets might contain proteins that stimulate helper T-lymphocytes, thus making them
21
22 inadequate for treating food hypersensitivity in all situations. Studies available on dogs
23
24 fed commercial hypoallergenic diets with partially hydrolyzed salmon ^[171] and fish
25
26 protein hydrolysate ^[186] reported a reduction in pruritus and the extent and severity of
27
28 atopic dermatitis, being thus considered to be useful for adverse food reaction diagnosis.
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32 Hydrolyzed protein diets have also been considered to manage inflammatory
33
34 bowel disease, an immunologically mediated intestinal disorder and one of the most
35
36 common dog gastrointestinal diseases ^[187]. Marks et al. ^[188] reported the resolution of
37
38 clinical signs in dogs with inflammatory bowel disease fed a commercially
39
40 hypoallergenic diet containing an enzymatically hydrolyzed defatted soy globulin as the
41
42 only protein source, and Ambrosini et al. ^[189] found improved intestinal membrane
43
44 integrity in dogs with inflammatory bowel disease fed a commercial hydrolyzed diet.
45
46 However, to the best of the author's knowledge, scientific studies with FBCPHs in dogs
47
48 with inflammatory bowel disease are lacking.
49

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51 Hydrolyzed diets are highly digestible and present reduced antigenicity; they
52
53 have also been considered interesting in managing exocrine pancreatic insufficiency,
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55 mainly when associated with dermatological disease. Indeed, despite the likelihood that
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4 endopeptidases from the intestinal brush border compensate for the loss of pancreatic
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6 proteases, adverse food reactions have been reported in dogs with exocrine pancreatic
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8 insufficiency [190]. Studies with FBCPHs to manage exocrine pancreatic insufficiency in
9
10 dogs were not found, but a study with a limited number of dogs (three German
11
12 Sheperds) [191] found improvement in dermatological signs of dogs with exocrine
13
14 pancreatic insufficiency fed a soy and chicken-based-hydrolysate diet.

15
16
17 Even though FBCPHs have been reported to improve the clinical signs of dogs,
18
19 namely reducing the intensity of chronic otitis externa [192], osteoarthritis severity [193],
20
21 joint degeneration and spondylitis deformans [174] and might have a role in the treatment
22
23 of keratoconjunctivitis sicca [194], it is impossible to consider the FBCPHs as the sole
24
25 responsible for the reported improvements due to the complexity of the dietary
26
27 treatments.

28
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30 Regarding effects on immune function, Zinn et al. [195] demonstrated that the
31
32 dietary inclusion of pink salmon hydrolysate (20%) in replacement of poultry by-
33
34 product meal does not dramatically impact the immune function of healthy, senior dogs.
35
36 Authors suggested that the lack of an effect may have resulted from the short length of
37
38 the treatment period (26 days) and that FBCPs may more effectively affect immune
39
40 function in diseased or challenged animals. Similarly, feeding adult Beagle dogs for 6
41
42 weeks with a diet including fish hydrolysate and oil from FBCPs in replacement of
43
44 shrimp hydrolysate and salmon oil did not affect systemic inflammatory markers,
45
46 cardiac structure, and function. However, it potentially benefited bacterial genera
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48 associated with a healthy microbiome [196]. However, it is not easy to distinguish the
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50 effects of protein hydrolysates and oil sources.

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53 Stress associated with fear and anxiety adversely affects animal health, and
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55 related behaviors can negatively affect human-pet relationships [197, 198]. Pet owners are
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4 reluctant to use psychotropic drugs for anxiety treatment, and despite several natural
5
6 products being marketed for behavior therapy, very few have unequivocal efficacy.
7
8 Animal protein hydrolysates may include opioid-like peptides that can act as anxiolytic
9
10 agents [199]. Indeed, bovine α 1-casein hydrolysate decreased anxious disorders [200, 201],
11
12 and caseinate hydrolysate alleviated stress in dogs [202]. Regarding the use of FPH
13
14 supplements, Landsberg et al. [145] reported benefits through decreased hyperactivity and
15
16 cortisol responses, while Titeux et al. [144] observed the promotion of dog-human
17
18 interactions and reduced subtle stress behaviors.
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24 -----Table 4-----
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27 The scarce studies available in the literature, along with the author's experience
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29 with the practical use of fish hydrolysates, support the potential of these feed resources
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31 in dog diets (Figure 5). Indeed, due to their high protein content, aminoacids profile,
32
33 high digestibility, and attractive taste, they are advantageous to be included in high
34
35 digestible and palatable diets. In addition, their effects on satiety make these ingredients
36
37 useful in diets for body weight control. Due to their novelty and peptide size, they are
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39 commonly used in elimination diets to manage adverse food reactions and in managing
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41 dermatological diseases. Finally, the eventual presence of opioid-like peptides might
42
43 play an important role in reducing anxiety and stress-related behaviors, improving the
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45 dogs' welfare and the human-dog relationship.
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49 However, more studies are needed to fully disclose the potential of FBCPHs as
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51 functional ingredients in pet food, even because the amino acid and peptide profiles of
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53 protein hydrolysates differ according to the fish source, enzyme source, and hydrolysis
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4 conditions [203–205], thus impacting their functionality. This information is essential to
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6 fully taking advantage of these resources in pet food.
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10 -----Figure 5-----
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14 Overall, FBCPHs represent a valuable and sustainable resource for pet food
15 formulations due to their high protein content, digestibility, palatability, and bioactive
16 properties. These ingredients are particularly beneficial for managing adverse food
17 reactions, dermatological issues, satiety, and stress-related behaviors in pets, making
18 them ideal for premium and functional diets. However, additional research is required to
19 explore their potential fully, as amino acid and peptide profiles of FBCPs vary with fish
20 source and hydrolysis conditions, affecting their bioactivity. The development of
21 standardized production methods is crucial to improving the reproducibility of
22 FBCPHs' bioactivity and consequently expanding their application in the pet food
23 industry. The growing trend towards personalized pet nutrition presents opportunities to
24 tailor FBCPHs for specific health needs while combining them with other bioactive
25 compounds like prebiotics and probiotics, which could further enhance gut health and
26 immune function. These future research pathways, combined with efforts to optimize
27 production processes and improve scalability, will contribute to developing innovative,
28 sustainable ingredients from FBCPs, supporting both the circular economy and
29 environmental sustainability in the pet food and fish industries.
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6. Conclusions

The fish processing industry faces the challenge of managing increasing amounts of FBCPs, which have significantly contributed to the sector's environmental and economic inefficiencies. FBCPHs represent a promising and sustainable solution, emerging as valuable functional ingredients for pet food. These hydrolysates provide numerous advantages, including high digestibility, essential amino acids, and potential bioactive properties, making them effective for addressing pet health concerns such as allergies, gastrointestinal issues, and anxiety-related behaviors. Their application in premium and functional diets highlights their versatility and value in pet nutrition.

However, to maximize the potential of FBCPH, further research is essential to optimize hydrolysis and extraction processes. This includes exploring various approaches, such as FBCP pretreatment techniques, enzyme selection, hydrolysis parameter optimization, utilization of novel technologies, and even the integration of renewable energy sources. Besides process optimization, the development of standardized production methods is crucial to enhance the yield, quality, and reproducibility of BCPHs' bioactivities, ensuring their effective and seamless application in pet food formulations.

While preliminary studies demonstrated the positive effects of FBCPHs in pet food formulations, more comprehensive research is needed to confirm these benefits, establish them, and ensure optimal efficacy in formulations. Additionally, regulatory considerations must be addressed to ensure compliance as these ingredients are integrated into the market. Despite all these challenges, FBCPHs align with the growing trend towards premium and health-focused pet foods. Future research will be crucial in improving FBCPH production, incorporating renewable energy, ensuring bioactivity, and meeting regulatory standards, all while addressing pet owners' expectations for high-quality, functional nutrition.

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Disclosure statement

The authors report that there are no competing interests to declare.

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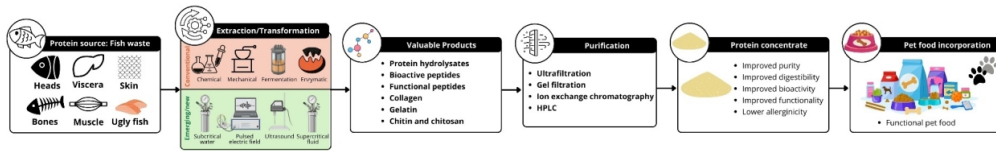


Figure 1. Summary of the main goals of the present review work.

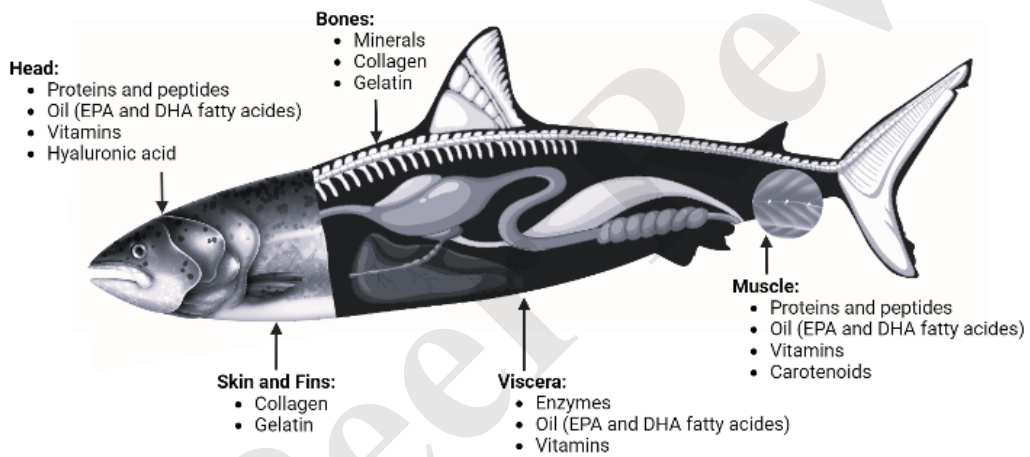


Figure 2. Fish by- and co-products (FBCPs) and their composition. Created with BioRender.com

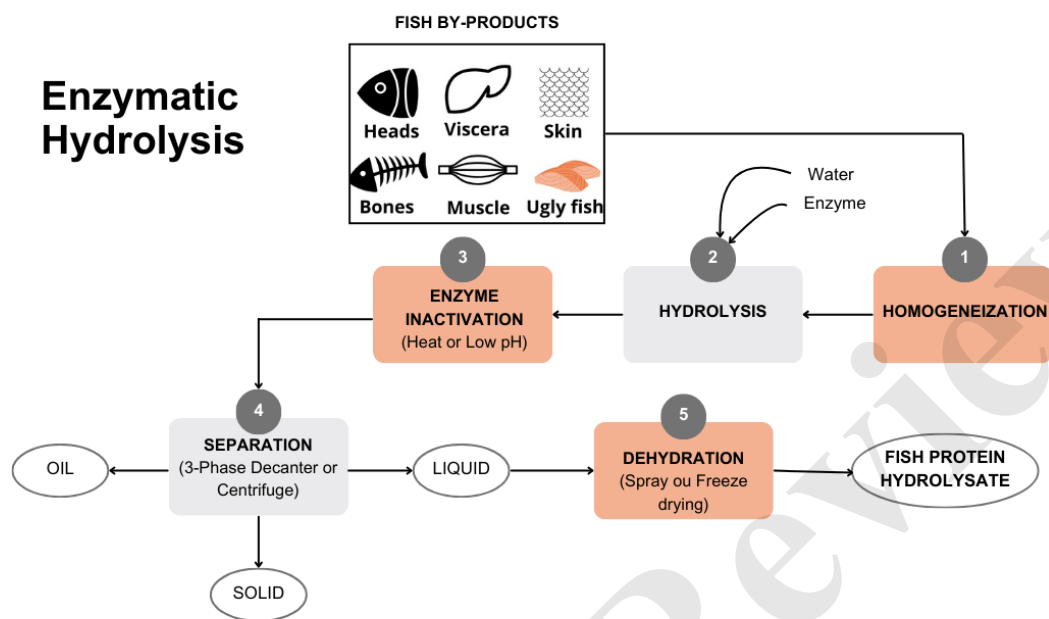


Figure 3. Schematic representation of the enzymatic hydrolysis process of fish by- and co-products (FBCPs).

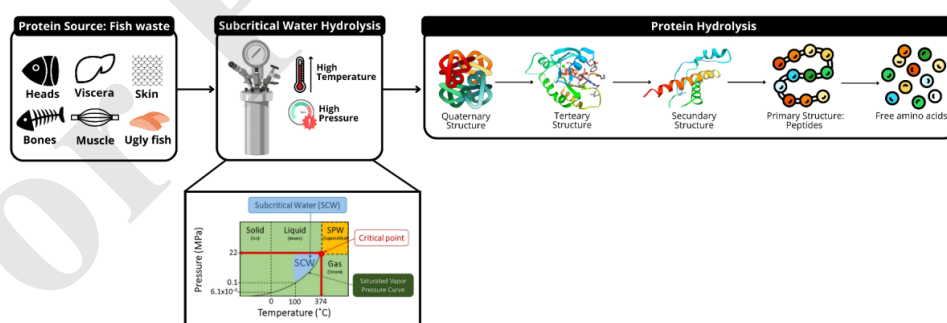


Figure 4. Schematic representation of the technique subcritical water hydrolysis (SWH) of fish by- and co-products (FBCPs).

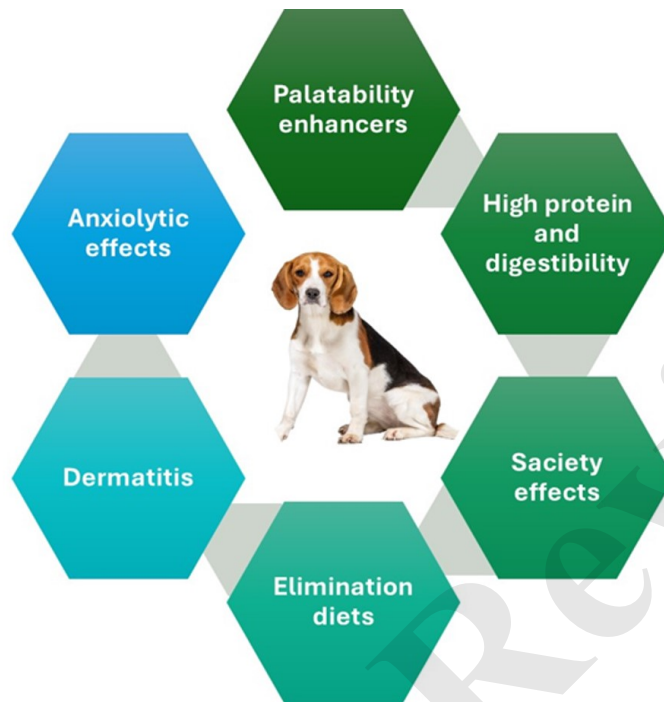


Figure 5. Potential effects of hydrolysates from fish by-products and co-products (FBCPs) on dog nutritional and health parameters.

Table 1. List of EU legislation concerning pet food safety and labeling and claims.

Scope	Legislation applied	Description
Pet food safety	Regulation 178/2002	General principles and requirements of food law (which also applies to feed)
	Regulation 183/2005	Requirements for feed hygiene
	Regulation 1069/2009	Animal by-product regulations with detailed rules on the safety of raw materials of animal origin used in pet food, their processing requirements, and model health certificates for imports to the EU
	Regulation 142/2011	
	Regulation 999/2001	Rules for the prevention, control, and eradication of certain transmissible spongiform encephalopathies
	Regulation 1831/2003	Requirements on additives for use in animal nutrition; all approved additives are listed in the Register published by the EU Commission
	Directive 2002/32	Directive dealing with undesirable substances in feed, which sets maximum limits for contaminants, with specifications for raw materials and finished feeds
Labeling and Claims	Regulation 767/2009	Rules for labeling, claims, and other forms of marketing communication
	Directive 2008/38	List of all approved PARNUT (products for particular nutritional purposes) indications

Table 2. Different types of functional claims and their general requirements are according to the FEDIAF Code ^[169].

Functional claims	General requirements
Nutrient function claims	<p>Simply links the presence of a nutrient or combination of nutrients in a product to the physiological role in the body's growth, development, and normal functions, without any further detail about the effect's level or degree/mechanism.</p>
Enhanced function claims	<p>Describes the specific beneficial effect of nutrients or other substances, alone or in combination, on physiological functions or biological activities in the body. Enhanced function means an effect that either exceeds its usual role in maintaining normal metabolic functions, including growth and development, or is related to a substance that is not essential for the animal but provides a benefit beyond nutrition. No reference should be made to particular diseases or pathological states.</p>
Health maintenance and decreased disease risk claims	<p>Related to the optimization of nutrition and the support or protection of physiological conditions. It can also be related to health maintenance and reducing the risk of disease development resulting from nutritional imbalances in a healthy animal.</p> <p>Such claims relate to the consumption of a product containing a nutrient or other substances, alone or in combination, that helps to reduce the risk of disease development or maintain physiological functions or health.</p> <p>Claims referring to the treatment or curing of a disease are considered medicinal claims and would cause a product to be medicinal by presentation. However, words such as prevent may be used if not related to a disease treatment. The Code provides a guideline with some examples of words that may be avoided since they are usually associated with authorized medicinal products.</p>

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	<p>When using substances supporting health maintenance and decreased disease risk claims, operators shall ensure that the relevant substance is properly classified as an additive, a feed material, or a veterinary medicinal product; if it is classified as a veterinary medicinal product, it may not be used in pet food.</p> <p>In case of unclear classification, the Commission Guidelines shall be consulted.</p>
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For Peer Review

Table 3. Studies of functional foods and diets containing fish hydrolysates in cats.

Animal species/model	Functional food/diet	Inclusion level	Study duration	Effects	Reference
Cats with allergic dermatitis (n= 12; 7 females, 5 males; 2-12 years)	Partially hydrolyzed salmon and pea commercial hypoallergenic diet	85% hydrolyzed salmon	2, 4, 6, 8, and 10 weeks	Partially hydrolyzed salmon and pea diet reduced pruritus and clinical symptoms in all animals after 8 weeks, showing to be useful for feline adverse food reaction diagnosis and treatment	[171]
Nonseasonally pruritic cats (n=32; 22 females, 10 male; 0.4-14 years)	Hydrolyzed fish protein and rice starch commercial hypoallergenic diet	No information available	56 days, followed by a challenge with the previous diet	Hydrolyzed fish protein and rice starch diet reduced pruritus in 77% of cats, showing to be useful for feline adverse food reaction diagnosis	[172]
Healthy cats (n=10; 5 males, 5 females, 3-5 years)	Prescription diet with L-tryptophan and supplemented with fish peptides (from sardines).	Supplement contained 0.1% fish peptides plus 0.1% lemon balm (minimum 5%	10 weeks supplementation	Supplementation of fish peptides, lemon balm, and oligofructose reduced the average 24-hour urinary cortisol/creatinine ratio, a stress marker in cats, and increased serotonin. Overall effects	[173]

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	lemon balm extract, and oligofructoset	rosmarinic acid) and 0.5% oligofructose		highlight its potential use in cats suffering from mild stress-related conditions.	
Cats with lameness caused by orthopedic diseases and spondylitis deformans (n=3)	Collagen peptide (from fish scales) and D-glucosamine (from crabs) supplement	Oral administration of 1 g each/animal/day, mixed with food	1, 2 and 3 months	Simultaneous administration of collagen peptide and D-glucosamine supplements from marine origin was effective in the recovery of several joint degeneration and spondylitis deformans in dogs after one month. The absence of side effects suggests the potential for long-term administration	[174]

Table 4. Studies of functional foods and diets containing fish hydrolysates in dogs.

Animal species/model	Functional food/diet	Inclusion level	Study duration	Effects	Reference
Healthy adult dogs (n=20; 10 Beagle, 10 pointers; 7.9-32.8 kg BW)	Salmon hydrolysate-supplemented diet	10% salmon protein hydrolysate	2 days	10% hydrolyzed salmon protein increased the consumption ratio compared to a chicken-based control diet, showing to be highly palatable to dogs	[11]
Healthy Beagle dogs (n=12; 6 females, 6 males; 5.4 ± 0.57 years; 11.8 ± 2.20 kg BW; BSC of 4.3 ± 0.69)	Fish hydrolysate and fish oil (from fish waste) diet	5% fish hydrolysate and 3.2% fish oil	2 periods of 6 weeks (crossover design)	Fish hydrolysate and oil diet increased intake of EPA and DHA without affecting palatability, digestibility, and coat quality.	[10]
<i>in vitro</i> static gut model of dog digestion	Tilapia byproduct protein hydrolysate	2 g of hydrolysate as sole substrate	2 min oral digestion, 2 h gastric digestion, 4 h	<i>In vitro</i> digestion of tilapia byproduct hydrolysate released several peptides with potential bioactive activity that may be involved in the regulation of food intake, glucose metabolism, intestinal hormones secretion, and dipeptidyl peptidase IV inhibitory activity, suggesting to exert satiety effects	[12]

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			intestinal digestion		
Dogs with atopic dermatitis signs (n=13)	Partially hydrolyzed salmon and pea commercial hypoallergenic diet	85% hydrolyzed salmon	2, 4, 6, 8, and 10 weeks	Partially hydrolyzed salmon and pea diet reduced pruritus and clinical symptoms in all animals after 10 weeks, showing to be useful for canine adverse food reaction diagnosis and treatment	[171]
Nonseasonally pruritic dogs (n=50; 24 females, 26 males; 0.4-14 years old)	Hydrolyzed fish protein and rice starch diet commercial hypoallergenic diet	No information available	56 days, followed by a challenge with the previous diet	Hydrolyzed fish protein and rice starch diet reduced pruritus in 63% of dogs, showing to be useful for adverse food reaction diagnosis, even in dogs allergic to fish and rice	[186]

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<p>Dog with chronic clinical otitis symptoms (n=30; 14 females, 16 males; 6.0 ± 0.15 years; 32.0 ± 1.17 kg BW)</p>	<p>Commercial diet (93-94%) supplemented with therapeutical microcapsules (6-7%) with fish and vegetable hydrolysates</p>	<p>Microcapsules contained 60-80% fish and vegetable hydrolysates, minerals, and 20-40% natural substances (<i>Melaleuca alternifolia</i>, <i>Tilia platyphyllos scapoli et cordata</i>, <i>Allium sativum</i>, <i>Rosa canina</i>, and zinc)</p>	<p>90 days</p>	<p>Supplementation of hydrolyzed fish and vegetable-based microcapsules relieved the intensity of otitis externa-related symptoms in combination with topical drugs, suggesting the efficacy of combined dietary and conventional drug therapy</p>	<p>[192]</p>
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<p>Healthy Labrador Retriever puppies (n=42)</p>	<p>Fish meal-based commercial diet (93-94%) and nutraceutical supplement (6-7%) with fish and vegetable hydrolysates</p>	<p>Supplement contained hydrolyzed fish and vegetable protein, supplemented with glucosamine, chondroitin sulfate, chitosamine, <i>Boswellia serrata</i>, <i>Harpagophytum procumbens</i>, green-lipped mussel, and fish-oil omega-3/6</p>	<p>From 3 to 12 months of age</p>	<p>Nutraceutical supplement with fish and vegetable hydrolysates did not affect the prevalence of hip and elbow dysplasia but reduced osteoarthritis severity at 12 months of age, showing to have beneficial effects on severe osteoarthritis development</p>	<p>[193]</p>
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		polyunsaturated fatty acids (1:1 ratio)			
Dogs with lameness caused by orthopedic diseases and spondylitis deformans (n=68)	Collagen peptide (from fish scales) and D-glucosamine (from crabs) supplement	Oral administration of 1 g each/animal/day, mixed with food	1, 2 and 3 months	Simultaneous administration of collagen peptide and D-glucosamine supplements from marine origin was effective in the recovery of several joint degeneration and spondylitis deformans in dogs after one month. The absence of side effects suggests the potential for long-term administration	[174]
Dogs with immune-mediated keratoconjunctivitis sicca (n=50; 19 females, 31 males; 6.5 ± 0.7 years)	Commercial diet (93-94%) supplemented with nutraceutical tablets (6-7%)	Tablets contained 60-80% fish and vegetable hydrolysates, minerals, and	60 days	Supplementation of hydrolyzed fish and vegetable-based tablets combined with immunosuppressive therapy reduce the immune-mediated ocular symptoms of keratoconjunctivitis sicca-affected dogs that had poor or no response to classical immunosuppressive drugs, unveiling its potential immune modulation effect	[194]

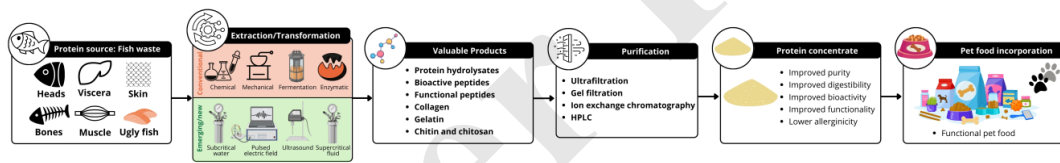
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	with fish and vegetable hydrolysates	20-40% botanical substances			
Healthy senior pointer dogs (n=12; 2 female, 10 male; 7.9 ± 1.24 years; 24.0 ± 2.4 kg BW)	Hydrolyzed pink salmon-supplemented diet	20% pink salmon hydrolysate	26 days	Pink salmon hydrolysate at 20% inclusion had no negative effect on the palatability, nutrient digestibility, and immune function of healthy senior dogs, revealing its potential as an alternative ingredient	[195]
Healthy Beagle dogs (n=12; 6 females, 6 males; 5.4 ± 0.57 years; 11.8 ± 2.20 kg BW; BSC of 4.3 ± 0.69)	Fish hydrolysate and fish oil (from fish waste) diet	5% fish hydrolysate and 3.2% fish oil	2 periods of 6 weeks (crossover design)	Fish hydrolysate and oil diet decreased plasma triglycerides and angiotensin-converting enzyme activity, while no negative impact on systemic inflammation markers, cardiac structure, and function was observed. Fecal microbiome modulation towards health-promoting bacterial genera suggests the potential of a functional diet	[196]
Adult Beagle dogs (n=45; female and male; 2.7-17 years)	Fish hydrolysate capsules	750 and 1500 mg/day	35 days	Fish hydrolysate supplementation reduced hyperactivity response and cortisol response, supporting its use to reduce dogs' fear and anxiety	[145]

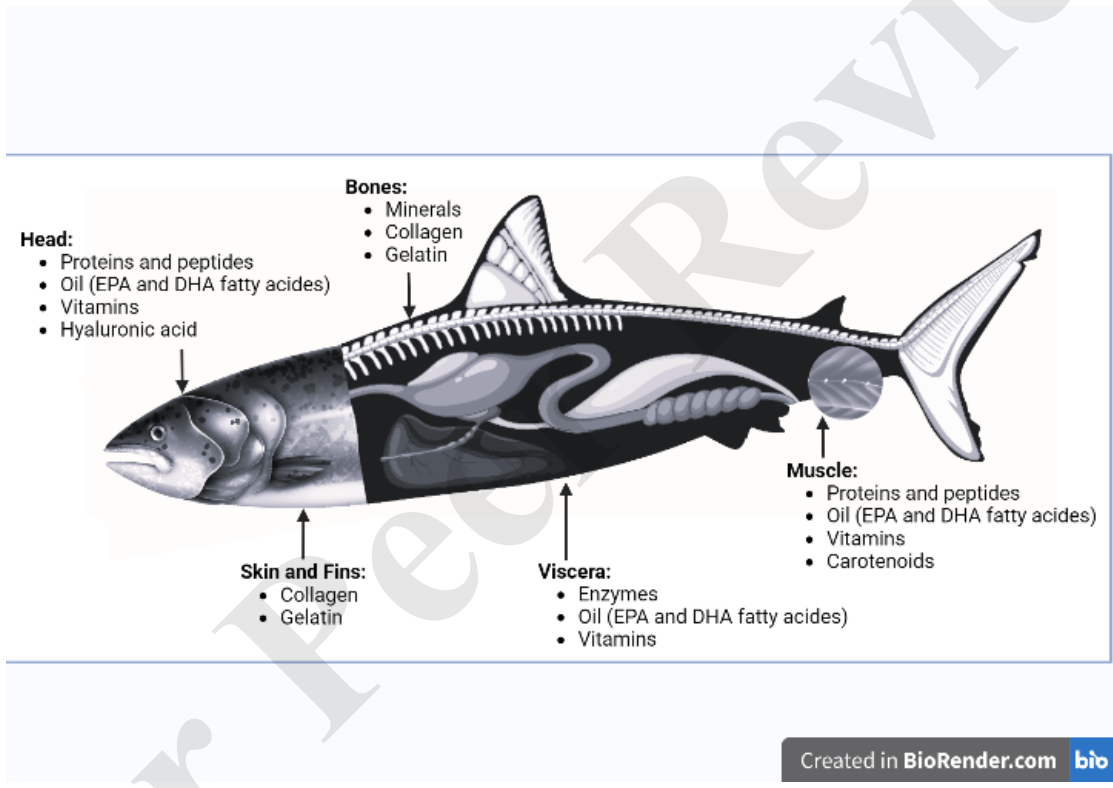
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<p>Dogs of several breeds (29) and crossbreeds (n=39; 26 females, 13 males; 1-6 years)</p>	<p>Fish hydrolysate capsules (500 mg GABOLYSAT PTP 55, 11 mg of SOD B Primo-antioxidant® M, 5 IU/mg)</p>	<p>Less than 10 kg BW: 1 capsule/day More than 10 kg BW: 2 capsules/day</p>	<p>30 days</p>	<p>Fish hydrolysate supplement promoted dog-human interactions and tend to reduce subtle stress behaviors, suggesting being effective in situations of mild stressors</p>	<p>[144]</p>
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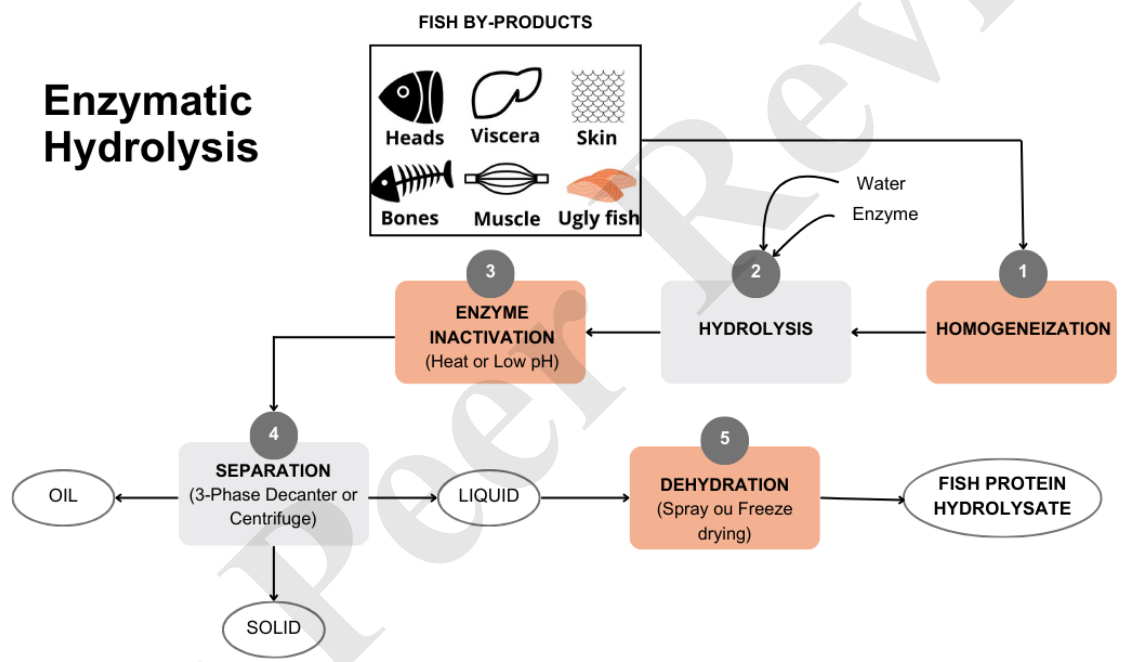


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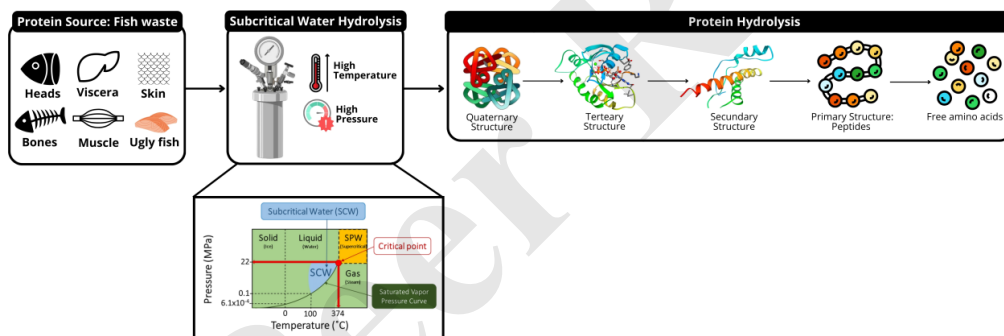


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Enzymatic Hydrolysis



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