



## Acromegaly in humans and cats: Pathophysiological, clinical and management resemblances and differences

Mariana Lopes-Pinto<sup>a</sup>, Patrícia Lunet Marques<sup>b,c</sup>, Ema Lacerda-Nobre<sup>a</sup>, Diego Miceli<sup>d</sup>, Rodolfo Oliveira Leal<sup>b,c</sup>, Pedro Marques<sup>e,f,\*</sup>

<sup>a</sup> Endocrinology Department, Unidade Local de Saúde de Santa Maria, Portugal

<sup>b</sup> CIISA – Centre for Interdisciplinary Research in Animal Health, Faculty of Veterinary Medicine, University of Lisbon, Portugal

<sup>c</sup> Associate Laboratory for Animal and Veterinary Sciences (ALAAV), Portugal

<sup>d</sup> IBYME - Institute of Experimental Biology and Medicine (CONICET), Buenos Aires, Argentina

<sup>e</sup> Pituitary Tumor Unit, Endocrinology Department, Hospital CUF Descobertas, Lisbon, Portugal

<sup>f</sup> Faculdade de Medicina, Universidade Católica Portuguesa, Lisbon, Portugal

### ARTICLE INFO

#### Keywords:

Acromegaly  
Gigantism  
Growth hormone (GH)  
Insulin-like growth factor 1 (IGF-1)  
Hypersomatotropism  
Cats

### ABSTRACT

**Objective:** Acromegaly is a disorder associated with excessive levels of growth hormone (GH) and insulin-like growth factor-1 (IGF-1). In general, GH/IGF-1 excess leads to morphologic craniofacial and acral changes as well as cardiometabolic complications, but the phenotypic changes and clinical presentation of acromegaly differ across species. Here, we review the pathophysiology, clinical presentation and management of acromegaly in humans and cats, and we provide a systematic comparison between this disease across these different species.

**Design:** A comprehensive literature review of pathophysiology, clinical features, diagnosis and management of acromegaly in humans and in cats was performed.

**Results:** Acromegaly is associated with prominent craniofacial changes in both species: frontal bossing, enlarged nose, ears and lips, and protuberant cheekbones are typically encountered in humans, whereas increased width of the head and skull enlargement are commonly found in cats. Malocclusion, prognathism, dental diastema and upper airway obstruction by soft tissue enlargement are reported in both species, as well as continuous growth and widening of extremities resulting in osteoarticular compromise. Increase of articular joint cartilage thickness, vertebral fractures and spine malalignment is more evident in humans, while arthropathy and spondylosis deformans may also occur in cats. Generalized organomegaly is equally observed in both species. Other similarities between humans and cats with acromegaly include heart failure, ventricular hypertrophy, diabetes mellitus, and an overall increased cardiometabolic risk. In GH-secreting pituitary tumours, local compressive effects and behavioral changes are mostly observed in humans, but also present in cats. Cutis verticis gyrata and skin tags are exclusively found in humans, while palmigrade/plantigrade stance may occur in some acromegalic cats.

Serum IGF-1 is used for acromegaly diagnosis in both species, but an oral glucose tolerance test with GH measurement is only useful in humans, as glucose load does not inhibit GH secretion in cats. Imaging studies are regularly performed in both species after biochemical diagnosis of acromegaly. Hypophysectomy is the first line treatment for humans and cats, although not always available in veterinary medicine.

**Conclusion:** Acromegaly in humans and cats has substantial similarities, as a result of common pathophysiological mechanisms, however species-specific features may be found.

**Abbreviations:** AIP, Aryl hydrocarbon receptor interacting protein; cDNA, Complementary deoxyribonucleotidic acid; DNA, Deoxyribonucleotidic acid; CT, Computerized tomography; GH, Growth hormone; GHRH, Growth hormone releasing hormone; GNAS, Guanine nucleotide binding protein alpha stimulating activity polypeptide; GPR, G-protein coupled receptor; IGF-1, Insulin-like growth factor 1; MAX, Myc-associated factor X; MEN1, Multiple endocrine neoplasia type 1; MRI, Magnetic resonance imaging; PIIP, Type III procollagen propeptide; SDHx, Succinate dehydrogenase; SRT, Stereotactic radiation therapy; SSTR, Somatostatin receptor.

\* Corresponding author at: Pituitary Tumor Unit, Endocrinology Department, Hospital CUF Descobertas, Lisbon, Portugal.

E-mail address: [pedro.miguel.sousa.marques@gmail.com](mailto:pedro.miguel.sousa.marques@gmail.com) (P. Marques).

<https://doi.org/10.1016/j.ghir.2024.101595>

Received 10 January 2024; Received in revised form 28 April 2024; Accepted 20 May 2024

Available online 24 May 2024

1096-6374/© 2024 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

The word “acromegaly” was coined by Pierre Marie, in the late 19th century from the Greek “akron”, meaning ‘tip or extremity’, and “megas”, meaning ‘great’. However, reports of clinical signs compatible with acromegaly have been made as early as 1567 [1]. In cats, the first report dates from 1976 [2]. Acromegaly, or hypersomatotropism, is an endocrine disease that can affect both humans and cats. Acromegaly is characterized by an excessive production of growth hormone (GH), usually by a GH-secreting pituitary tumour in both species. In cats, a clear distinction between hypersomatotropism and acromegaly is usually made. While hypersomatotropism implies only a state of excess GH, acromegaly refers to the syndrome and associated clinical signs that result from GH excess. Hypersomatotropism may or may not result in acromegaly in cats, especially in the earlier stages of the disease [3].

Chronic excessive GH levels lead to increased synthesis of insulin-like growth factor 1 (IGF-1) by the liver [4]. High IGF-1 concentrations promote somatic overgrowth, which culminates in a constellation of signs and symptoms, including several acromegaly-related morphologic features, as well as cardiometabolic and skeletal complications [5].

We aimed to review the pathophysiology and clinical features of acromegaly in humans and cats in a systematic comparative manner, highlighting the overlapping and distinctive features of the disease in both species. Additionally, by identifying important similarities, we aimed to assess the potential use of cats as the animal model *par excellence* for the study of acromegaly in human medicine and gain further insights into this complex disease. On the other hand, the current knowledge of the disease in humans, reviewed here, can also benefit the understanding and management of acromegaly in cats.

## 2. Epidemiology and pathophysiology

### 2.1. Humans

Acromegaly is a rare disease affecting humans, with an estimated annual incidence of 7.7–10 cases per million population per year, and with a prevalence ranging from 3.6 to 13 cases per 100,000 individuals [6–10]. Acromegaly is mostly diagnosed during the 5th decade of life, and is associated with an increased mortality risk when untreated, particularly in older patients and in women [11–15].

Over 95% of patients diagnosed with acromegaly harbour a GH-secreting pituitary tumour, which stimulates the synthesis of IGF-1 by the liver, leading to somatic overgrowth and several acromegaly-related manifestations, such as cardiometabolic and skeletal complications [6,9]. Such pituitary tumours commonly arise from somatotroph cells, and may display different granulation patterns: densely-granulated, sparsely-granulated or mixed pattern. Densely-granulated somatotroph tumours represent approximately 50% of the cases, and are typically smaller and respond better to somatostatin analogues, whereas sparsely-granulated somatotroph tumours are more common in younger individuals and are often associated with an invasive behaviour as well as with poorer response to medical therapy [6,9,15]. Somatic mutations in the guanine nucleotide binding protein alpha stimulating activity polypeptide gene (*GNAS*) are responsible for 40% of sporadic GH-secreting pituitary tumours [16,17]. Environmental factors may also contribute for the onset of this disease in humans, as suggested by data from an Italian study reporting increased prevalence of acromegaly in highly-polluted areas [18].

Acromegaly may also be caused by the ectopic hypersecretion of GH or Growth Hormone-Releasing Hormone (GHRH) by a hypothalamic tumour or by a neuroendocrine tumour [6,19].

Acromegaly may occur in a familial setting, as part of a syndrome or in isolation. The syndromic forms of acromegaly include: i) multiple endocrine neoplasia type 1 (MEN1), most typically due to GH-secreting or GH and prolactin-secreting pituitary tumours, concomitantly with primary hyperparathyroidism and pancreatic neuroendocrine tumours,

but GH or GHRH excess may also derive from a MEN1-related neuroendocrine tumour [20,21]; ii) MEN type 4 iii) “3PAs syndrome” due to mutations in the genes for succinate dehydrogenase (*SDHx*) or Myc-associated factor X (*MAX*); iv) Carney complex, which is associated with pituitary hyperplasia, mucosal and atrial myxomas, adrenal hyperplasia and hyperpigmentation; and also, iv) McCune-Albright syndrome, a hypersecretory endocrinopathy which comprises pituitary hyperplasia or adenoma, hyperthyroidism, hypercortisolism, hyperprolactinemia, cafe-au-lait pigmentation and polyostotic fibrous dysplasia. In contrast, familial acromegaly or gigantism may occur in isolation due to mutations in the aryl hydrocarbon receptor-interacting protein (*AIP*) [22], or more rarely, in the setting of X-linked acro-gigantism due to G-protein coupled receptor 101 gene (*GPR101*) micro-duplications [6,23–27].

### 2.2. Cats

Hypersomatotropism is thought to affect 1 in 800 cats in the United Kingdom, thus being about 10 times more prevalent than in humans [28,29]. Feline hypersomatotropism is closely related with diabetes mellitus, with about 15–25% of diabetic cats exhibiting excessive levels of IGF-1 [28,30]. Earlier studies reported that 100% of cats with hypersomatotropism had diabetes, however it is currently known that hypersomatotropism may also occur in non-diabetic cats [31,32]. Cats with hypersomatotropism tend to be older/geriatric (mean age at diagnosis 11.3 years, for a median lifespan of 14 years, with an inter-quartile range of 9–17 years [33]), with 70% being neutered males; this is similar to cats with diabetes without GH/IGF-1 excess [34].

Like in humans, feline hypersomatotropism usually results from a neoplastic or hyperplastic transformation of somatotroph cells in the anterior pituitary, predominantly in the form of a pituitary adenoma [35]. In cats, such pituitary tumours normally secrete only GH [36], contrasting with humans where prolactin co-secretion may occur in up to 30% of GH-secreting pituitary adenomas [34,37]. Plurihormonal tumours have also been reported in cats, with increased adrenocorticotrophic hormone (ACTH) and  $\alpha$ -melanocyte-stimulating hormone ( $\alpha$ -MSH), as well as increased GH [36,38,39].

Some environmental causes of feline hypersomatotropism have been suggested. Higher levels of organohalogenated contaminants have been found in cats with hypersomatotropism in comparison with healthy cats or diabetic cats, suggesting a possible connection between these endocrine-disrupting chemicals and feline hypersomatotropism [40]. Hypersomatotropism may also decrease the capacity to metabolize persistent chemicals like polychlorinated biphenyls [34,37].

Like in humans, hypersomatotropism in cats may also have a genetic component. In one study, 2 out of 10 cats with hypersomatotropism had a single non-conservative single nucleotide polymorphism in exon 1 of the *AIP* gene, while no *AIP* mutation was present in 10 control cats [41]. MEN-1 like syndrome has also been identified in cats, although no GH excess was detected [42,43]. Thus, genetic causes of hypersomatotropism in cats are yet to be described. [42]

## 3. Clinical features in humans and cats with acromegaly

### 3.1. Facial features

#### 3.1.1. Humans

Clinical features of acromegaly usually show an insidious course, and subtle physical changes may be traced back in up to a decade before the diagnosis [6]. The most common manifestations include facial dysmorphism and acral overgrowth, leading to a particular acromegaloid appearance. A coarse face with frontal bossing, enlarged nose and ears, fleshy lips, prognathism, protuberant cheekbones and wrinkles are typical facial features of acromegaly (Fig. 1a-d) [6,9]. Other features may be found, such as maxillary prominence with dental malocclusion, increased interdental space (i.e. diastema) and macroglossia (Fig. 1c-d).

Besides contributing to esthetical compromise, these changes may also result in partial obstruction of the respiratory tract, leading to obstructive sleep apnoea, excessive snoring and narcolepsy [6,9,44,45]. Progressive hoarse voice in acromegaly results from laryngeal and paranasal sinuses hypertrophy [46].

### 3.1.2. Cats

Like in humans, facial bone growth leads to broad and prominent facial features in cats (Fig. 1e-f), increased width of the head, enlargement of the skull, inferior prognathia (Fig. 1g) and dental diastema (Fig. 1h). In the upper airways, soft tissue growth leads to inspiratory stridor and/or increased reporting of “snoring” by owners in 38% of cats with acromegaly [34,37,47]. In one study from Buenos Aires, physical changes consistent with acromegaly were observed in 61% of diabetic cats with excess GH [30]. In other studies, however, physical changes were only observed in 5–24% of a similar population [48,49]. In a series of 68 diabetic cats with hypersomatotropism, frontal and parietal bones as well as the mandibular rami were thicker, and the distance between the zygomatic arches was greater than in control cats. The skin and subcutis dorsal to the frontal bone, lateral to the zygomatic arch and ventral to the mandibular rami were also thicker in cats with hypersomatotropism. Prognathia inferior and signs of temporo-mandibular joint malformation were more often observed in cats with hypersomatotropism [50].

Broad facial features have also been reported in up to 37% of cats with acromegaly and prognathism in 18% [47].

## 3.2. Extremities

### 3.2.1. Humans

The enlargement of the extremities is a key feature of acromegaly (Fig. 2a-b). Continuous growth and widening of hands and feet, often revealed by the increase in ring and shoe size, should raise suspicion of acromegaly [6,9,51], although there are also other conditions unrelated to GH/IGF-1 that may lead to acral enlargement and/or acromegaloid appearance (pseudoacromegaly) [52,53].

### 3.2.2. Cats

The enlargement of extremities may also occur in cats, with large and clubbed paws, with distal limbs exhibiting a clubbed appearance (Fig. 2c) [34]. Increase in joint space secondary to thickening of the articular cartilage and cortical thickening contribute to enlargement of extremities, as well as periarticular periosteal reaction and osteophyte proliferation [54]. Joint stiffness and pain has been reported in 10% of cats with acromegaly and clubbed extremities in 13% [47].

## 3.3. Skeletal manifestations

### 3.3.1. Humans

Humans with acromegaly often show osteoarticular compromise due to IGF-1 induced increased bone metabolism. Skeletal changes in patients with acromegaly are mainly induced by excessive levels of IGF-1, and typically include: mono or polyarticular arthropathy, with painful, swollen and stiff joints (in about 70% of cases), carpal tunnel syndrome, in response to local fluid retention and wrist swelling (in more than half of patients), generalized increase of articular joint cartilage thickness, ossification of ligaments and vertebral fractures which jeopardize patients’ mobility and quality of life [55–58]. Skeletal deformities may be severe and ultimately change rib cage structure, and spine alignment with dorsal kyphosis scoliosis in response to anteroposterior vertebral growth, widened disc spaces, osteoporosis, and vertebral fractures; calvaria thickening have also been reported in patients with acromegaly (Fig. 3a-d) [55,57–60].

### 3.3.2. Cats

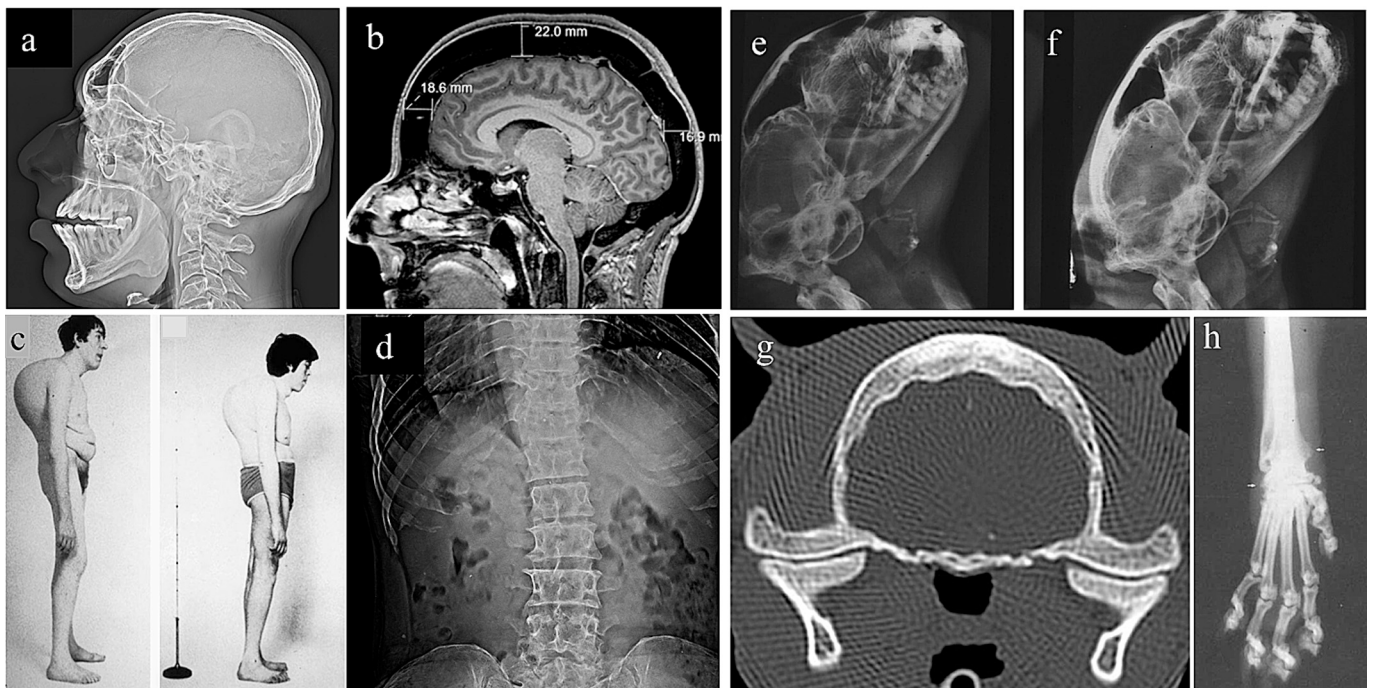
In acromegalic cats, increased bone and cartilage growth can lead to arthropathy and *spondylosis deformans*. Radiographic changes include periarticular periosteal reaction and osteophytes with soft tissue swelling and collapse of the joint spaces (Fig. 3h). Although these findings are compatible with osteoarthritis, lameness is not often described in affected cats [54]. In fact, the pathogenesis of acromegalic arthropathy in cats is thought to be noninflammatory, with cartilage hypertrophy and hyperplasia leading to the disruption of joint geometry and chondrocyte metabolism [54,61]. Thickened skull bone may also be found in acromegalic cats (Fig. 3f-g).



**Fig. 1.** Acromegalic facial appearance in humans (a-d) and cats (e-h). Acromegalic features in humans include large faces (a-b), frontal bossing (b), enlarged nose and ears (a-b), fleshy lips (a-c), prognathism (b), protuberant cheekbones (a-b), dental malocclusion, increased interdental space (c) and macroglossia (d). Acromegalic cats may exhibit similar characteristics, including broad facial features (e, f), prognathism (f,g) and increased interdental space (h). Pictures a-h are unpublished original images.



**Fig. 2.** Acral extremities enlargement in humans (a-b) and cats (c-d) with acromegaly. Patients with acromegaly may present with widened hands (a-b) and feet. Cats with acromegaly present with “clubbed paws” due to acral enlargement of extremities (c-d). The sources from the pictures are: a-c) unpublished original images.



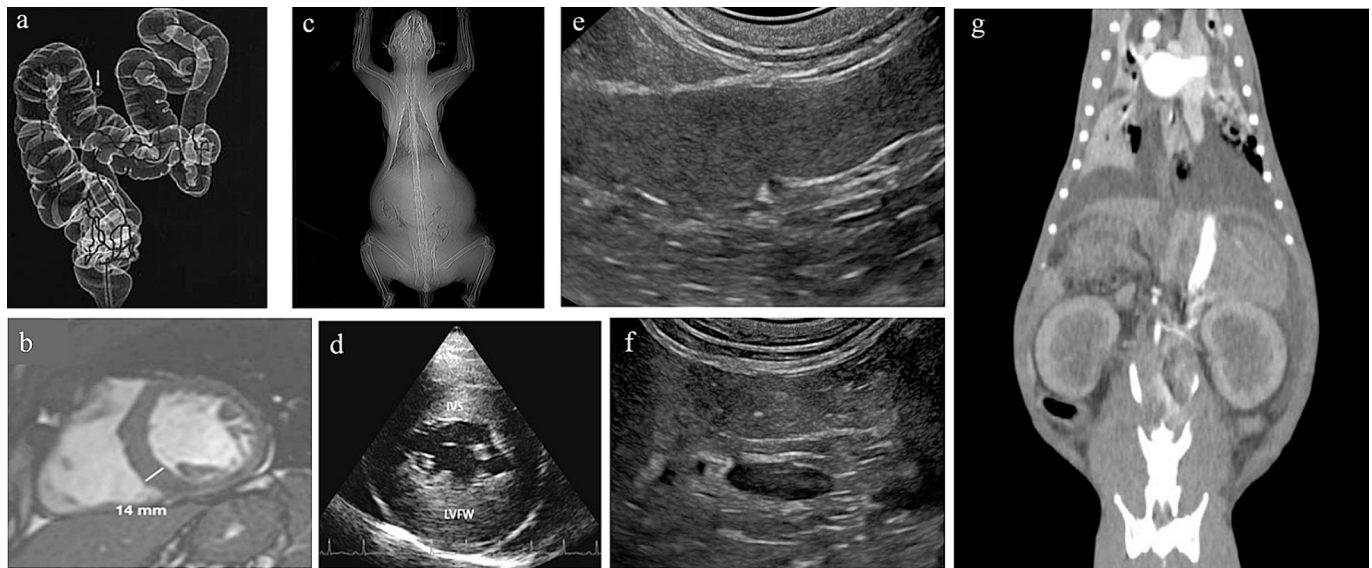
**Fig. 3.** Skeletal features of acromegaly in humans (a-d) and cats (e-h). Skull X-ray and Computed Tomography (CT) scan of patients with acromegaly may show increased calvaria thickness, frontal sinus overgrowth and malocclusion of teeth with prognathism (a-b). Spine alignment may be compromised and dorsal kyphosis scoliosis may occur (c) increased disc-facet and intervertebral joint space with mild kyphosis of lumbar vertebrae, hyperostosis of T11-L3 anterior vertebrae body and posterior scalloping of the lower lumbar vertebrae may be seen in frontal spine X-ray (d). Cats with acromegaly may also present with thickening of the skull, which can be seen on X-ray (skull of the same cat before (e) and after developing acromegaly (d)) or on CT scan (g). Osteoarthritis is also a common finding in acromegalic cats, with associated soft tissue swelling, collapse of joint spaces and periarticular periosteal reaction (h). The sources from the pictures are: a,d) Jones, Acromegaly. Reference article, [Radiopaedia.org, https://doi.org/10.53347/rID-5748](https://doi.org/10.53347/rID-5748) (accessed on 19 Dec 2023); b) Cosme 2023 Med Clin (Barc) [PMID: 36456415]; c) Whitehead 1982 Clin Endocrinol (Oxf) [PMID: 7165969]; e-f) Greco 2012 Top Companion Anim Med [PMID: 22958795]; g) Fracassi 2016 JFMS Open Rep [PMID: 28491423]; h) Peterson 1990 J Vet Intern Med [PMID: 2401966].

### 3.4. Organomegaly

#### 3.4.1. Humans

There is a tendency for generalized organomegaly in patients with acromegaly, often presenting with enlarged thyroid, salivary glands,

tongue, liver, spleen, kidney, heart (Fig. 4b) and prostate. An abnormally long and large colon, termed dolichomegacolon, may also be present and carries the risk of abnormal intestinal rotation and volvulus (Fig. 4a) [62,63].



**Fig. 4.** Organomegaly examples in humans (a-b) and cats (c-g) with acromegaly. Computed tomography (CT) colonoscopy of a patient with acromegaly showed a large colon - dolico-megacolon (a); cardiac magnetic resonance imaging may identify marked left ventricular hypertrophy in acromegaly (b). Cats with acromegaly also present with organomegaly: generalized abdominal distension may be noted on X-ray (c); cardiac ultrasound may show signs of hypertrophic cardiomyopathy, with concentric thickening of the left ventricle (d); ultrasound may show enlarged spleen and adrenal glands (e-f); CT scan may show exuberant enlargement of the kidneys (g). The sources from the pictures are: a) Resmini 2009 J Clin Endocrinol Metab [PMID: 18957501]; b) Bogazzi 2010 J Endocrinol Invest [PMID: 31143230]; c) Niessen 2015 PLoS One [PMID: 26023776]; d) Ferasin 2012 In Practice, <https://doi.org/10.1136/inp.e2271> (accessed on 19 Dec 2023); Pictures e-g are unpublished original images.

### 3.4.2. Cats

Abdominal enlargement may be noted in cats with acromegaly due to GH/IGF-1 excess-related organomegaly (Fig. 4c). Hepatomegaly, splenomegaly, renomegaly, as well as enlargement of the adrenal glands and pancreas have been reported (Fig. 4e-g) [37,64].

## 3.5. Cardiovascular and metabolic comorbidities

### 3.5.1. Humans

Cardiovascular disease represents a major cause of morbidity and mortality in patients with acromegaly [6]. Chronic excess of GH and IGF-1 lead to hypertension, left ventricular hypertrophy and diastolic dysfunction, which are often the first cardiovascular manifestations of acromegaly [65,66]. Heart failure, arrhythmia and valvular disease may also occur in acromegaly [67,68]. Metabolic comorbidities such as dyslipidaemia, insulin resistance, glucose intolerance or diabetes owing to the lipolytic and insulin-antagonistic effects of GH may also increase the cardiovascular risk [9,15,69]. Diabetes may be identified in 19–28% of patients with acromegaly [70].

### 3.5.2. Cats

Cardiovascular abnormalities in cats with hypersomatotropism include systolic murmur (present in 18% of cats with acromegaly [47]), cardiomegaly, cardiomyopathy and ventricular hypertrophy (Fig. 4d); congestive heart failure may also occur in some acromegalic cats [71]. Many of the metabolic complications seen in cats with hypersomatotropism may be attributed to diabetes rather than to GH/IGF-1 excess. Cats usually present with polyuria/polydipsia and polyphagia. However, polyphagia (and weight gain) independent of glycaemic control and extreme polyphagia (due to stimulation of the hunger centre by GH) are more indicative of hypersomatotropism than diabetes mellitus [34]. Insulin resistance or increased glycaemic variability, in the absence of other causes, is often the only clue pointing towards feline hypersomatotropism in diabetic cats [72]. GH, especially in carnivores, has a powerful diabetogenic activity and appears to provoke hyperglycaemia by inducing peripheral insulin resistance [34]. In one study involving 14 acromegalic cats, after prolonged illness, half of them

developed azotemia and signs of renal disease [54]. Cats in general are prone to developing chronic kidney disease, with some studies reporting a prevalence that can reach close to 70% in geriatric cats [73]. However, diabetic nephropathy is not commonly reported and chronic kidney disease is usually linked to chronic interstitial glomerulonephritis. In feline acromegaly, however, the pathogenesis of kidney disease is not entirely clear. Inflammatory interstitial changes are mild or non-existent in acromegalic cats, and pathological changes have been characterized as diffuse thickening of the mesangium, resembling diabetic glomerulopathy or glomerulosclerosis lesions seen in humans [54].

## 3.6. Local tumour effects

### 3.6.1. Humans

The local growth of a pituitary tumour may be accompanied by severe headaches, which are particularly common in acromegaly [74], as well as visual field defects, and more rarely, cranial nerve palsies, diplopia, facial pain, and/or compression of the hypothalamus or the frontal lobe [9,65,75,76]. Pituitary stalk compression due to large pituitary tumours may contribute to hyperprolactinemia, which can lead to galactorrhoea and hypogonadotropic hypogonadism that can translate clinically as menstrual disturbances in women, and erectile dysfunction and low libido in male patients [6,77]. Hypopituitarism due to the mass effect on the normal pituitary gland may occur in up to 20% of patients [6,9,75].

### 3.6.2. Cats

Pituitary mass growth may have consequences in acromegalic cats such as peripheral blindness or mental dullness, but these might be difficult to recognise in early stages of the disease [34]. In cats with large pituitary masses, overt signs of central nervous system dysfunction may appear, such as seizures, circling, complete blindness, ataxia, tetraparesis and altered mentation [31].

### 3.7. Other clinical features

#### 3.7.1. Humans

Patients with acromegaly are more prone to depression, anxiety, apathy and low self-esteem than healthy subjects [78–82]. Skin problems are also common in patients with acromegaly. Cutaneous thickening, acne, oily skin, hyperhidrosis, hirsutism, skin tags (Fig. 5a), acanthosis nigricans, may be seen in patients with acromegaly, and typically result from the trophic effects of GH/IGF-1 in the subcutaneous tissue and sebaceous glands [83,84]. Other skin lesions, such as cutis verticis gyrata, may also be found in patients with acromegaly (Fig. 5b) [85,86].

#### 3.7.2. Cats

Changes in behaviour and demeanour are difficult to assess in cats, however, central nervous manifestations such as circling, blindness and seizures, may be seen in about 1.7% of acromegalic patients [31]. Lethargy, regardless of origin, may be present in 25% of these cats [47].

Some cats with hypersomatotropism (about 3% [47]) may display a palmigrade/plantigrade stance (Fig. 5c-d). This manifestation is however more common in uncomplicated diabetes, usually secondary to peripheral neuropathy, but it can still be found in non-diabetic cats with hypersomatotropism [34]. In one case study, a non-diabetic acromegalic cat which presented with posterior paraparesis and hypotonia, as well as plantigrade stance, broad facial features and respiratory stridor, was eventually diagnosed with sciatic nerve neuropathy [87].

## 4. Cancer

#### 4.1.1. Humans

An increased incidence of cancer in patients with acromegaly has been suggested, however, this is not well established [88,89]. Recently, a modest increase in this risk has been shown in thyroid, colorectal, anal, central nervous system, gastric, urinary, haematological, pancreatic and small intestine tumours [88,89]. Despite the inconsistent evidence, patients with acromegaly should undergo a screening colonoscopy at diagnosis, considering the increased risk of colonic polyps. Further evaluation should be performed according to the general population, particularly when acromegaly is controlled or in remission [83].

#### 4.1.2. Cats

To our knowledge, there is no data regarding the incidence (or risk) of cancer in cats with acromegaly.

## 5. Diagnosis of acromegaly

#### 5.1. Humans

When acromegaly is suspected, a hormonal work-up should be performed, which can begin with assessing the serum levels of IGF-1 adjusted for age and gender [15,83]. High IGF-1 levels (1.3 times above the upper limit of the normal range), and lack of suppression of GH on an oral glucose tolerance test (GH nadir cutoff of  $<0.4$   $\mu\text{g/L}$  for non-obese patients and  $<0.2$   $\mu\text{g/L}$  for obese patients) confirm the diagnosis of acromegaly [90]. However, IGF-1 and GH levels may be discordant in some situations, and elevated levels of IGF-1 or GH may not always be a synonym of acromegaly [91]. GH response to oral glucose load may also be abnormal in females, obese or older patients [6,15,68].

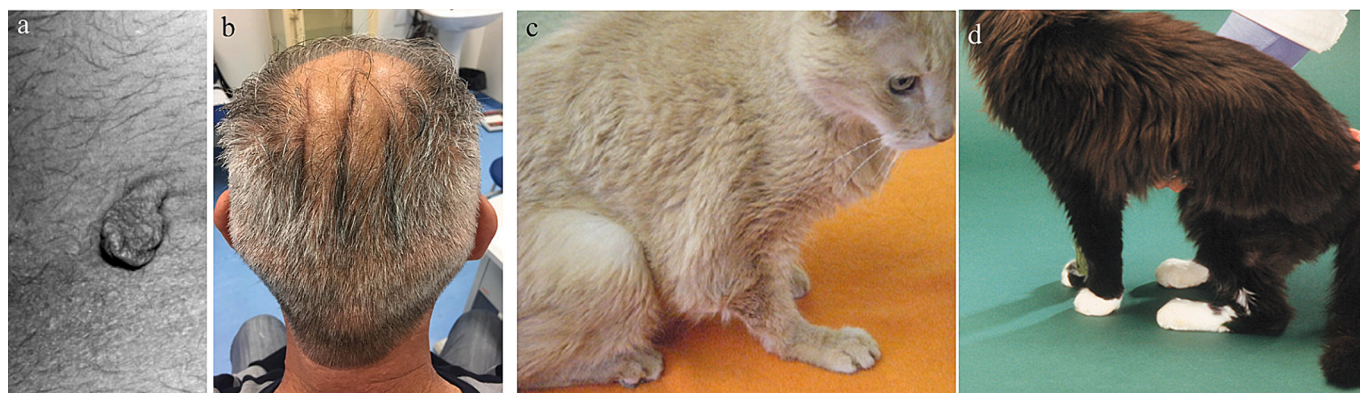
After biochemical confirmation of acromegaly, a pituitary magnetic resonance imaging (MRI) scan should be performed, which allows the detection and characterization of the pituitary tumour (Fig. 6a) [9,15,83]. In the rare cases of ectopic acromegaly, measurement of serum GHRH, functional somatostatin receptor imaging and/or whole-body cross-sectional imaging may be crucial [92,93].

#### 5.2. Cats

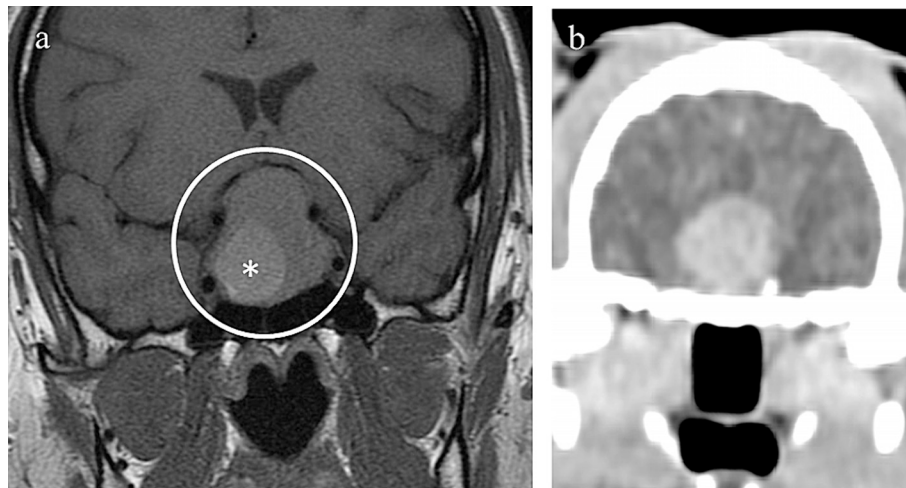
The diagnosis of acromegaly in cats also relies on the measurement of serum IGF-1, which is typically lower than 600 ng/mL. In a study performed on diabetic cats from the United Kingdom, values  $>1000$  ng/mL were shown to have a positive predictive value for the diagnosis of hypersomatotropism of 95% [34]. Values between 600 and 1000 ng/mL are considered abnormal but not diagnostic of hypersomatotropism. In these cases, vigilance regarding blood glucose and IGF-1 concentrations is recommended [34]. In contrast with humans, neither age nor sex appear to influence IGF-1 concentrations in cats [94,95]. It is important to note that, in cats, a diagnosis of acromegaly entails the presence of phenotypical changes. Otherwise, the preferred nomenclature is hypersomatotropism.

GH suppression tests are not routinely performed because there are no commercially available GH assays validated for cats [96]. Moreover, GH suppression test following a glucose load are not useful, since glucose does not inhibit GH secretion in cats [34,97].

Although not routinely performed in daily practice, serum type III procollagen propeptide (PIIP) also appears to have diagnostic value and indicates the presence of active acromegaly. PIIP correlates positively with serum IGF-1, and its concentration increases in the presence of abnormal tissue growth. PIIP is independent of portal insulin availability [34]. Portal insulin is required for hepatic IGF-1 production. In parallel



**Fig. 5.** Other clinical features of acromegaly in humans (a-b) and cats (c-d). Pigmented skin tags (a); convoluted folds and deep furrows of the scalp termed cutis verticis gyrata (b), and, may be found in patients with acromegaly (b). In cats, the most distinctive features from humans are palmigrade and plantigrade stance (c-d). The sources from the pictures are: a) Ben-Shlomo 2006 Clin Dermatol [PMID: 16828406]; b) unpublished original image; c) Romich 2014 ISBN 978-1133125761; d) Feline Medicine Q&A 15 Wikivet.net (accessed 5 Nov 2023).



**Fig. 6.** Radiological appearance of growth hormone-secreting pituitary adenomas in humans (a) and in cats (b). Pituitary magnetic resonance imaging (MRI) scan reveals a large pituitary mass with suprasellar extension and optic chiasm compression (a), with an area corresponding to a probable subacute haemorrhage (\*). Brain CT scan showing a pituitary tumour with a mass extending dorsally and to the right-hand side in an acromegalic cat (b). The sources from the pictures are: a) Weerakkody Y. Pituitary macroadenoma. Reference article, [Radiopaedia.org](https://doi.org/10.53347/rID-9801), <https://doi.org/10.53347/rID-9801> (accessed on 19 Dec 2023); b) Unpublished original image.

with people, endogenous insulin concentrations can be low in feline diabetes mellitus, a low IGF-1 reading could be obtained when sampling takes place prior to initiation of exogenous insulin treatment despite the presence of excess GH due to acromegaly [48]. In order to reduce the occurrence of false negatives, it is important to measure IGF-1 concentrations 2–4 weeks after initiating exogenous insulin therapy [34,98].

Other biochemical abnormalities that may also be found in acromegalic cats include hyperglycaemia (>120 mg/dL or 6.6 mmol/L), hypercholesterolaemia, mild increases in serum alanine aminotransferase and in alkaline phosphatase, hyperproteinemia and elevated serum urea nitrogen [61]. Blood electrolytes may also be abnormal. Mild to moderate hyperphosphataemia without overt renal dysfunction (i.e., azotemia) has been described in almost a third of the reported acromegalic cats [54,61,99]. The most reported haematological change in feline acromegalics is erythrocytosis, which has been attributed to GH or IGF-1 effects on the bone marrow [61]. IGF-1 plays a key role in the proliferation and osteogenic differentiation of bone marrow mesenchymal stem cells, a type of stem cells in the bone marrow tissue with osteogenic differentiation potential [100]. An *in vitro* study has also showed that IGF-1 has a marked positive effect on the proliferative activity of stem cell cultures derived from the cat bone marrow [101]. None of these biochemical abnormalities are pathognomonic of acromegaly or hypersomatotropism and may appear in other endocrinopathies such as uncomplicated diabetes. Their stand-alone diagnostic ability is, therefore, quite limited.

After establishing the biochemical diagnosis of acromegaly in a cat, diagnostic imaging investigations should be performed. Most cats with serum IGF-1 >1000 ng/mL have a large (>4 mm) pituitary neoplasm extending beyond the dorsal rim of the sella turcica (Fig. 6b). MRI is more sensitive than computed tomography (CT) scan, but 95% of acromegalic cats will display abnormalities on the CT scan [34].

## 6. Management of acromegaly

### 6.1. Humans

The main treatment goals are achieving normal GH and IGF-1 serum levels, control tumour growth, prevent or ameliorate systemic comorbidities, improve the quality of life and reduce the mortality risk [102,103]. Acromegaly in humans is managed with transsphenoidal surgery to resect the GH-secreting pituitary tumour, medical therapy and radiotherapy [104]. A tendency for a “personalized” approach

considering patient demographics, and tumour radiological and pathological characterization is emerging, favouring disease control over time [15,83,105].

Current recommendations favour transsphenoidal surgery as the first-line treatment approach for most cases [58,106]. Surgical remission is considered when normal IGF-1 and undetectable GH levels are obtained [9,15], but IGF-1 normalization and cure of acromegaly may be delayed [107]. If the disease persists after surgery or the patient is not suitable for an operation, medical treatment with somatostatin analogues, dopamine agonists or pegvisomant (GH receptor antagonist) may be used [9,15,104]. Biochemical control may be achieved in 30–40% and tumour shrinkage can be observed in two thirds of patients treated with first generation somatostatin analogues; biochemical and structural responses to pasireotide may be found in 54% and 80% of cases [108]. Cabergoline may control IGF-1 levels in around 20% of cases, particularly those with mild elevations of IGF-1, and tumour shrinkage may be obtained in 33% of cases [108]. Pegvisomant is highly effective in blocking the action of GH, normalising IGF-1 levels in 90% of patients in clinical trials, and in about 73% of patients in real-life studies [109–113]. Stereotactic or conventional radiotherapy may also be employed to control the disease, particularly in cases refractory to surgery and medical therapy [114,115].

Acromegaly-related co-morbidities, including diabetes mellitus, hypertension, cardiovascular disease, obstructive sleep apnoea and osteoarthritis, should also be addressed and managed individually [15,116,117]. Acromegaly patients with hypopituitarism at diagnosis due to mass effect, or post-operative/post-radiotherapy hypopituitarism, should initiate adequate hormone replacement therapy [118].

### 6.2. Cats

Hypophysectomy is considered the gold-standard treatment, given its high success rates: insulin may be withdrawn in up to 85% of acromegalic cats with diabetes, and serum IGF-1 levels normalize in up to 90% of cats after surgery [34]. In one case series of 25 acromegalic cats undergoing hypophysectomy, 3 died within 8 weeks of surgery [119]. However, only one death was linked to the procedure. 96% of cats achieved normalization of serum IGF-1 and 92% achieved diabetic remission. Median survival time was 1347 days. Another study reported 15% of postoperative deaths, with 95% of the surviving cats having improved diabetic control and 71% achieving remission. Median survival time was 853 days [120]. Hypophysectomy, however, requires

lifelong glucocorticoid and thyroid supplementation [34]. The follow-up entails the measurement of IGF-1, and remission is considered in case of normal IGF-1 one year post-surgery [34,119]. Cryohypophysectomy, the destruction of the hypophysis through extreme cold, usually a liquid nitrogen cooled cryoprobe, has also been reported in two cats: one with a favorable outcome, with diminished insulin resistance, although with no IGF-1 measurements available; the other maintained peripheral insulin resistance with no significant reduction of plasma IGF-1 [121].

Medical therapy in acromegalic cats is widely unavailable and is associated with only modest success rates. Cats with acromegaly have higher expression of somatostatin receptor (SSTR) type 1 and SSTR type 5, and lower expression of SSTR type 2, which renders the first-generation somatostatin analogues (octreotide and lanreotide) rather ineffective, as they mainly act via SSTR type 2 [35]. Pasireotide inhibits SSTR types 1, 2, 3 and 5, and has shown modest success in treating cats with acromegaly [122]. While dopamine agonists such as cabergoline were initially considered ineffective [123], recent studies [124,125], including one case series of 3 cats that achieved remission following cabergoline treatment [125], have advocated its use as a promising option when other treatments are not available. Cabergoline and pasireotide appear to have similar efficacy regarding normalization of IGF-1 (25% vs 26%) [124,125]. Given that pasireotide has a higher rate of adverse effects, similar efficacy and an often-prohibitive cost for owners, cabergoline could be a treatment option for cats with hypersecretion and diabetes [122,124,125]. One study [124] has shown that, in cats undergoing treatment with cabergoline, median IGF-1 decreased significantly. However, the initial median pituitary height cats that experienced an IGF-1 reduction was significantly lower compared with those that did not experience an IGF-1 reduction [124].

This may indicate cabergoline is more suitable for treatment of “milder” cases of acromegaly, in cats with smaller pituitary tumours.

Conventional radiotherapy may also be an option if hypophysectomy is not available or the tumour is too large. Extent and timing of success are unpredictable, since improvement may take a more than a year. Stereotactic radiotherapy (SRT), however, appears to be more effective. In one study [126], 95% of cats exhibited a decrease in the required insulin dose. Studies regarding conventional radiotherapy show a more variable response, with 55 to 92% of cats being able to reduce insulin dose. The overall median survival time of cats undergoing SRT also appears to be better than those undergoing conventional radiotherapy (1072 vs 508–840 days) [126–130]. Diabetic remission can occur in some acromegalic cats following radiotherapy given as primary treatment (both SRT and conventional radiotherapy), but the efficacy rate is lower than in cats undergoing hypophysectomy. Hypophysectomy has been performed successfully in patients with post-radiation relapse. Radiation may also be a useful neoadjuvant treatment, allowing a pre-surgical reduction of large pituitary tumours in selected cases [34].

In countries where hypophysectomy or radiotherapy are not available, the treatment of acromegaly is purely symptomatic: exogenous insulin for diabetes, medication for hypertrophic cardiomyopathy, drugs for hypertension, and analgesia for joint pain [34,131].

## 7. Conclusions

Acromegaly in humans and cats share some key similarities summarised in Table 1. The metabolic effects of excess GH and IGF-1 in both species lead to complications such as hypertension, left ventricular hypertrophy, diastolic dysfunction, diabetes mellitus, arthropathy and

**Table 1**  
Epidemiology, pathophysiology and clinical features of acromegaly in humans and cats.

		Humans	Cats
General features	Epidemiology	3.6–13 cases per 100,000 individuals More common in older women	125 cases per 100,000 individuals (1 in 800 cats) More common in older neutered male cats (70%)
	Pathophysiology	GH-secreting pituitary tumour (>95%) – May be plurihormonal (up to 30% also secrete prolactin) Genetic causes identified (e.g. <i>AIP</i> , <i>GNAS</i> and <i>GPR101</i> mutations) Environmental causes possible (pollution)	GH-secreting pituitary tumour – Very rarely plurihormonal. Prolactin secretion has not been documented No strong evidence for genetic causes Environmental causes possible (pollution)
Clinical features	Facial features	Frontal bossing Prognathism Dental malocclusion	Broad facial features (37%) Diastema Prognathism (18%) Dental malocclusion
	Cardiovascular and metabolic	Hypertension (17–51%) Left ventricular hypertrophy Diastolic dysfunction Heart failure	Heart murmur (18%) Hypertension Left ventricular hypertrophy Diabetes (15–25%) Azotemia and kidney disease
	Skeletal	Joint stiffness/pain (70%) Carpal tunnel syndrome (25–51%) Increased joint cartilage thickness Calvaria thickening	Joint stiffness/pain (10%) <i>Spondylosis deformans</i> Increased joint cartilage thickness Calvaria thickening Enlarged extremities/clubbed paws (13%)
	Organomegaly	Thyroid enlargement/goiter Salivary glands Hepatomegaly Splenomegaly	Hepatomegaly Splenomegaly Nephromegaly Abdominal enlargement (40%)
	Local tumour effects	Headaches (32–87%) Visual field defects (4–62%) Cranial nerve palsies	Cardiomegaly CNS signs excluding lethargy (1.7%) Lethargy (25%)
	Other notable features	Respiratory stridor/snoring Sleep apnoea (7–30%) Hirsutism	Respiratory stridor/snoring (38%) Palmigrade/plantigrade stance (3%)
	Cancer	Modest increase in risk in certain cancers: Thyroid Gastrointestinal tract	Not consistently reported Urinary tract Haematological Pancreatic

*AIP*: Aryl hydrocarbon receptor interacting protein; *CNS*: Central nervous system; *GNAS*: Guanine nucleotide binding protein alpha stimulating activity polypeptide gene; *GPR101*: G-protein coupled receptor gene; *HCM*: hypertrophic cardiomyopathy.

nephropathy. Physical attributes are also similar in affected individuals, including enlarged/club-like extremities, broad facial features, dental diastema and inferior prognathia.

Some signs in humans, such as galactorrhoea, menstrual disturbances, erectile dysfunction and low libido are not observed in cats since most are spayed or neutered. Cats also do not usually have specific dermatological signs associated with acromegaly, while in humans skin manifestations, such as skin tags, oily skin or acne, are common. Unkempt coat can sometimes be observed but it is a universal sign of illness in cats. In affected cats, the typical acromegaloid phenotype is not always present. Cats can have very high serum IGF-1 concentrations and a normal appearance. Often, the only sign is insulin-resistant or increased glycaemia /diabetes in cats with diabetes mellitus. While not all acromegalic cats are diabetic, the overwhelming majority is.

Some studies suggest that the prevalence of acromegaly in humans may be vastly underestimated [102,103], since, like in cats, some patients with acromegaly may have very mild acromegalic features [132].

Given the overall similarities between the presentation of acromegaly in both species and the shorter lifespan of cats, feline acromegaly may be a useful model for studying this disease in human patients. On the other hand, since data regarding acromegaly in humans is significantly vaster, it may provide some insights and further knowledge about the disease in other species, including cats, especially given the same pathophysiological mechanisms underlying to the excess of GH and IGF-1, and the several similarities, as illustrated here.

#### Formatting of funding sources

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

#### CRediT authorship contribution statement

**Mariana Lopes-Pinto:** Writing – review & editing, Writing – original draft, Methodology, Investigation, Formal analysis, Data curation, Conceptualization. **Patrícia Lunet Marques:** Writing – review & editing, Writing – original draft, Methodology, Investigation, Data curation, Conceptualization. **Emilia Lacerda-Nobre:** Writing – review & editing, Validation, Resources. **Diego Miceli:** Writing – review & editing, Validation, Resources. **Rodolfo Oliveira Leal:** Writing – review & editing, Validation, Supervision, Resources, Methodology, Conceptualization. **Pedro Marques:** Writing – review & editing, Validation, Supervision, Resources, Methodology, Conceptualization.

#### Declaration of competing interest

None.

#### Acknowledgements

N/A

#### References

- [1] W.W. de Herder, Acromegaly and gigantism in the medical literature. Case descriptions in the era before and the early years after the initial publication of Pierre Marie (1886), *Pituitary* 12 (3) (Sep. 2009) 236–244.
- [2] C. Gemhardt, H. Loppnow, Pathogenesis of spontaneous diabetes mellitus in the cat. II. Acidophilic adenoma of the pituitary gland and diabetes mellitus in 2 cases, *Berl. Munch. Tierarztl. Wochenschr.* 89 (17) (Sep. 1976) 336–340.
- [3] S.J.M. Niessen, Update on feline acromegaly, *In Pract.* 35 (1) (Jan. 2013) 2–6.
- [4] Z. Laron, Insulin-like growth factor 1 (IGF-1): a growth hormone, *Mol. Pathol.* 54 (5) (Oct. 2001) 311–316.
- [5] I.M. Holdaway, R.C. Rajasoorya, G.D. Gamble, Factors influencing mortality in acromegaly, *J. Clin. Endocrinol. Metab.* 89 (2) (Feb. 2004) 667–674.
- [6] S. Melmed, Acromegaly, in: *The Pituitary*, 5th ed., Elsevier, 2022, pp. 449–493.
- [7] A. Lavrentaki, A. Paluzzi, J.A.H. Wass, N. Karavitaki, Epidemiology of acromegaly: review of population studies, *Pituitary* 20 (1) (Feb. 2017) 4–9.
- [8] G.T. Hoskuldottir, S.B. Fjalldal, H.A. Sigurjonsdottir, The incidence and prevalence of acromegaly, a nationwide study from 1955 through 2013, *Pituitary* 18 (6) (Dec. 2015) 803–807.
- [9] A. Colao, et al., Acromegaly, in: *Nature Reviews Disease Primers* Vol. 5, Nature Publishing Group, Dec. 01, 2019 no. 1.
- [10] T.T. Agustsson, et al., The epidemiology of pituitary adenomas in Iceland, 1955–2012: a nationwide population-based study, *Eur. J. Endocrinol.* 173 (5) (Nov. 2015) 655–664.
- [11] J. Dal, et al., Sex differences in acromegaly at diagnosis: a nationwide cohort study and meta-analysis of the literature, *Clin. Endocrinol.* 94 (4) (Apr. 2021) 625–635.
- [12] T. Burton, E. Le Nestour, M. Neary, Incidence and prevalence of acromegaly in a large US health plan database, *Pituitary* 19 (2016) 262–267.
- [13] N.F. Lenders, A.I. McCormack, K.K.Y. Ho, MANAGEMENT OF ENDOCRINE DISEASE: does gender matter in the management of acromegaly? *Eur. J. Endocrinol.* 182 (5) (May 2020) R67–R82.
- [14] A. Tjörnstrand, et al., The incidence rate of pituitary adenomas in western Sweden for the period 2001–2011, *Eur. J. Endocrinol.* 171 (4) (Oct. 2014) 519–526.
- [15] L. Katznelson et al., “Acromegaly: an endocrine society clinical practice guideline,” *J. Clin. Endocrinol. Metab.*, vol. 99, no. 11. Endocrine Society, pp. 3933–3951, Nov. 01, 2014.
- [16] A. Boguslawska, M. Korbonits, Genetics of acromegaly and gigantism, *J. Clin. Med.* 10 (7) (Mar. 2021) 1377.
- [17] P. Marques, M. Korbonits, Genetic aspects of pituitary adenomas, *Endocrinol. Metab. Clin. N. Am.* 46 (2) (Jun. 2017) 335–374.
- [18] S. Cannavò, et al., Increased prevalence of acromegaly in a highly polluted area, *Eur. J. Endocrinol.* 163 (4) (Oct. 2010) 509–513.
- [19] P. Petrossians, et al., Acromegaly at diagnosis in 3173 patients from the Liège acromegaly survey (LAS) database, *Endocr. Relat. Cancer* 24 (10) (Oct. 2017) 505–518.
- [20] I. Potorac, et al., Pituitary MRI features in acromegaly resulting from ectopic GHRH secretion from a neuroendocrine tumor: analysis of 30 cases, *J. Clin. Endocrinol. Metab.* 107 (8) (Jul. 2022) e3313–e3320.
- [21] L. Garby, et al., Clinical characteristics and outcome of acromegaly induced by ectopic secretion of growth hormone-releasing hormone (GHRH): a French Nationwide series of 21 cases, *J. Clin. Endocrinol. Metab.* 97 (6) (Jun. 2012) 2093–2104.
- [22] P. Marques, et al., Significant benefits of AIP testing and clinical screening in familial isolated and young-onset pituitary tumors, *J. Clin. Endocrinol. Metab.* 105 (6) (Jun. 2020) e2247–e2260.
- [23] O. Vierimaa, et al., Pituitary adenoma predisposition caused by germline mutations in the AIP gene, *Science* (1979) 312 (5777) (May 2006) 1228–1230.
- [24] S. Melmed, C. Ezrin, K. Kovacs, R.S. Goodman, L.A. Frohman, Acromegaly due to secretion of growth hormone by an ectopic pancreatic islet-cell tumor, *N. Engl. J. Med.* 312 (1) (Jan. 1985) 9–17.
- [25] L. Rostomyan, A. Beckers, Screening for genetic causes of growth hormone hypersecretion, *Growth Hormon. IGF Res.* 30–31 (Oct. 2016) 52–57.
- [26] L. Cazabat, et al., Germline AIP mutations in apparently sporadic pituitary adenomas: prevalence in a prospective single-center cohort of 443 patients, *J. Clin. Endocrinol. Metab.* 97 (4) (Apr. 2012) E663–E670.
- [27] F. Caimari, M. Korbonits, Novel genetic causes of pituitary adenomas, *Clin. Cancer Res.* 22 (20) (Oct. 2016) 5030–5042.
- [28] S.J.M. Niessen, et al., Feline acromegaly: an underdiagnosed Endocrinopathy? *J. Vet. Intern. Med.* 21 (5) (Sep. 2007) 899–905.
- [29] D.A. Elliott, E.C. Feldman, P.D. Koblik, V.F. Samii, R.W. Nelson, Prevalence of pituitary tumors among diabetic cats with insulin resistance, *J. Am. Vet. Med. Assoc.* 216 (11) (Jun. 2000) 1765–1768.
- [30] D.D. Miceli, et al., Prevalence of hypersomatotropism and hyperthyroidism in cats with diabetes mellitus from referral centers in Buenos Aires (2020–2022), *J. Feline Med. Surg.* 25 (2) (Feb. 2023), p. 1098612X2211485.
- [31] J.M. Fletcher, et al., Hypersomatotropism in 3 cats without concurrent diabetes mellitus, *J. Vet. Intern. Med.* 30 (4) (Jul. 2016) 1216–1221.
- [32] Research Communications of the 33<sup>rd</sup> Ecvim-Ca Congress, *J. Vet. Intern. Med.* 38 (1) (Dec. 2023) 730–863.
- [33] D.G. O'Neill, D.B. Church, P.D. McGreevy, P.C. Thomson, D.C. Brodbelt, Longevity and mortality of cats attending primary care veterinary practices in England, *J. Feline Med. Surg.* 17 (2) (Feb. 2015) 125–133.
- [34] E.C. Feldman, F. Fracassi, M.E. Peterson, *Feline Endocrinology*, 1st ed., Edra S.p.A, Milan, 2019.
- [35] C.J. Scudder, et al., Pituitary pathology and gene expression in Acromegalic cats, *J. Endocr. Soc.* 3 (1) (Jan. 2019) 181–200.
- [36] K. Sanders, S. Galac, B.P. Meij, Pituitary tumour types in dogs and cats, *Vet. J.* 270 (Apr. 2021) 105623.
- [37] S.J. Ettinger, E.C. Feldman, *Textbook of Veterinary Internal Medicine Diseases of The Dog and The Cat* Eighth Edition, 2016.
- [38] B.P. Meij, R.H. van der Vlugt-Meijer, T.S.G.A.M. van den Ingh, A. Rijnberk, Somatotroph and Corticotroph pituitary adenoma (double adenoma) in a cat with diabetes mellitus and hyperadrenocorticism, *J. Comp. Pathol.* 130 (2–3) (Feb. 2004) 209–215.
- [39] M. Sharman, L. FitzGerald, M. Kiupel, Concurrent somatotroph and plurihormonal pituitary adenomas in a cat, *J. Feline Med. Surg.* 15 (10) (Oct. 2013) 945–952.
- [40] A.C. Dirtu, S.J.M. Niessen, P.G. Jorens, A. Covaci, Organohalogenated contaminants in domestic cats' plasma in relation to spontaneous acromegaly and

- type 2 diabetes mellitus: a clue for endocrine disruption in humans? *Environ. Int.* 57–58 (Jul. 2013) 60–67.
- [41] C.J. Scudder, S.J. Niessen, B. Catchpole, R.C. Fowkes, D.B. Church, Y. Forcada, Feline hypersomatotropism and acromegaly tumorigenesis: a potential role for the AIP gene, *Domest. Anim. Endocrinol.* 59 (Apr. 2017) 134–139.
- [42] P. Roccabianca, et al., Multiple endocrine neoplasia type-I-like syndrome in two cats, *Vet. Pathol.* 43 (3) (May 2006) 345–352.
- [43] B. Kipperman, R. Nelson, S. Griffey, E. Feldman, Diabetes mellitus and exocrine pancreatic neoplasia in two cats with hyperadrenocorticism, *J. Am. Anim. Hosp. Assoc.* 28 (5) (1992) 415–418.
- [44] P. Attal, P. Chanson, Endocrine aspects of obstructive sleep apnea, *J. Clin. Endocrinol. Metab.* 95 (2) (Feb. 2010) 483–495.
- [45] R.R. Grunstein, K.Y. Ho, M. Berthon-Jones, D. Stewart, C.E. Sullivan, Central sleep apnea is associated with increased ventilatory response to carbon dioxide and hypersecretion of growth hormone in patients with acromegaly, *Am. J. Respir. Crit. Care Med.* 150 (2) (Aug. 1994) 496–502.
- [46] G. Mazzotti, A.G.A. Lania, E. Canalis, MANAGEMENT OF ENDOCRINE DISEASE: bone disorders associated with acromegaly: mechanisms and treatment, *Eur. J. Endocrinol.* 181 (2) (Aug. 2019) R45–R56.
- [47] E. Côté, S.J. Ettinger, E.C. Feldman, *Ettinger's Textbook of Veterinary Internal Medicine* vol. 2, 2024.
- [48] S.J.M. Niessen, et al., Studying cat (*Felis catus*) diabetes: beware of the acromegalic imposter, *PLoS One* 10 (5) (May 2015) p. e0127794.
- [49] S. Schaefer, et al., Evaluation of insulin-like growth factor-1, total thyroxine, feline pancreas-specific lipase and urinary corticoid-to-creatinine ratio in cats with diabetes mellitus in Switzerland and the Netherlands, *J. Feline Med. Surg.* 19 (8) (Aug. 2017) 888–896.
- [50] C.R. Lamb, et al., Computed tomographic signs of acromegaly in 68 diabetic cats with hypersomatotropism, *J. Feline Med. Surg.* 16 (2) (Feb. 2014) 99–108.
- [51] R. Castellanos-Bueno, et al., Clinical and epidemiological characteristics, morbidity and treatment based on the registry of acromegalic patients in Colombia: RAPACO, *Growth Hormon. IGF Res.* 60–61 (Oct. 2021) 101425.
- [52] P. Marques, M. Korbonits, Approach to the patient with Pseudoacromegaly, *J. Clin. Endocrinol. Metab.* 107 (6) (May 2022) 1767–1788.
- [53] P. Marques, M. Korbonits, Pseudoacromegaly, *Front. Neuroendocrinol.* 52 (Jan. 2019) 113–143.
- [54] M.E. Peterson, et al., Acromegaly in 14 Cats, 1990.
- [55] A. Skandhan, P. Desai, Acromegaly, in: *Radiopaedia.org*, Radiopaedia.org, 2009.
- [56] K. Vouzourneraki, et al., Carpal tunnel syndrome in acromegaly: a nationwide study, *Eur. J. Endocrinol.* 184 (2) (Feb. 2021) 209–216.
- [57] S. Melmed, Acromegaly, in: *The Pituitary*, Elsevier, 2022, pp. 449–493.
- [58] A. Colao, et al., Acromegaly, *Nat. Rev. Dis. Primers.* 5 (1) (Mar. 2019) 20.
- [59] E.M. Whitehead, S.M. Shalet, D. Davies, B.A. Enoch, D.A. Price, C.G. Beardwell, Pituitary gigantism: a disabling condition, *Clin. Endocrinol.* 17 (3) (Sep. 1982) 271–277.
- [60] O.J. Ogedegbe, et al., A comprehensive review of four clinical practice guidelines of acromegaly, *Cureus* 14 (9) (Sep. 2022) e28722.
- [61] C.A. Hurty, B. Flatland, Feline acromegaly: a review of the syndrome, *J. Am. Anim. Hosp. Assoc.* 41 (5) (Sep. 2005) 292–297.
- [62] C. Capatina, J.A.H. Wass, 60 years of neuroendocrinology: acromegaly, *J. Endocrinol.* 226 (2) (Aug. 2015) T141–T160.
- [63] O. Hancerliogullari, R. Senocak, H. Sinan, An uncommon cause of acute abdomen in an acromegalic patient: colonic volvulus, *Ann. Ital. Chir.* (89) (2018) 572576.
- [64] B.N. Lourenço, E. Randall, G. Seiler, K.F. Lunn, Abdominal ultrasonographic findings in acromegalic cats, *J. Feline Med. Surg.* 17 (8) (Aug. 2015) 698–703.
- [65] D.R. Clemmons, et al., Optimizing control of acromegaly: integrating a growth hormone receptor antagonist into the treatment algorithm, *J. Clin. Endocrinol. Metab.* 88 (10) (Oct. 2003) 4759–4767.
- [66] A.F. Casini, L.V. Neto, R. Fontes, R.F. França, S.S. Xavier, M.R. Gadelha, Aortic root ectasia in patients with acromegaly: experience at a single center, *Clin. Endocrinol.* 75 (4) (Oct. 2011) 495–500.
- [67] C. Berg, et al., Cardiovascular risk factors in patients with uncontrolled and long-term acromegaly: comparison with matched data from the general population and the effect of disease control, *J. Clin. Endocrinol. Metab.* 95 (8) (Aug. 2010) 3648–3656.
- [68] S. Melmed, Acromegaly pathogenesis and treatment, *J. Clin. Investig.* 119 (11) (Nov. 2009) 3189–3202.
- [69] G. Vila, J.O.L. Jørgensen, A. Luger, G.K. Stalla, Insulin resistance in patients with acromegaly, *Front. Endocrinol. (Lausanne)* 10 (Jul. 2019).
- [70] K. Borgeat, et al., Time spent with cats is never wasted: lessons learned from feline acromegalic cardiomyopathy, a naturally occurring animal model of the human disease, *PLoS One* 13 (3) (Mar. 2018) p. e0194342.
- [71] A.N. Sharma, M. Tan, E.A. Amsterdam, G.D. Singh, Acromegalic cardiomyopathy: epidemiology, diagnosis, and management, *Clin. Cardiol.* 41 (3) (Mar. 2018) 419–425.
- [72] T. Komiya, et al., Relationship between anti-insulin antibody production and severe insulin resistance in a diabetic cat, *J. Vet. Med. Sci.* 83 (4) (2021) 661–665.
- [73] C.L. Marino, B.D.X. Lascelles, S.L. Vaden, M.E. Gruen, S.L. Marks, Prevalence and classification of chronic kidney disease in cats randomly selected from four age groups and in cats recruited for degenerative joint disease studies, *J. Feline Med. Surg.* 16 (6) (Jun. 2014) 465–472.
- [74] S. Kaniuka-Jakubowska, et al., A study of acromegaly-associated headache with somatostatin analgesia, *Endocr. Relat. Cancer* 30 (3) (Dec. 2023).
- [75] M.R. Drange, N.R. Fram, V. Herman-Bonert, S. Melmed, Pituitary tumor registry: a novel clinical resource <sup>1</sup>, *J. Clin. Endocrinol. Metab.* 85 (1) (Jan. 2000) 168–174.
- [76] P. Caron, et al., Signs and symptoms of acromegaly at diagnosis: the physician's and the patient's perspectives in the ACRO-POLIS study, *Endocrine* 63 (1) (Jan. 2019) 120–129.
- [77] K. Kovacs, E. Horvath, S.L. Asa, L. Stefanescu, T. Sano, Pituitary cells producing more than one hormone human pituitary adenomas, *Trends Endocrinol. Metab.* 1 (2) (Nov. 1989) 104–107.
- [78] M.R. Gadelha, L. Kasuki, D.S.T. Lim, M. Flesteriu, Systemic complications of acromegaly and the impact of the current treatment landscape: an update, *Endocr. Rev.* 40 (1) (Feb. 2019) 268–332.
- [79] E.B. Geer, et al., Observed discordance between outcomes reported by acromegaly patients and their treating endocrinology medical provider, *Pituitary* 23 (2) (Apr. 2020) 140–148.
- [80] G.A. Fava, N. Sonino, M.A. Morphy, Psychosomatic view of endocrine disorders, *Psychother. Psychosom.* 59 (1) (1993) 20–33.
- [81] P. Pantanetti, N. Sonino, G. Arnaldi, M. Boscaro, Self image and quality of life in acromegaly, *Pituitary* 5 (1) (2002) 17–19.
- [82] M. Flesteriu, et al., A pituitary society update to acromegaly management guidelines, *Pituitary* 24 (1) (Feb. 2021) 1–13.
- [83] A. Ben-Shlomo, S. Melmed, Skin manifestations in acromegaly, *Clin. Dermatol.* 24 (4) (Jul. 2006) 256–259.
- [84] J. Leavitt, Skin tags: a cutaneous marker for colonic polyps, *Ann. Intern. Med.* 98 (6) (Jun. 1983) 928.
- [85] J. Roque, P. Marques, Acromegaly-related cutis verticis gyrata, *Growth Hormon. IGF Res.* 66 (Oct. 2022) 101497.
- [86] J. Roque, P. Marques, Clinico-radiological features of acromegaly-related cutis verticis gyrata, *Hormones* 21 (3) (Sep. 2022) 519–520.
- [87] A. Corsini, E. Bianchi, A. Volta, M. Bonazzi, Sciatic neuropathy in an acromegalic cat without concurrent diabetes mellitus, *J. Feline Med. Surg. Open Rep.* 6 (1) (Jan. 2020), 205511692090693.
- [88] D. Esposito, O. Ragnarsson, G. Johannsson, D.S. Olsson, Incidence of benign and malignant tumors in patients with acromegaly is increased: a nationwide population-based study, *J. Clin. Endocrinol. Metab.* 106 (12) (Aug. 2021) 3487–3496.
- [89] Z. Xiao, P. Xiao, Y. Wang, C. Fang, Y. Li, Risk of cancer in acromegaly patients: an updated meta-analysis and systematic review, *PLoS One* 18 (11) (Nov. 2023) e0285335.
- [90] A. Giustina, et al., Consensus on criteria for acromegaly diagnosis and remission, *Pituitary* 27 (1) (Feb. 2024) 7–22.
- [91] C. Peixe, M. Sánchez-García, A.B. Grossman, M. Korbonits, P. Marques, Biochemical discrepancies in the evaluation of the somatotroph axis: elevated GH or IGF-1 levels do not always diagnose acromegaly, *Growth Hormon. IGF Res.* 64 (Jun. 2022) 101467.
- [92] S. Daud, A.H. Hamrahian, R.J. Weil, M. Hamaty, R.A. Prayson, L. Olansky, Acromegaly with negative pituitary MRI and no evidence of ectopic source: the role of transphenoidal pituitary exploration? *Pituitary* 14 (4) (Dec. 2011) 414–417.
- [93] F. Borson-Chazot, L. Garby, G. Raverot, F. Claustrat, V. Raverot, G. Sassolas, Acromegaly induced by ectopic secretion of GHRH: a review 30 years after GHRH discovery, *Ann. Endocrinol. (Paris)* 73 (6) (Dec. 2012) 497–502.
- [94] C.E. Reusch, S. Kley, M. Casella, R.W. Nelson, J. Mol, J. Zapf, Measurements of growth hormone and insulin-like growth factor 1 in cats with diabetes mellitus, *Vet. Rec.* 158 (6) (Feb. 2006) 195–200.
- [95] A. Maxwell, R. Butterwick, R.M. Batt, C. Camacho-Hübner, Serum insulin-like growth factor (IGF)-I concentrations are reduced by short-term dietary restriction and restored by refeeding in domestic cats (*Felis catus*), *J. Nutr.* 129 (10) (Oct. 1999) 1879–1884.
- [96] M. Rosca, Y. Forcada, G. Solcan, D.B. Church, S.J. Niessen, Screening diabetic cats for hypersomatotropism: performance of an enzyme-linked immunosorbent assay for insulin-like growth factor 1, *J. Feline Med. Surg.* 16 (2) (Feb. 2014) 82–88.
- [97] N. Kokka, J.F. Garcia, M. Morgan, R. George, Immunoassay of plasma growth hormone in cats following fasting and administration of insulin, arginine, 2-Deoxyglucose and hypothalamic extract, *Endocrinology* 88 (2) (Feb. 1971) 359–366.
- [98] E.M. Strage, et al., Effect of insulin treatment on circulating insulin-like growth factor I and IGF-binding proteins in cats with diabetes mellitus, *J. Vet. Intern. Med.* 32 (5) (Sep. 2018) 1579–1590.
- [99] E.C. Feldman, R.W. Nelson, Acromegaly and hyperadrenocorticism in cats: a clinical perspective, *J. Feline Med. Surg.* 2 (3) (Sep. 2000) 153–158.
- [100] J. Feng, Z. Meng, Insulin growth factor-1 promotes the proliferation and osteogenic differentiation of bone marrow mesenchymal stem cells through the Wnt/ $\beta$ -catenin pathway, *Exp. Ther. Med.* 22 (2) (Jun. 2021) 891.
- [101] A. Mazurkevych, et al., The effect of a fibroblast growth factor, insulin-like growth factor, growth hormone, and Biolaminin 521 LN on the proliferative activity of cat stem cells, *Acta Vet. Brno* 90 (1) (2021) 77–85.
- [102] A. Giustina, et al., Multidisciplinary management of acromegaly: a consensus, *Rev. Endocr. Metab. Disord.* 21 (4) (Dec. 2020) 667–678.
- [103] A. Giustina, et al., A consensus on the diagnosis and treatment of acromegaly comorbidities: an update, *J. Clin. Endocrinol. Metab.* 105 (4) (Apr. 2020) e937–e946.
- [104] R. Pivonello, et al., Complications of acromegaly: cardiovascular, respiratory and metabolic comorbidities, *Pituitary* 20 (1) (Feb. 2017) 46–62.
- [105] S. Melmed, Pituitary medicine from discovery to patient-focused outcomes, *J. Clin. Endocrinol. Metab.* 101 (3) (Mar. 2016) 769–777.
- [106] M.-S. Shin, et al., Long-term changes in serum IGF-1 levels after successful surgical treatment of growth hormone-secreting pituitary adenoma, *Neurosurgery* 73 (3) (Sep. 2013) 473–479.

- [107] B. Maia, L. Kasuki, M.R. Gadelha, Novel therapies for acromegaly, *Endocr. Connect.* 9 (12) (Dec. 2020) R274–R285.
- [108] A.J. van der Lely, et al., Long-term treatment of acromegaly with pegvisomant, a growth hormone receptor antagonist, *Lancet* 358 (9295) (Nov. 2001) 1754–1759.
- [109] T. Feola, et al., Pegvisomant improves glucose metabolism in acromegaly: a Meta-analysis of prospective interventional studies, *J. Clin. Endocrinol. Metab.* 104 (7) (Jul. 2019) 2892–2902.
- [110] P.J. Trainer, et al., Treatment of acromegaly with the growth hormone–receptor antagonist pegvisomant, *N. Engl. J. Med.* 342 (16) (Apr. 2000) 1171–1177.
- [111] L. Katznelson, Pegvisomant for the treatment of acromegaly—translation of clinical trials into clinical practice, *Nat. Clin. Pract. Endocrinol. Metab.* 3 (7) (Jul. 2007) 514–515.
- [112] T. Brue, et al., Diabetes in patients with acromegaly treated with pegvisomant: observations from arostudy, *Endocrine* 63 (3) (Mar. 2019) 563–572.
- [113] A.M. Abu Dabrh, et al., Radiotherapy versus radiosurgery in treating patients with acromegaly: a systematic review and meta-analysis, *Endocr. Pract.* 21 (8) (Aug. 2015) 943–956.
- [114] M.J. Hannon, A.L. Barkan, W.M. Drake, The role of radiotherapy in acromegaly, *Neuroendocrinology* 103 (1) (2016) 42–49.
- [115] M. Sherlock, et al., Mortality in patients with pituitary disease, *Endocr. Rev.* 31 (3) (Jun. 2010) 301–342.
- [116] I.M. Holdaway, M.J. Bolland, G.D. Gamble, A meta-analysis of the effect of lowering serum levels of GH and IGF-1 on mortality in acromegaly, *Eur. J. Endocrinol.* 159 (2) (Aug. 2008) 89–95.
- [117] M. Flseriu, et al., Hormonal replacement in hypopituitarism in adults: an Endocrine Society clinical practice guideline, *J. Clin. Endocrinol. Metab.* 101 (11) (Nov. 2016) 3888–3921.
- [118] K.L. van Bokhorst, et al., Evaluation of hypophysectomy for treatment of hypersomatotropism in 25 cats, *J. Vet. Intern. Med.* 35 (2) (Mar. 2021) 834–842.
- [119] J. Fenn, et al., Efficacy of hypophysectomy for the treatment of hypersomatotropism-induced diabetes mellitus in 68 cats, *J. Vet. Intern. Med.* 35 (2) (Mar. 2021) 823–833.
- [120] S.L. Blois, D.L. Holmberg, Cryohypophysectomy used in the treatment of a case of feline acromegaly, *J. Small Anim. Pract.* 49 (11) (Aug. 2008) 596–600.
- [121] R. Gostelow, et al., Pasireotide long-acting release treatment for diabetic cats with underlying Hypersomatotropism, *J. Vet. Intern. Med.* 31 (2) (Mar. 2017) 355–364.
- [122] L. Abraham, S. Helmond, R. Mitten, J. Charles, S. Holloway, Treatment of an acromegalic cat with the dopamine agonist L-deprenyl, *Aust. Vet. J.* 80 (8) (Aug. 2002) 479–483.
- [123] D.D. Miceli, et al., Cabergoline treatment in cats with diabetes mellitus and hypersomatotropism, *J. Feline Med. Surg.* 24 (12) (Dec. 2022) 1238–1244.
- [124] D.D. Miceli, P.N. Vidal, G.A. Pompili, V.A. Castillo, E.A. Soler Arias, S.J. Niessen, Diabetes mellitus remission in three cats with hypersomatotropism after cabergoline treatment, *J. Feline Med. Surg. Open Rep.* 7 (1) (Jan. 2021), p. 205511692110189.
- [125] T.L. Wormhoudt, et al., Stereotactic radiation therapy for the treatment of functional pituitary adenomas associated with feline acromegaly, *J. Vet. Intern. Med.* 32 (4) (Jul. 2018) 1383–1391.
- [126] R.K. Sellon, J. Fidel, R. Houston, P.R. Gavin, Linear-accelerator-based modified Radiosurgical treatment of pituitary tumors in cats: 11 cases (1997–2008), *J. Vet. Intern. Med.* 23 (5) (Sep. 2009) 1038–1044.
- [127] M.N. Mayer, D.S. Greco, S.M. LaRue, Outcomes of pituitary tumor irradiation in cats, *J. Vet. Intern. Med.* 20 (5) (Sep. 2006) 1151–1154.
- [128] M.D. Dunning, C.S. Lowrie, N.H. Bexfield, J.M. Dobson, M.E. Herrtage, Exogenous insulin treatment after hypofractionated radiotherapy in cats with diabetes mellitus and acromegaly, *J. Vet. Intern. Med.* 23 (2) (Mar. 2009) 243–249.
- [129] M.J. Brearley, G.A. Polton, R.M. Littler, S.J.M. Niessen, Coarse fractionated radiation therapy for pituitary tumours in cats: a retrospective study of 12 cases, *Vet. Comp. Oncol.* 4 (4) (Dec. 2006) 209–217.
- [130] A. Corsini, et al., Quality of life and response to treatment in cats with hypersomatotropism: the owners' point of view, *J. Feline Med. Surg.* 24 (8) (Aug. 2022) e175–e182.
- [131] P.W. Rosario, M.R. Calsolari, Screening for acromegaly by application of a simple questionnaire evaluating the enlargement of extremities in adult patients seen at primary health care units, *Pituitary* 15 (2) (Jun. 2012) 179–183.
- [132] P.W. Rosario, Frequency of acromegaly in adults with diabetes or glucose intolerance and estimated prevalence in the general population, *Pituitary* 14 (3) (Sep. 2011) 217–221.